TAXONOMIC LISTING OF PACIFIC NORTHWEST GENERA

* - indicates definite records from the Pacific Northwest

	OD A TITAL		20 /0 Fine 02 04
PALAEOSOMATA (=BIFEMO	ORATINA)	*Maerkelotritia	39-40, figs. 83-84
Archeonothroidea		(=Oribotritia se	
	2, fig. 64	*Mesotritia	40
	2, fig. 63	*Microtritia	40-41, fig. 85
Palaeacaroidea	0 fig 61	*Oribotritia	39, figs. 81-82
	2, fig. 61	(=Plesiotritia)	10
Ctenacaroidea		*Rhysotritia	40
	2, fig. 59	*Synichotritia	41
	2, fig. 62	Perlohmannioidea	(5. fina 164-166 100
	2, fig. 60	*Perlohmannia	65, figs. 164-166, 188
ENARTHRONOTA (=ARTHRO	UNUTINA)	Epilohmannioidea	(E ((Fige 167-160
Brachychthonioidea	0 20 fig 52	*Epilohmannia	65-66, figs. 167-169,
	9-30, fig. 53	P 1 1	187
*Eobrachychthonius 29		Eulohmannioidea	25 figo 67-69
	9, figs. 54, 55, 306		35, figs. 67-68
Mixochthonius 29	-	DESMONOMATA	
Neobrachychthonius29		Crotonioidea (=Noth	
Neoliochthonius 29		*Camisia	36, 68, figs. 70-71,
(=Paraliochthonius		II and the attraction of	73, 177-178, 308
Poecilochthonius 29		Heminothrus	71 26 fig 74
*Sellnickochthonius		*Malaconothrus Mucronothrus	36, fig. 74 36
(=Brachychochthoni	-		
*Synchthonius 29 Verachthonius 29		Neonothrus	71
	9	*Nothrus	69, figs. 179-182,
Hypochthonioidea	° figo 51-52	hDl a tarra a thanks	186, 310
	8, figs. 51-52	*Platynothrus	69-71, figs. 183-185, 309
(=Hypochthoniella)		WT-share a shake and in a	
*Eohypochthonius 27	7-28, figs. 44-45	*Trhypochthonius	35, fig. 69
*Eohypochthonius 27 *Hypochthonius 28	7-28, figs. 44-45 8, figs. 49-50	Trhypochthoniellus	35, fig. 69 s 35
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307	Trhypochthoniellus *Trimalaconothrus	35, fig. 69
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea)	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea	35, fig. 69 s 35 36, fig. 72
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43	Trhypochthoniellus *Trimalaconothrus	35, fig. 69 ³⁵ 36, fig. 72 95-96, figs. 264-265,
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=H *Atopochthonius 26 *Pterochthonius 25	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea)	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia	35, fig. 69 s 35 36, fig. 72
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 25 Protoplophoroidea	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea	35, fig. 69 5 35 36, fig. 72 95-96, figs. 264-265, 311
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 25 Protoplophoroidea *Cosmochthonius 26	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia	35, fig. 69 ³⁵ 36, fig. 72 95-96, figs. 264-265,
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA	35, fig. 69 5 35 36, fig. 72 95-96, figs. 264-265, 311
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea	35, fig. 69 5 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=H *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella	35, fig. 69 5 35 36, fig. 72 95-96, figs. 264-265, 311
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea	<pre>35, fig. 69 s 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes	<pre>35, fig. 69 35, fig. 72 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes	<pre>35, fig. 69 35, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes	<pre>35, fig. 69 5 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176 68, fig. 173-175, 297 68</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea *Gehypochthonius 36	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47 4, fig. 65	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes Teleioliodes	35, fig. 69 35, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 1775, 297 68 68
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=H *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea *Gehypochthonius 36	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47 4, fig. 65 4, fig. 65	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes Plateremaeoidea (=0	<pre>35, fig. 69 s 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176 68, fig. 173-175, 297 68 68 Gymnodamaeoidea)</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea *Gehypochthonius 36 *Parhypochthonius 36	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47 4, fig. 65 4, fig. 65	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes Plateremaeoidea (=0 *Gymnodamaeus	<pre>35, fig. 69 s 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176 68, fig. 173-175, 297 68 68 Gymnodamaeoidea) 63, fig. 159</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea *Gehypochthonius 36 *Parhypochthonius 36 MIXONOMATA (=PTYCTIM	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47 4, fig. 65 4, fig. 66 INA pt)	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes Teleioliodes Plateremaeoidea (=0 *Gymnodamaeus *Jacotella	<pre>35, fig. 69 s 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176 68, fig. 173-175, 297 68 68 Gymnodamaeoidea) 63, fig. 159 64, figs. 162, 298-299</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=H *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea *Gehypochthonius 34 *Parhypochthonius 34 MIXONOMATA (=PTYCTIM) Phthiracaroidea *Atropocarus 38	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47 4, fig. 65 4, fig. 65 1NA pt) 8	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes Teleioliodes Plateremaeoidea (=0 *Gymnodamaeus *Jacotella *Johnstonella	<pre>35, fig. 69 s 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176 68, fig. 173-175, 297 68 68 Symnodamaeoidea) 63, fig. 159 64, figs. 162, 298-299 63</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=1 *Atopochthonioidea (=1 *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea *Gehypochthonius 34 *Parhypochthonius 34	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47 4, fig. 65 4, fig. 65 4, fig. 66 INA pt) 8 8, fig. 78	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes Flateremaeoidea (=0 *Gymnodamaeus *Jacotella *Johnstonella	<pre>35, fig. 69 s 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176 68, fig. 173-175, 297 68 68 Gymnodamaeoidea) 63, fig. 159 64, figs. 162, 298-299 63 64, figs. 163</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=H *Atopochthonioidea (=H *Atopochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea *Gehypochthonius 34 *Parhypochthonius 34 *Hoplophorella 35 Hoplophthiracarus 35	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47 4, fig. 65 4, fig. 65 4, fig. 66 INA pt) 8 8, fig. 78	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes Plateremaeoidea (=0 *Gymnodamaeus *Jacotella *Johnstonella *Joshuella Licnodamaeus	<pre>35, fig. 69 s 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176 68, fig. 173-175, 297 68 68 Gymnodamaeoidea) 63, fig. 159 64, figs. 162, 298-299 63 64, fig. 163 62, fig. 159</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=H *Atopochthonius 26 *Pterochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea *Gehypochthonius 34 *Parhypochthonius 34 *Parhypochthonius 34 *Parhypochthonius 34 *Parhypochthonius 34 *Hoplophorella 38 *Hoplophorella 38 *Phthiracarus 38	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47 4, fig. 65 4, fig. 65 4, fig. 66 INA pt) 8 8, fig. 78	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes Plateremaeoidea (=0 *Gymnodamaeus *Jacotella *Johnstonella Licnodamaeus *Nortonella	<pre>35, fig. 69 s 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176 68, fig. 173-175, 297 68 68 Gymnodamaeoidea) 63, fig. 159 64, figs. 162, 298-299 63 64, fig. 163 62, fig. 159 64</pre>
*Eohypochthonius 27 *Hypochthonius 28 *Mesoplophora 37 Atopochthonioidea (=H *Atopochthonioidea (=H *Atopochthonius 26 Protoplophoroidea *Cosmochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Phyllozetes 26 *Sphaerochthonius 26 Protoplophora 37 Unplaced genus Nipponiella 26 PARHYPOSOMATA Parhypochthonioidea *Gehypochthonius 34 *Parhypochthonius 34 *Parhypochthonius 34 *Parhypochthonius 34 *Parhypochthonius 34 *Hoplophorella 36 *Hoplophorella 36 *Phthiracarus 38 *Phthiracarus 38	7-28, figs. 44-45 8, figs. 49-50 7, figs. 76-77, 307 Phyllochthonoidea) 6, fig. 43 5, figs. 39-40 6, figs. 41-42, 48 6 6, fig. 46 7, fig. 75 6, fig. 47 4, fig. 65 4, fig. 65 4, fig. 66 INA pt) 8 8, fig. 78	Trhypochthoniellus *Trimalaconothrus Nanhermannioidea *Nanhermannia Hermannioidea *Hermannia BRACHYPILINA Hermannielloidea *Hermanniella Liodoidea Liodes *Platyliodes Poroliodes Plateremaeoidea (=0 *Gymnodamaeus *Jacotella *Johnstonella *Joshuella Licnodamaeus	<pre>35, fig. 69 s 35 36, fig. 72 95-96, figs. 264-265, 311 96, figs. 267-268, 317 67, figs. 170-172, 305 68, fig. 176 68, fig. 176 68, fig. 173-175, 297 68 68 Gymnodamaeoidea) 63, fig. 159 64, figs. 162, 298-299 63 64, fig. 163 62, fig. 159</pre>

D1 J	()		101 100 51- 000
Pleodamaeus			101-102, fig. 288
Damaeoidea (=Belbo:	73, figs. 193-195	Conoppia	78
*Belba *Caapabalba	73, figs. 193-193, 300	*Eupterotegaeus	98, figs. 273-274, 293
*Caenobelba		*Ommatocepheus *Oribatodes	101, figs. 289, 296
*Damaeus	72 74 figs 107-109		101-102
*Dasybelba	74, figs. 197-198	*Sphodrocepheus	101, figs. 282-283,
Dyobelba	73	a 1 1 1 1	294-295
*Epidamaeus	73, 75, figs. 199-201	Carabodoidea * <i>Carabodes</i>	102 Fina 004 006
*Hungarobelba	74, figs. 196, 302 72		103, figs. 284-286
*Quatrobelba	72, figs. 190-191	*Dolicheremaeus	88, 103, fig. 291
*n. genus	74, figs. 202-203	Niphocepheus	103, fig. 290
*n. g <i>enus</i> Cymbaeremaeoidea	74, II93. 202-203	Odontocepheus Tootoophooidee	103
*Ametroproctus	99-100, figs. 280-281	Tectocepheoidea	102, figs. 287, 318
(=Coropoculia)	99-100, 1198 . 200-201	*Tectocepheus	102, 1195. 207, 318
Micreremus	95	<i>Tegeocranellus</i>	105
*Scapheremaeus	95, 99, figs. 189,	Oppioidea <i>Aeroppia</i>	91
*Scapiler emaeus	262-263, 272	Allosuctobelba	90
Amerobelboidea (=E		Ameroppia	
Amerus	84	*Autogneta	91, fig. 261 91
*Basilobelba	74, 97, fig. 269	*Banksinoma	91, fig. 252
Dameolus	97	Brachioppiella	91, 119. 252 91
*Eremobelba	97, fig. 271	Cosmoppia	92
Eremulus	97	*Gemmazetes	91-92
Fosseremus	97	Microppia	92
Eremaeoidea		Multioppia	91
*Eremaeus	85, figs. 224, 231-235,		91, figs. 255, 258
EI Omacub	304	*Oppiella	91-92, figs. 225, 254,
*Kartoeremaeus	85, 97, fig. 270	oppiolid	256-257, 259
*Megeremaeus	85, figs. 230, 303	*Oribella	91
Gustavioidea (=Lia		Pantelozetes	92
Adoristes	78-79	*Quadroppia	89, 91, figs. 250-251
*Ceratoppia	76, figs. 205-207, 312	Ramusella	91
*Cultroribula	81	*Rhinosuctobelba	86, figs. 236-238, 240,
Dorycranosus	79		249
*Furcoribula	81, fig. 222	*Rhynchobelba	86, fig. 239
*Gustavia	79, figs. 209, 211, 213	3 *Suctobelba	90, fig. 247
*Kodiakella	78, fig. 212	*Suctobelbella	90, figs. 248, 253
*Leuroxenillus	79, 100, figs. 275-276	Suctobelbila	90
*Liacarus	79, figs. 115, 214-221,	*Veloppia	91, 96
	313	Hydrozetoidea	
Metapyroppia	78	*Hydrozetes	88, figs. 244-246
*Metrioppia	78, figs. 208, 210	Limnozetes	59, fig. 147
Opsioristes	79	Ameronothroidea	
Paenoppia	79	*Alaskozetes	89, fig. 260
*Parapyroppia	76	Ameronothrus	96
*Peltenuiala	46, figs. 97, 99, 102	(=Hygroribates)	
Procorynetes	79	Licneremaeoidea (=	
Pyroppia	76	*Exochocepheus	103, fig. 292
Rhaphidosus	79	Licneremaeus	96, fig. 266
Stenoxenillus	100, fig. 277	*Scutovertex	102, fig. 279
*Stonyxenillus	78-79, 100, fig. 77	PTEROGASTERINA	
*Tenuiala	46, figs. 96, 100-101,		
4m	104	*Dometorina	88 82 8/ fin 200
*Tenuialoides	46	*Eporibatula	83-84, fig. 229
*Xenillus	79, 100	*Gerloubia	83-84
Cepheoidea		Gymnobates	83

Gymnobatoides Haplozetes	83 57, fig. 143	Pelopsis (=Parapelops)	58
Hemileius	88	*Podoribates	59
Liebstadia	84	Punctoribates	58
Lucoppia	82	*Sphaerozetes	58-59, figs. 144-145,
Metaleius	88		148
Neoribates	43, fig. 89	*Trichoribates	59
Oribatula	88, fig. 241	*Zachvatkinibates	59
*Oripoda	57, 83, fig. 226	Zetomimus	61
Parakalumna	43	Phenopelopoidea	
Paraleius	88	*Eupelops	49-50, figs. 107, 111-
*Paraphauloppia	84		112, 114, 296
Parapirnodus	82	*Peloptulus	50, fig. 108
*Peloribates	57, fig. 146	*Propelops	49, figs. 106, 110,
*Phauloppia	88, figs. 242-243		113, 297
*Scheloribates	56-57, figs. 117-118,	Oribatelloidea	
***********	135-142	Adoribatella	53
*Xylobates	57	*Oribatella	52-53, figs. 120, 124-
*Zygoribatula	82, figs. 223, 227-228		126
Ceratozetoidea		Achipterioidea	-
*Ceratozetes	59-61, figs. 116, 151- 155	*Achipteria	45-46, 53, figs. 95, 98, 103
*Chamobates	59	*Anachipteria	47-49, 53, figs. 105,
Dentizetes	54, fig. 130	Anachipteria	109, 119, 121-122,
*Diapterobates	58		131
Euzetes	59	*Lepidozetes	54, fig. 127
*Fuscozetes	59	*Parachipteria	45
Heterozetes	59	Pseudachipteria	45
*Humerobates	59	Scutozetes	54
*Hypozetes	60, fig. 156	Tectoribates	53
Iugoribates	61	(=Anoribatella)	
*Jugatala	54, figs. 132-134	Tegoribates	54
*Melanozetes	59, figs. 149-150	Galumnoidea	
Minuthozetes	57	Acrogalumna	43, fig. 91
*Mycobates	54, 57, figs. 128-129	*Galumna	43
(=Calyptozetes)		*Pergalumna	43, figs. 92-94
Neogymnobates	83	Pilogalumna	43, fig. 90

* indicates definite records from the Pacific Northwest.

ABSTRACT

Moldenke, Andrew; Fichter, Becky. Invertebrates of the H. J. Andrews Experimental Forest, western Cascade Mountains, Oregon: IV. The oribatid mites (Acari: Cryptostigmata). Gen. Tech. Rep. PNW-GTR-217. Portland, OR: U. S. Department of Agriculture, Forest Service, Pacific Northwest Research Station; 1988. 112 p.

A fully illustrated key is presented for identifying genera of oribatid mites known from or suspected of occurring in the Pacific Northwest. The manual includes an introduction detailing sampling methodology; an illustrated glossary of all terminology used; two color plates of all taxa from the H. J. Andrews Experimental Forest; a diagrammatic key to the 16 major groups of adult oribatids; illustrations to aid in quick recognition of the 16 adult groups; keys to all adult genera; and a diagrammatic key to the major types of immature oribatid mites with local species illustrated. The text on adults is broken into 16 sections, each structured around a key to the genera within each major type of oribatid mite; the text associated with the keys describes each species from the Andrews Forest, its microhabitat preference, pattern of abundance, and seasonal phenology. A total of 246 scanning electron micrographs (with pointers indicating characters useful for identification) are provided to help in identifying species.

Keywords: oribatid mites, soil ecology, key.

TABLE OF CONTENTS

Taxonomic Listing of Pacific Northwest Oribatid Genera	i-iii
Introduction Map 1: Andrews Forest Table 1: Sample Design	1 2 3
Methods	5
Andrews Forest Oribatid Fauna Table 2-3: Summary of Distributional Preferences Fig 1: Seasonal Abundance of Andrews Forest Oribatids	5 6 8
Acknowledgments	9
References	10
Illustrated Morphologic Oribatid Structures	11
Glossary	12
Color Comparison of all Andrews Forest Species	18
Diagrammatic Key to Major Types of Adult Oribatid Mites	20
Representative Illustrations of Major Oribatid Types	22
Text for Adults Section A: Enarthronotine mites Section B: Palaeosomatine mites Section C: Translucent oribatids Section D: Penknife mites Section E: Galumnoid winged wites Section F: Spine-shouldered winged mites Section G: Winged mites Section G: Winged mites Section H: Flat-backed stilt mites Section I: Lohmannioid mites Section J: Hermanniellioid mites Section J: Hermanniellioid mites Section L: Damaeoid Mites Section M: Lamellate smooth-backed mites Section N: Non-lamellate smooth-backed mites Section P: Non-lamellate patterned-backed mites	25 32 34 37 43 45 48 62 65 67 68 72 76 82 95 98
Representative Illustrations of Major Types of Immatures	105
Diagrammatic Key to Major Types of Immatures	108
Index	110

INTRODUCTION

Densities of adult oribatids on the forest floor in H. J. Andrews Experimental Forest range from 15,000 to 120,000 per square meter. About 60 species are abundant and found in most litter samples collected there; these species are also the most abundant and widespread species throughout the coniferous biome of the Pacific Northwest.

The Andrews Forest has been designated a Long Term Ecological Reserve by the National Science Foundation, and functions as an integral cornerstone in understanding the ecology of North America as part of a continent-wide system of special research sites (Map 1). These sites serve as foci of integrative studies of ecological dynamics, especially involving changes spanning temporal phenomena longer than the term of typical research grants. Scientists at these sites have the responsibility to facilitate correlative studies throughout the ecological biome they represent, hence every effort was made in the present work to prepare a key of generalized utility throughout the entire Pacific Northwest.

Almost nothing is known about how these species function in nutrient cycling or in regulating populations of other soil fauna and microbes. Ecological investigations into the decomposition guild have been seriously hampered for many years because mites are so small and taxonomic reviews are lacking. We have designed this paper for nonspecialists to present detailed information on the distribution, seasonality, and abundance of the mites in the Andrews Forest; to provide a key to the species; and to stimulate ecological studies on the functional roles of oribatid mites throughout the Northwest, by providing a utilitarian key to all the genera likely to be found.

The Pacific Northwest coniferous biome extends from southern British Columbia through Washington and Oregon to coastal central California and the southernmost Cascades (Mount Lassen, California). Of the 209 genera treated in this key, about threequarters are definitely known to occur in the Pacific Northwest (indicated by * in Taxonomic List of Northwest Genera). The other sixty are deemed likely to be found in additional studies in the region (Valin Marshall, Pacific Research Centre, Canadian Department of Forestry, Victoria; and Roy Norton, Syracuse University, Syracuse, New York; pers. comm.). About 100 genera known to occur in North America (Marshall et al. 1986) were not considered in the present work based on Norton's knowledge of their distribution patterns.

The three major sections of this paper are:

a) A pictorial key to major morphological groups of adult oribatids,

based on characters easily recognized under a dissecting microscope. Some groups are artificial, but each one is a coherent morphological unit. The first composite plate (figs. 3 to 23) displays typical representatives of each group. Figures 24 to 26 show uropodine mesostigmatid mites which occur in most litter samples from the Andrews Forest. They may be confused with oribatids because mites in the two groups are similar in size, shape, and color.

b) Keys to genera in each broad morphological group of adult oribatid mites. We provide 16 separate keys to decrease the confusion inherent in identifying an unfamiliar group.

c) A pictorial key to the major morphological groups of immature oribatids, using characters easily visible under a dissecting microscope.

INTRODUCTION

Densities of adult oribatids on the forest floor in H. J. Andrews Experimental Forest range from 15,000 to 120,000 per square meter. About 60 species are abundant and found in most litter samples collected there; these species are also the most abundant and widespread species throughout the coniferous biome of the Pacific Northwest.

The Andrews Forest has been designated a Long Term Ecological Reserve by the National Science Foundation, and functions as an integral cornerstone in understanding the ecology of North America as part of a continent-wide system of special research sites (Map 1). These sites serve as foci of integrative studies of ecological dynamics, especially involving changes spanning temporal phenomena longer than the term of typical research grants. Scientists at these sites have the responsibility to facilitate correlative studies throughout the ecological biome they represent, hence every effort was made in the present work to prepare a key of generalized utility throughout the entire Pacific Northwest.

Almost nothing is known about how these species function in nutrient cycling or in regulating populations of other soil fauna and microbes. Ecological investigations into the decomposition guild have been seriously hampered for many years because mites are so small and taxonomic reviews are lacking. We have designed this paper for nonspecialists to present detailed information on the distribution, seasonality, and abundance of the mites in the Andrews Forest; to provide a key to the species; and to stimulate ecological studies on the functional roles of oribatid mites throughout the Northwest, by providing a utilitarian key to all the genera likely to be found.

The Pacific Northwest coniferous biome extends from southern British Columbia through Washington and Oregon to coastal central California and the southernmost Cascades (Mount Lassen, California). Of the 209 genera treated in this key, about threequarters are definitely known to occur in the Pacific Northwest (indicated by * in Taxonomic List of Northwest Genera). The other sixty are deemed likely to be found in additional studies in the region (Valin Marshall, Pacific Research Centre, Canadian Department of Forestry, Victoria; and Roy Norton, Syracuse University, Syracuse, New York; pers. comm.). About 100 genera known to occur in North America (Marshall et al. 1986) were not considered in the present work based on Norton's knowledge of their distribution patterns.

The three major sections of this paper are:

a) A pictorial key to major morphological groups of adult oribatids,

based on characters easily recognized under a dissecting microscope. Some groups are artificial, but each one is a coherent morphological unit. The first composite plate (figs. 3 to 23) displays typical representatives of each group. Figures 24 to 26 show uropodine mesostigmatid mites which occur in most litter samples from the Andrews Forest. They may be confused with oribatids because mites in the two groups are similar in size, shape, and color.

b) Keys to genera in each broad morphological group of adult oribatid mites. We provide 16 separate keys to decrease the confusion inherent in identifying an unfamiliar group.

c) A pictorial key to the major morphological groups of immature oribatids, using characters easily visible under a dissecting microscope.



PLANT COMMUNITY SAMPLING DESIGN

	\longrightarrow successional gradient \longrightarrow						
	herb-dominated (1-7 yrs since clear cut)	shrub-dominated (7-15 yrs since clear cut)		old growth forest (300-400 yrs)			
xeric	open bare ground; sparse grasses & herbs			dense overstory Acer & Corylus; moderate Gaul- theria cover; dense moss			
	dense <i>Epilobium</i> & some with dense <i>Rubus</i>	dense <i>Ceanothus</i> & <i>Acer</i> ; bare ground	<pre>dense Douglas-fir & Acer; moderate to dense Gaultheria; moss present</pre>	dense moss cover; Berberis & Gaul- theria OR Rhododendron & bare soil			
mesic	sparse herbs; some <i>Rubus</i>	dense <i>Ceanothus</i> & <i>Acer</i> ; scat- tered herbs & fern; moss present	dense Douglas-fir & Acer; moderate Gaultheria, Poly- stichum & Oxalis; dense moss	dense Polystichum Oxalis, Berberis; dense moss			

TABLE 1: At least 4 replicate samples were taken at six times each during a two year period for each one of the 4 successional x 3 moisture matrix communities. Franklin & Dyrness (1973) community designations for the different climax communities are: xeric: Pseudotsuga menziesii/Tsuga heterophylla; Corylus cornuta;

mid-moisture: Tsuga heterophylla; Rhododendron macrophyllum/Berberis nervosa; mesic: Tsuga heterophylla; Polystichum munitum/Oxalis oregona. The four sections of photographs are:

d) Two color plates with Andrews Forest oribatids arranged in a continuous sequence by size. Resolution is not sufficient to identify them without a key. All taxa are placed in both the size and color context of an actual sample. In fieldcollected samples, teneral specimens (not yet fully melanized) in the process of moulting are encountered; only practical experience in sorting samples produces the ability to associate these unusual color forms with the appropriate fully adult morph.

e) A "Rogue's Gallery" of generalized adult oribatid morphotypes (figs. 3 to 23). These 16 taxa represent groups to which any oribatid collected from the forests of the Pacific Northwest can be assigned with cursory examination. Occasionally, these categories are congruent with currently recognized taxonomic entities (one to several superfamilies), but often they are artificial categories useful only in identification. This non-natural approach recognizes that many taxonomically important characters are extremely difficult for the novice taxonomist or ecologist to recognize.

f) Photographs of Andrews Forest oribatids and representative genera from other areas of the Pacific Northwest. The photographs were not chosen on esthetic grounds (many specimens carry material or cerotegument as they normally do in nature) nor to serve in a monographic revision. Most taxa are not illustrated on all surfaces, nor are all setae adequately demonstrable. We believe the pictures are appropriate to this introductory effort, however, as a means of identifying oribatid mites to genus and to show morphologic variability likely to be significant in sorting to the level of morphospecies.

g) A "Rogue's Gallery" of immature oribatids from the Andrews Forest. The life cycles of this fauna have not been studied in detail, hence this section represents an introductory aid to identifying the general group to which an immature oribatid belongs. Generic and specific distinctions are beyond the scope of this work.

All genera found in the Andrews Forest or suspected of inhabiting this region (Balogh 1972; Balogh and Mahunka 1982; Marshall et al. 1986; Norton, pers. comm.) are included in this treatment. The nomenclature adopted is that recommended in the oribatid catalog (Marshall et al. 1986), likely the only reference available to a general audience. Reference is provided to Balogh (1972) when nomenclatural treatments differ. Information on distribution of genera not yet found in the Pacific Northwest has been provided by the editors of the oribatid catalog (in press) and represents published citations; presumption that these genera may occur in the Pacific Northwest is based on the professional experience of specialists who edited this manuscript. The authors learned of the potential Northwestern distribution of the genera Allodamaeus (Plateremaeoidea), Grypoceramerus (Amerobelboidea), Spinozetes (Oppioidea), Provertex and Passalozetes (Passalozetoidea) for inclusion in the present text.

In some families, such as the Oppiidae and Brachychthoniidae, closely similar genera and species may be too difficult for novices to separate. Descriptive characters for all the North American genera in these groups are presented in tables for quick comparison. Taxonomic specialists should be consulted for assistance with these groups.

METHODS

A sampling design encompassing the principal tree associations of the Andrews Forest Long Term Ecological Reserve (Franklin and Dyrness 1973) was adapted in 1982 by Gary Parsons (Graduate Research Assistant, Oregon State University) to study insect fauna of the soil and litter. Each of eight climax vegetation associations was sampled in each of four successional stages; from this sampling design, we analyzed mesic, midmoisture, and xeric sequences of successional seres (Table 1). Sampling was conducted at different seasons during a 2-year period. Random samples of litter and soil were taken to a depth of 7 cm in two subsamples (later combined) of 25 x 25 cm each. Differential contribution of litter versus soil to the sample was not recorded. Samples were processed in Tullgren funnels and collected in 75-percent ethanol.

At least four replicate samples were taken from each of 12 community types (matrix of four successional seres by three moisture regimes). A total of 318 samples were were brought to the laboratory, agitated with mineral oil, and the inorganic debris allowed to settle to remove inorganic debris. Larger invertebrates were removed and the remaining mesofauna placed in a 50- \times 9-mm petri plate in a film of mineral oil for identification and permanent storage. A subsample (a half or quarter of the total) was sorted in samples with abundant oribatids. Original data and specimens are available in the H. J. Andrews reference collection at Oregon State University.

ANDREWS FOREST ORIBATID FAUNA

The response of the oribatid fauna to the moisture gradient is presented in Table 2. Preference for a particular stage in succession or microhabitat is presented in Table 3. Species without a consistent preference are omitted from the tables (principally rare or uncommon species).

The pattern of seasonal abundance for each species is described in the text of the key. Most Andrews Forest oribatid species follow the same generalized phenological abundance curve, and hence the abundance of each species is compared to the phenology of total oribatids (fig. 1). Total oribatid seasonal abundance is strongly bimodal within narrow estimates of standard deviation. Abundance of nearly all species of oribatids (and likewise of total oribatids) is highest in spring and fall.

Table 2Summary of Distribution Preferences in Andrews Forest

driest conditions

Anachipteria small Brachychthonius Ceratozetes 310µ Ceratozetes 460µ Ceratozetes 580µ Ceratoppia small Eohypochthonius Hypochthonius Jacotella Leuroxenillus Nanhermannia Scheloribates 380µ Scheloribates oblong Zygoribatula Oribatella 290-340µ

driest & mid-moisture

Anachipteria big Cultroribula Gustavia Oribotritia megale Perlohmannia small Sphaerozetes

mid-moisture

Eporibatula Eupelops Hungarobelba Propelops

mid-moisture to mesic conditions

Anachipteria (cerotegument) Liacarus tan Liochthonius Scheloribates 280µ Scheloribates 460µ

most mesic conditions

Ceratoppia big Ceratozetes 380µ Epilohmannia Euphthiracarus Hermanniella small Metrioppia Oppiella small Oribatella mid Oribatella big Peltenuiala pacifica Perlohmannia big Quadroppia quadricarinata Sphodrocepheus anthelionis

no preference exhibited

Achipteria	Nothrus silvestris			
<i>Caenobelba</i>	<i>Maerkelotritia</i> big			
Carabodes	Maerkelotritia small			
Ceratoppia	<i>Oppiella</i> big			
Damaeid n.genus #2	Phthiracarus big			
Epidamaeus	Phthiracarus small			
Eremaeus	Rhinosuctobelba			
<i>Hermanniella</i> big	Sphodrocepheus			
Hermanniella small	tridactylus			
Liacarus big spp.	Tectocepheus velatus			

Table 3

Summary of Distribution Preferences in Andrews Forest

<u>herb stage of clear-cut</u> Ceratozetes 460µ Ceratozetes 580µ Jacotella

<u>herb stage & shrub stage</u> Achipteria Brachychthonius Propelops Scheloribates 380µ Anachipteria (cerotegument)

shrub stage after clear-cut Eupelops Gustavia Hungarobelba Maerkelotritia big Oribatella mid Oribatella big Oribotritia megale Scheloribates 460µ Scheloribates oblong Zygoribatula

shrub stage & tree stage Hermanniella small Leuroxenillus Liochthonius Maerkelotritia small Metrioppia Nothrus silvestris Perlohmannia big

<u>tree stage regrowth</u> Eporibatula Euphthiracarus Nanhermannia Sphodrocepheus anthelionis tree regrowth & old growth Anachipteria small Carabodes Ceratoppia mid Damaeid n.genus #2 Epilohmannia Oribatella small Phthiracarus small Tenuiala

old growth forests Ceratoppia big Eremaeus Hermanniella mid Hermanniella big Liacarus tan Oppiella small Peltenuiala pacifica Phthiracarus big Sphaerozetes Sphodrocepheus tridactylus

arboreal old growth canopy Camisia carrolli Eremaeus 2 spp. Hypozetes Jugatala 2 spp. Odontodamaeus veriornatus Platyliodes macroprionus Scheloribates sp. Scapheremaeus

no preference indicated Caenobelba Ceratozetes 310µ Eohypochthonius Epidamaeus Liacarus big spp. Maerkelotritia big Oppiella large Perlohmannia small Quadroppia quadricarinata Rhinosuctobelba dicerosa Suctobelbella spp. Tectocepheus velatus



Figure 1: Bimodal pattern of abundance of total oribatid species at Andrews Forest. Means and standard deviations are calculated from the percentage of annual capture of each of the 90 commonest oribatid species. Data are calculated species by species to avoid bias inherent in the sampling design in which all sites were not sampled equivalently at all seasons.

ACKNOWLEDGMENTS

Specimens were taken primarily from those collected by Gary Parsons for his thesis research on insects of the soil and litter layers of the Andrews Forest Long Term Ecologic Reserve. Correlative unpublished studies of oribatid mites from nearby Washington, and California add significantly localities in Oregon, to the generalizations presented herein (Topic 1982; Forest floor accumulation and decomposition in the western Cascades of Oregon, Ph.D. thesis, University of Oregon; D. Walter, 1981; Patterns in a decomposer community: the oribatid mites of Monterey pine litter; M.S. thesis, University of California, Berkeley; and Seastedt and Cline, Microarthropod densities and composition in fallen and decaying trees of coniferous Special thanks go to David Walter and Gerald Krantz for forests (in preparation). sharing their expertise. Valin Marshall permitted access to an unpublished manuscript for checking nomenclatural usage and distributions, as well as providing specimens for photographs.

Valuable editorial suggestions were provided by Martha Brookes, Gerald Krantz, J. D. Lattin, Joanne Lawrence, Valin Marshall, and Roy Norton.

The scanning electron micrographs were taken by Albert Soeldner and his capable staff of the Botany and Plant Pathology Department, Oregon State University. Illustrative layout was meticulously performed by Stuart Sloan, who also prepared the half-tones for publication.

This paper represents a partial contribution (no. 17) of the project entitled "The Fallen Tree--an Extension of the Live Tree." Cooperating on the project are the U.S. Department of Agriculture, Forest Service, Pacific Northwest Research Station; Oregon State University, Department of Forest Science; U. S. Department of Agriculture, Agricultural Research Service; and Oregon Department of Fish and Wildlife.

Funding for this research was provided in part by the National Science Foundation (DEB 8012162 and BSR 8514325: Long-Term Ecological Research on the HJ Andrews Forest; and BSR 8300370: Support of the HJ Andrews Experimental Forest as a National Field Research Facility).

REFERENCES

Andre, Henri M.; Voegtlin, David J. Some observations on the biology of *Camisia carrolli* (Acari: Oribatida). Acarologia 23: 81-89. 1981.

Balogh, J. The oribatid genera of the world. Budapest: Akad. Kaido; **1972**. 188 p. + 71 plates.

Balogh, J.; Mahunka, S. Primitive oribatids of the Palaearctic region. Oxford: Elsevier Press; 1983. 372 p.

Franklin, Jerry F.; Dyrness, C.T. Natural vegetation of Oregon and Washington. Gen. Tech. Rep. PNW-8. Portland, OR: U.S. Department of Agriculture, Forest Service, Pacific Northwest Forest and Range Experiment Station. **1973.** 417 p.

Grandjean, F. Essai de classification des Oribates (Acariens). Bulletin Societe Zoologie de France 78: 421-446. **1954**.

Krantz, G.W. A manual of Acarology, 2d ed. Corvallis, OR: Oregon State University Bookstore; 1978. 509 p.

Marshall, V.G.; Reeves, R.M.; & Norton, R.A. Catalogue of the Oribatida (Acari) of Continental United States and Canada. Memoirs Entomological Society Canada #139. 1987. 418 p.

Rockett, C.L. Nematode predation by oribatid mites (Acari: Oribatida). International Journal of Acarology 6: 219-224. 1976.

Voegtlin, D. Invertebrates of the H. J. Andrews Experimental Forest: a survey of arthropods associated with the canopy of old growth *Pseudotsuga menziesii*). Forest Research Laboratory Special Publication #4, School of Forestry, Oregon State University, Corvallis, OR. **1982**.

Walker, N.A. Euphthiracaroidea of California sequoia litter: with a reclassification of the families and genera of the world. Fort Hays Studies (New Series) Science #3. 1964. 154 p.

Wallwork, J.A. Some basic principles underlying the classification and identification of cryptostigmatid mites. pp. 155-168 In: Sheals, J. G., ed. The soil ecosystem. London: Systematics Association Publ. #8, 1969.

Wooley, Tyler A. North American Liacaridae III: new genera and species (Acari: Cryptostigmata). Journal Kansas Entomological Society 42: 183-184. 1969.



ILLUSTRATED MORPHOLOGIC ORIBATID STRUCTURES

GLOSSARY

(abbreviations in parentheses are utilized on the figures)

ABUNDANCE: relative density of one species compared to all others in sample, averaged over all samples.

ADANAL SCLERITE (AAS): ventral shield just to the side of the anal sclerite. Very few genera have adanal sclerites.

ADANAL SETAE: group of setae located just to the side of the anal sclerite, either on adanal sclerite, if present, or on the ventral surface.

ADGENITAL SCLERITE: pair of shields located just to the side of the genital sclerites. Very few of our genera have adgenital sclerites.

ADGENITAL SETAE: group of setae located just to the side of the genital plates or on the adgenital sclerite, if present.

ADNATAE: see spinae adnatae.

ANAL SCLERITE (AS): ventral shield located lateral to the anus and posterior to the genital shield.



Figure 2: Diagrammatic ventral views of advanced (left) and primitive oribatids, showing the position and number of ventral sclerites.

ANTERIORLY: toward the front of the body.

ANTEROMESALLY: on the midline towards the front of the body.

APOPHYSIS: projection from the body surface.

AREAE POROSAE (AP): thinner portions of the notogaster supplied with dense fine tubular pores and thus appearing micropunctate. Usually in four pairs, if present.

BIPECTINATE: referring to a branched seta, both branches of which are comblike.

BIRAMOUS: with two branches, like a Y or a T.

BEADED LEGS: the legs of the Damaeoidea, Plateremaeoidea, and some small Oripodoidea, which are alternately swollen and narrow, resembling a necklace made from pop beads ("moniliform").

BOTHRIDIUM (B): the socket on the prosoma that gives rise to the sensillum. Usually directed laterally; if directed dorsally or hidden by the dorsosejugal suture, it is useful in identification.

CAPITATE: shaped like a ball (head) on top of a thin neck; referring to setae or the sensilla.

CARINA: a thickened ridge (usually linear) rising above the surrounding flat surface.

CARINATE: with a carina or ridge.

CEROTEGUMENT (CT): a waxy exudate covering the external body surfaces, often thick enough to obscure the setae or surface patterning.

CHELICERAE (CH): appendages near the mouth, which are pincher-like. In *Gustavia*, *Eupelops*, *Rhinosuctobelba*, and *Suctobelbella* they are so modified that they no longer resemble lobster-claws.

CLAVATE: club-shaped; referring to setae or the sensillum.

CONSTANCY: the number of subsamples in any of the 12 community types in which the species occurred in an abundance of more than three individuals.

COSTULA: very reduced lamellae on the surface of the prodorsum, which appear as inconspicuous longitudinal ridges ending anteriorly at the insertions of the lamellar setae.

CRISTA: a saddle-like prominence on the anteromesal portion of the notogaster of the Oripodoidea. Varies from a pair of lateral tubercules (concave between) to a pair of long, high ridges flanking a deep depression.

CUSP: tooth; generally referring to the shape of the tip of the lamella.

DECUMBENT: lying down; referring to setae relative to the body surface.

DENTATE: with tooth-like projections, usually at the tip.

DORSAL SETAE: setae located on the top surface of the notogaster.

DORSOSEJUGAL SUTURE (DSJ): the groove separating the prodorsum from the notogaster in dorsal view.

EMARGINATE: with an irregular margin, as if a bite had been removed from the edge.

EPIMERES: shields on the venter of the prosoma to which the legs and leg muscles are attached.

EPIPHYTIC: referring to mosses and lichens located on the branches of trees.

EUPTERY: presence of pteromorphs capable of flexion.

EXUVIAE: the shed skins of the nymphal stages; attached in regular geometric patterns to the surface of the adults of certain species.

FEMORAL: pertaining to the upper leg segment.

FILIFORM: thread-shaped.

FUSIFORM: spindle-shaped, wide in the middle and thin at both ends; referring to the shape of the sensillum.

GENITAL SCLERITES (GS): pair of shields surrounding the genital aperture in the center of the venter. A fully formed and distinct genital sclerite indicates adulthood.

GENITO-ANAL PLATES (GAP): mid-ventral longitudinal sclerites representing the fusion of the genital/anal sclerites and adgenital/adanal sclerites (in the Euphthiracaroidea only).

GENU: the leg segment between the femur and the tibia.

GLABROUS: smooth, shining, without setae.

GYMNONOTIC: refers to a notogaster without area porosae, sacculi, or pores.

HUMERAL ANGLE: region of the notogaster at the termini of the dorsosejugal suture; refers to the deviation from a smooth curve produced by any protrusion of the notogaster, such as pteromorphs.

INTERLAMELLAR SETAE (ILS): the prominent pair of setae on the posterior dorsal surface of the prosoma between the bases of the lamellae.

LAMELLA (L): flanged protective shield on the dorso-lateral portion of the prodorsum; fused to the surface along posteromesal edge (usually) and assuming many diverse forms (see illustration on next page for descriptive terminology).

LAMELLAR RIDGE (LR): refers to a pair of longitudinal ridges or "costulae" on the surface of the prodorsum, never present along with true lamellae.

LAMELLAR TOOTH (LT): the tooth-like anterior tip of the lamella.

LAMELLATE: with a pair of true lamellae.

LAMELLAR SETA: the seta located at the anterior tip of the lamellar cusp or lamellar ridge.

LANCEOLATE: spear-shaped, refers to setae or sensilla.

14



LATERO-ABDOMINAL GLAND: a glandular structure located under the postero-lateral portion of the notogaster and opening to the surface by a large pore. Visible in very few genera of the Northwest.

LENTICULUS: a blanched lens-shaped region in the anteromedial portion of the notogaster; found on species which are otherwise darkly melanized.

MANDIBULAE: the opposable joint at the tip of the chelicerae.

MENTAL TECTUM (MT): a broad shield on the vertral surface of the prosoma concealing the chelicerae in ventral view.

MESAL: towards the midline.

MONODACTYLY: having a single tarsal claw per leg. Characteristic of all immature oribatids but very few adults except for the smallest species.

MULTISETOSE: with many setae.

NOTOGASTER: the shield covering the dorsum of the opisthosoma.

NYMPHAL SHED SKINS (NSS): exuviae of immature stages carried on the dorsum of the adult of certain species.

OPISTHONOTAL SUTURES: transverse sutures across the entire width of the opisthosoma, characterisitc of the Enarthronota.

OPISTHOSOMA: the posterior body segment; the region of the body posterior to the fourth pair of legs.

PECTINATE: comb-shaped; refers to the sensillum.

PEDOTECTUM (PT): a protrusion arising from above the base of the legs and extending forward and laterally over the leg.

PICTURE HEIGHT: the height (in microns = μ) of the full frame pictured.

PERIANAL PLATES: see adanal sclerites.

15

2

ini.

PLUMOSE: feather-like; refers to setae that are covered with finer setae.

PORI: four pairs of sac-like structures under the notogaster and opening to the surface by a constricted pore (see sacculi).

PORONOTIC: having pori under the notogaster.

PRODORSUM: the dorsal portion of the mite anterior to the notogaster.

PTEROMORPHS (PTEROGASTERS) (P): wing-like extensions of the notogaster in the humeral region.

PTEROMORPH SUTURE (PS): a sulcus resulting in an axis of flexure at the base of the pteromorphs. Few mites with pteromorphs are capable of flexing them.

PUBESCENT: with setae.

RETICULATE: a netlike surface pattern formed by fine surface ridges and grooves.

ROBUST: oval in outline, not elongate.

ROSTRAL SETAE: a pair of setae on all oribatid mites on the anterior tip of the snout.

ROSTRUM (R): the anterior portion of the dorsal surface of the snout.

RUTELLUM: horizontal plate enclosing the mouth region from below.

SACCULONOTIC: notogaster with sacculi.

SACCULI: four pairs of sac-like structures located under the notogaster and opening to the surface through a slit (see pori).

SCLERITE: a hardened, inflexible shield on the body surface.



SENSILLUM (S): the antenna-like structure arising from the bothridial socket. (for descriptive terminology see illustration).

SERRATE: saw-like, with a linear row of teeth.

SETIFORM: formed like a simple seta.

SETOSE: bearing setae.

SPATULATE: resembling the shape of a spatula (somewhat spoon-like).

SMOOTH PATCHES: smooth regions on the dorsal surface of the prosoma in the Suctobelboidea, contrasting with the warty or carinate remainder.

SPINAE ADNATAE: a pair of tubercular teeth arising at the end of the dorsosejugal suture from the anterior edge of the notogaster.

STIPPLING: a surface pattern of fine, dense, pore-like depressions.

SULCUS: a linear groove in the surface integument.

SUPRAPLEURAL PLATES: in the Brachychthoniidae, this term refers to a series of sclerites on the side of the body between the notogaster and the usual ventral sclerites (see fig. 58 for position designation).

SUTURE: a linear groove in the surface integument, along which movement is usually possible.

SYNLAMELLATE: refers to the lamellae when they are fused for most of their length and form a shelf over the prodorsum.

TRANSLAMELLA (TL): refers to lamellae fused for their basal half to the prodorsum, apical teeth connected by a transverse carina or ridge.

TRANSLAMELLAR TEETH (TT): any tooth-like structures found along the carina joining the tips of the translamella.

TRANSVERSE: across the longitudinal body axis; usually refers to a straight line perpendicular to body length.

TRANSVERSE SUTURE (TS): a region of flexure entirely across either the notogaster or opisthosomal venter.

TRIDACTYLY: the normal condition in adult oribatids, possessing three tarsal claws per foot.

TRUNCATE: cut off square-transversely at the tip.

TUBERCULATE: with a short, oval protrusion from the surface.

COLOR COMPARISON OF ANDREWS FOREST SPECIES



LEGEND:

ROW A: 1)Oribotritia megale; 2) large Maerkelotritia; 3) large Phthiracarus; 4) medium Phthiracarus; 5) Euphthiracarus tanythrix; 6) small Maerkelotritia; 7) small Phthiracarus; 8) Microtritia paeniminima.

ROW B: 1) Rhinosuctobelba dicerosa; 2) large Perlohmannia; 3) small Perlohmannia; 4) Epilohmannnia n. sp.; 5) Platylioides macroprioides; 6) Trachytes sp. (non-oribatid); 7) non-oribatid; 8) non-oribatid.

ROW C: 1) common Eremaeus; 2) large Hermanniella; 3) Jugatala tuberosa; 4) large Ceratoppia; 5) large Ceratoppia; 6) medium Ceratoppia; 7) arboreal Eremaeus; 8) medium Hermanniella; 9) Great Basin Eremaeus.

ROW D: 1) small Hermanniella; 2) Ametroproctus sp.; 3) Pergalumna sp; 4) small tan Liacarus; 5) Sphaerozetes sp.; 6) Zygoribatula sp.; 7) tear-shaped Scheloribates; 8) 160 μ Ceratozetes sp.; 9) Phauloppia sp.; 10) Metrioppia oregonensis; 11) 380 μ Ceratozetes sp.; 12) small Ceratoppia; 13) Trhypochthonius americanus; 14) Nanhermannia sp.; 15) Scapheremaeus sp.

ROW E: 1) 400 μ Scheloribates sp.; 2) 310 Ceratozetes sp.; 3) common small Oribatella; 4) tear-shaped Ceratozetes; 5) Tectocepheus velatus; 6) small Carabodes; 7) Oppiella nova; 8) large Suctobelbella; SPACE ; 9) Oppia sp.; 10) Jacotella sp.; 11) small Caenobelba; 12) Hungarobelba sp.; 13) new genus #2 of damaeid, near Hungarobelba; 14) Hungarobelba sp.; 15) Epidamaeus sp.

ROW F: 1) Epidamaeus sp.; 2) Brachychthonius sp.; 3) Eporibatula sp.; 4) Furcoribula sp.; 5) common Suctobelbella; 6) Quadroppia quadricarinata; 7) Oppiella sp.; 8) Zachvatkinella sp.; 9) Eohypochthonius sp.; 10) Hypochthoniella sp.; 11) Hypochthonius rufulus; 12) Eulohmannia ribagei.

COLOR COMPARISON OF ANDREWS FOREST SPECIES



LEGEND:

ROW A: 1) Caeculus sp. (non-oribatid); 2) Camisia spiniger; 3) Belba californica; 4) Platynothrus sp.; 5) Nothrus silvestris; 6) Camisia carrolli.

ROW B: 1) large Stonyxenillus; 2) Liacarus spiniger; 3) small Stonyxenillus; 4) Leuroxenillus sp.; 5) Megeremaeus ditrichosus; 6) Liacarus robustus.

ROW C: 1) Hermannia sp.; 2) Sphodrocepheus tridactylus; 3) Eupterotegaeus sp.; 4) Eupelops sp.; 5) subalpine Pergalumna; 6) large Carabodes; 7) warty cerotegument Anachipteria; 8) Belba californica.

ROW D: 1) Uropodid (non-oribatid); 2) large Hermanniella; 3) Peltenuiala californica; 4) small tan Liacarus; 5) large Oribatella; 6) Liacarus bidentatus; 7) large Caenobelba.

ROW E: 1) Achipteria sp.; 2) common Anachipteria; 3) Propelops canadensis; 4) Anachipteria with pedotecta; 5) large Caenobelba; 6) Odontodamaeus veriornatus.

DIAGRAMMATIC KEY TO MAJOR TYPES OF ADULT ORIBATIDS





DIAGRAMMATIC KEY TO MAJOR TYPES OF ADULT ORIBATIDS





ABUNDANT non - ORIBATID TYPES



NANORCHESTID

GAMASID

TROMBIDIID

PHALANGID

SECTION A: Enarthronota

All genera included here are small to very small oribatids, which require slide mounting and high power for a clear view of structures, except for the large, flat and pink *Hypochthonius rufulus* Koch. All these genera have 1 to 4 transverse sutures across the notogaster. They are often dorso-ventrally flattened. Large prominent setae, often highly modified and ornate, are characteristic. The guts of this entire group of mites seldom contain dark-pigmented food boli. This is a diverse group, all of which have transverse opisthonotal sutures. Immatures resemble adults and care must be exercised not to confuse them with adults of similar species.

1A	Dorsal setae modified (bipectinate, plate-like, or spatulate and very long) 2
1 B	Dorsal setae abundant but rather inconspicuous
	Dorsal setae plate-like or long-spatulate; notogaster with three transverse sutures
2B	Dorsal setae pectinate, long-pectinate, or elongate (never spatu- late) 4

3A Dorsum covered with large, scale-like setae that completely hide body in dorsal and lateral views (figs. 39 and 40); sensillum filiform; tan body Pterochthonius

Pterochthonius angelus (Berlese) (275 μ), collected rarely in Andrews Forest, has been seen sporadically from other locales in western Oregon.

40





Figure 39-40: 39, 40: *Pterochthonius angelus* with plate-like setae and filiform sensillum (S); picture height $39 = 300 \ \mu$, picture height $40 = 300 \ \mu$.



Figure 41-42: 41, 42: Cosmochthonius lanatus with three transverse sutures (3TS) and pectinate setae (A); picture height $41 = 300 \ \mu$, picture height $42 = 300 \ \mu$.

3B Dorsal setae of varying lengths and shapes, but the posterior transverse row always greatly elongated with bases much wider than apices; sensillum filiform with minute bristles; body color transparent (fig. 43) Atopochthonius

Atopochthonius artiodactylus Grandjean (200 μ) was encountered only once in these litter samples, but was found in rotting logs (Cline & Seastedt, unpub.). Phyllozetes, recorded from Texas, differs in having compoundly dentate setae on the prodorsum.

Only one Cosmochthonius nr. lanatus (Michael) (200 μ) was found in these litter samples; this species was more abundant in rotting logs (Cline & Seastedt, unpub.). Nipponiella, reported by Walter from central California, is quite similar but has four transverse sutures and less conspicuously plumose notogastral setae (fig. 47); the prosomal setae are elongate and not conspicuously plumose (platelike in Atopochthonius).

4B Dorsal setae pectinate biramous (fig. 46)..... Sphaerochthonius

Sphaerochthonius occurs widely in both eastern and western Oregon; however, it was found only once in the Andrews Forest research grid (midmoisture clearcut).

- reddish-pink; one transverse suture; body 5A Dorsum with bothridium not distinctly enlarged; sensillum pectinate 6 Dorsum with two or apparently two transverse sutures; body 5B not pink; bothridium usually hypertrophied, with a clubbed
- 6A Body oval (fig. 44) and smaller, ca. 350 μ; genital plates appear transversely divided (fig. 45) Eohypochthonius



.

sensillum ...

Figure 43-45; 43: Atopochthonius sp. with elongate flattened setae posteriorly and 3 transverse sutures (3TS); 44: Eohypochthonius sp. with single transverse suture (1TS) and parallel sides of the notogaster; 45: Eohypochthonius sp. with transversely divided genital plates in ventral view.



Figure 46-48; 46: Sphaerochthonius sp. (Oregon desert) with two transverse sutures (2TS; second difficult to see) and pectinate biramous setae (A); picture height = 250 μ ; 47: Nipponiella sp. with elongate minutely plumose setae and two prominent transverse sutures; 48: Cosmochthonius lanatus with surface sculpturing and detail; note that one prosomal seta has been removed to reveal integumental pattern below (S = sensillum); picture height = 125 μ .

..... 7

Eohypochthonius was infrequently found in the Andrews Forest, located in only 19 of the 52 sites and then not abundant. It seemed to favor dry sites but showed no preference for successional sere. Seasonal abundance paralleled that for total oribatids but with bimodal peaks and troughs of greater amplitude.

Hypochthonius rufulus Koch (600 μ) was present in 32 of the 52 sites but never exceeded 3-percent relative abundance, except in the xeric clearcut. Seasonal abundance followed the norm of total oribatids. Hypochthonius rufulus avoided oldgrowth communities and was more abundant in dry than in midmoisture or mesic sites.



Figure 49-50: 49-50: Hypochthonius rufulus with one transverse suture (TS), plumose sensillum (S) and a wide posterior, dorsal and lateral views; picture height $49 = 525 \mu$, picture height $50 = 575 \mu$.

7A Transverse opisthonotal sutures convergent, anterior one incomplete (fig. 51); body yellow; prosoma elongate, length more than twice the width; body widest posterior to middle; sensillum long and indistinctly pectinate Eniochthonius

Eniochthonius (=Hypochthoniella) minutissima Berlese (figs. 51 and 52; 350 μ) was infrequently encountered in the Andrews Forest but, when present, was frequently abundant (exceeds 3-percent relative abundance in 60 percent of the instances). Its distribution was limited to the dry tree sere and the mesic clearcut, hence revealing no definable pattern. Within these two sites, however, its frequency exceeded 90 percent. Seasonal abundance patterns were distinctive; it was abundant in early winter and infrequent in early summer relative to total oribatid populations. Populations still peaked bimodally in spring and dry summer.



Figure 51-52; 51-52: Eniochthonius minutissima with two transverse convergent notogastral sutures (TS) and apparently undivided genital sclerites (GS) dorsal and ventral views; picture height $51 = 400 \ \mu$, picture height $52 = 400 \ \mu$.

7B Transverse sutures parallel (fig. 53); body white, orange, or yellow; prosoma wider than long; body rather quadrate, sides subparallel; bothridium prominent with clubbed or spatulate sensillum; body smaller (<250 μ) Brachychthoniidae

Table 4 - Characteristics of genera in the family Brachychthoniidae

Suprapleu plate (fig.	es l			Sensillum ma	ergum Irgina setae	al Color	Notes
Liochthonius	Р	simple/ blade	clear	clavate/ emarginate	2	+transp.	
Neoliochthonius	С	simple	clear	capitate	2	transp.	
Verachthonius	E	-	clear	fusiform/ setulose	2	transp.	'
Synchthonius	Е	simple	weak	fusiform/ setulose	2	+transp.	rostral denticu- lation
Mixochthonius	PE	plumose	clear	fusiform/ setulose	2	transp.	
Poecilochthonius	CE	simple	strong	fusiform/ setulose	3	yellow	rostral denticu- lation
Neobrachychthonius	CE	simple	clear	fusiform/ setulose	2	yellow	
Brachychthonius	CE	simple	strong	fusiform/ setulose	3	yellow/ orange	anal setae simple
Sellnickochthonius	CEF	blade/ simple	strong	fusiform/ setulose	3	pale yellow	some rostral denticulation
Eobrachychthonius (CDEFP	-	scatt.		3	yellow	terga with

circles

29

marginal suture

Distinctions between genera of Brachychthoniidae involve the nearly invisible suprapleural plates (column 2 in Table 4). The plates are designated by their position relative to the terga (see schematic illustration; fig. 58): P = prosomal, C = anterior edge of tergum 1, D = middle of tergum 1, E = at first transverse suture, and F = at second transverse suture. Brachychthonius, Sellnickochthonius, and Poecilochthonius show an integumental design under the scanning electron microscope that is not readily apparent otherwise because they are so small; the other genera are much less conspicuously patterned.

Brachychthonius sp. (fig. 53; 130 μ) was a widespread and abundant species in our samples, encountered in 42 of the 52 sites. It was characterized by a mean density greater than 20 in 8 of the 12 communities, but its occurrence was not consistent (constancy values were greater than 0.50 in only four communities). Though not among the most abundant oribatids, it exceeded 10-percent relative abundance in the xeric clearcut. This species was absent from old growth and preferred the clearcut and shrub seres. It showed a strong preference for dry > midtypical > mesic conditions. Within-replicate abundance was highly variable. Seasonal abundance was bimodal as usual but with an extremely high peak in dry summer and fall (59 percent of total yearly abundance).

Liochthonius sp. (figs. 54 and 55; 140 μ) apparently preferred shrub and tree seres, primarily in midmoisture and mesic edaphic gradients; total density was very low. It was abundant in rotting log samples taken by Cline & Seastedt (unpub.) from old growth of the Cascade Range.

Sellnickochthonius (=Brachychochthonius) (fig. 56 and 57) was encountered once in the Andrews Forest rotten-log study (Cline & Seastedt, unpub.).

Most of these genera are thought to be widespread and transcontinental. Distinguishing genera within this group will not be possible without assistance from an oribatid specialist; most of the characters cited are visible only in slide-mounted specimens.



Figure 53-55; 53: Brachychthonius sp. with two transverse sutures (2TS) and faint integumental pattern, sensillum indicated with an (S); picture height = 175 μ ; 54: Liochthonius sp. with two transverse sutures (2TS), little or no integumental design and blade-like simple setae (A) (large object between sutures is an artifact); picture height = 150 μ ; 55: Liochthonius sp. note absence of suprapleural plates; picture height = 175 μ .


Figure 56-58; 56-57: Sellnickochthonius with very prominent integumental patterning and 3 pairs of stippled suprapleural plates; 58: Diagrammatic placement of suprapleural plates in the Brachychthoniidae.

SECTION B: Palaeosomata (=Bifemorata)

These primitive mites are small and distinct. They resemble weakly sclerotized acaridid mites more than their heavily pigmented oribatid relatives. These mites may be overlooked in samples containing many specimens.

Nearly all species may be recognized by the huge melanized dorsal setae. Generic designations are based largely on setal pattern, which is difficult to distinguish because the body does not remain rigid in slide mounts.

1A Dorsum of mite without melanized long setae; longest setae shorter than body; sensillum strap-shaped; minute and distinguishable from prostigmatid mites only on slides (fig. 59); 440µ Aphelacarus

Aphelacarus acarinus (Berlese) was found in a nearly decomposed log in the Andrews Forest.

- 1B Dorsum with large black setae; longest setae nearly as long or longer than body (figs. 60 to 64); sensillum long and filamentous
- 2A Large dorsal black setae nonplumose; anterodorsal end of prosoma pointed and projecting over cheliceral bases; anterior transverse row of setae on opisthosoma with four short setae between two huge setae; three claws on legs II to IV (fig. 60)..... Ctenacarus 2B Large dorsal black setae plumose 3
- 3A First transverse row of opisthosomal setae comprised of four short setae (on a single sclerite) between two huge plumose setae (equal to largest on dorsum) (fig. 62)..... Beklemishevia

Walter (1981) cites *Beklemishevia* from central California litter. *Palaeacarus*, differing in having two instead of three claws on legs II to IV might also occur in the Pacific Northwest and would be separable only on the basis of slide-mounted material (fig. 61).

3B First transverse row without huge sublateral plumose setae Acaronychus/Zachvatkinella

Both of these genera have been found near Andrews Forest. They were never more common than 0.1-percent relative abundance in these samples. Zachvatkinella sp. (300 to 350 μ ; fig. 63) was present in 10 of the 52 sample sites, scattered through the habitat types, but never more than three individuals per sample. It has also been taken in old growth from the nearby Salmon Creek drainage (Topic 1982). Acaronychus tragardhi Grandjean was encountered twice in rotten wood samples and only once in our litter samples; it was also collected in the Salmon Creek litter studies of Topic (1982) and Walter (1981) and found to be dominant in soils from the edge of the Great Basin at Sisters (Moldenke & Lattin, unpub. data). These genera are very similar; the key character that separates them is the location of the medial pair of the first transverse row of setae on the same sclerite in Acaronychus (fig. 64).



Figure 59-64; 59: Aphelacarus without thickened melanized setae; setae simple; 60: *Ctenacarus* with thickened non-plumose melanized setae; 61: *Palaeacarus* with thickened melanized plumose setae; 62: *Becklemishevia* with thickened melanized plumose setae; 63: *Zachvatkinella* with thickened melanized plumose setae; first transverse row of setae equal in size; 64: *Acaronychus* with thickened melanized plumose setae.

SECTION C: small transparent oribatids (unnatural grouping)

These genera are largely transparent, difficult to see, and easily confused with immatures of more advanced oribatids or adult Acarida or Prostigmata. The Palaeosomata, distinguished by large setae and more leg joints, and the Enarthronota, distinguished by a segmented opisthosoma, might initially be confused with this group.

The maturity of transparent mites is most easily distinguished by examination of the genital region; fully formed genital plates are found only in the adult.





Figure 65-68; 65: Gehypochthonius rhadamanthus with transverse notogastral suture and non-protruding lateroabdominal gland; 66: Parhypochthonius aphidinus with transverse notogastral suture and protruding latero-abdominal glands; 67-68: Eulohmannia ribagai with elongate body, adanal plates and U-shaped ventral groove (TG), ventral and dorsal views; picture height 67 = 525 μ , picture height 68 = 525 μ .

- 2A Posterolateral portion of opisthosoma with a prominent cannon-shaped projection (ca. 100 μ; fig. 66)..... Parhypochthonius
 2B Posterolateral portion of opisthosoma with latero-abdominal gland, but lacking raised projection (ca. 400 μ; fig. 65)..... Gehypochthonius

Gehypochthonius rhadamanthus Jacot and Parhypochthonius aphidinus Berlese represent our only native parhypochthonioid arthronotine genera. They differ from immature Epilohmannia by their generally smaller size (100 to 400 μ compared with 350 to 600 μ), a scarcely visible transverse opisthonotal suture, and generally plumose sensilla (clubbed in Epilohmannia nymphs). Both genera appear to feed on xylem and have been taken in heavily decomposed logs (Cline and Seastedt, unpub.). Gehypochthonius was found only in the mesic tree stage; total density was very low.

brown stain in the region of the latero-3A With conspicuous abdominal glands (centers and pores of glands usually transparent) Trhypochthonius/Trhypochthoniella

Trhypochthonius americanus Ewing (fig. 69; 300 to 400 μ) has large latero-abdominal glands, which resemble those of the dorsoventrally flattened tannish Perlohmannia. Trhypochthonius occurred too infrequently in our samples (8/312) for association with any habitat; it was abundant only in dry shrub collection of May, 1983. Trhypochthoniella differs in lacking the bothridium and sensillum.

- 3B Without contrastingly colored latero-abdominal glands 4
- unicolored (more or less transparent 4A Elongate, cylindric, without contrastingly yellowish) body (fig. 68); darkened latero-abdominal gland; adanal plates present; ventral portion of opisthosoma with large indistinct U-shaped sulcus (fig. 69); sensillum pectinate; opisthosoma with longitudinal rows of prominent transparent setae Eulohmannia

Eulohmannia ribagai (Berlese) (ca. 650 μ) was rarely encountered in Andrews Forest samples. Only in the tree-stage midmoisture sere was it present in even 29 percent of the samples, never attaining 1-percent relative abundance. It seemed abundant only in dry summer. Walter (unpub.) reports that it is a species of the mineral soil horizon, which matches our results from the Coast Range.

Robust species; often with cerotegument; notogaster white; prosoma 4B either white or brown; venter without U-shaped sulcus; sensillum not pectinate; adanal plates absent





Figure 69-71; 69: Trhypochthonius americanus dorsal view, note that the pore of the latero-abdominal gland (LAG) is visible but there is no halo from the brown pigmentation surrounding the pore in the scanning electron microscope; picture height = 400 μ ; 70-71; Camisia carrolli with longitudinal carinae on the notogaster and a weakly concave posterior margin between prominent tubercles (TUB); picture height 70 = 900 μ , picture height 71 = 925 μ .

5A Posterior margin of notogaster concave (fig. 70, 71, & 73); bothridium and sensillum present Camisia

Camisia carrolli Andre (figs. 70 and 71) was the most abundant and noticeable oribatid of the high canopy. Andre and Voegtlin (1981) have published on its biology; it is strictly arboreal, never appearing in litter samples. *Camisia biuris* (fig. 73) is occasionally encountered in conifer litter from Oregon and Washington; it has not yet been taken in the Andrews Forest. *Camisia segnis* (Hermann) is also recorded from western Oregon.

5B Posterior margin of notogaster convex (fig. 72, 74); bothridium absent; sensillum sometimes absent Malaconothrus/Trimalaconothrus

These two genera have been found widely throughout western Oregon and Washington. They are distinguished by the number of tarsal claws: one in *Malaconothrus monodactylus* (Michael) (fig. 74) and three in *Trimalaconothrus* simplex (Banks) (fig. 72). Both species occurred in the Andrews Forest, but no habitat association was noted. *Mucronothrus* may also occur in the Pacific Northwest; it is monodactylous but has 18 to 20 pairs of genital setae. The other two genera have only 4 to 12 pairs.







Figure 72-74; 72: Trimalaconothrus sp. note absence of bothridium and presence of ridged notogaster; 73: Camisia biuris with weak longitudinal carinae on the notogaster and strongly concave posterior margin; picture height = 1650 μ ; 74: Malaconothrus sp. also with absence of bothridium and presence of ridged notogaster.

SECTION D: Mixonomata (in part) + Mesoplophora, the penknife mites (=Ptyctimina of older classifications)

All species in this group can flex the prosoma into the antero-ventral portion of the opisthosoma to accomplish a turtle-like protective function (compare figures 7 and 8). Whether fully expanded or contracted, nearly all specimens lie on their sides, which distinguishes them from other preserved oribatids. Published reports and our studies implicate these ptyctimine species as wood and leaf feeders. Collectively, they comprise the greatest biomass of oribatids in tree-stage and old-growth communities. A most useful reference on this group is Walker (1964); however, the generic placement of nearly all species (except *Euphthiracarus*) has been changed subsequently in the catalogue (Marshall et al. 1986).

This couplet separates the Protoplophoroidea together with the genera related to *Mesoplophora* (Mesoplophoroidea of older classifications) from typical Mixonomata penknife mites. In *Protoplophora* (recorded by Walter from coastal central California), the notogaster is divided by a transverse suture. In *Mesoplophora* (recorded by Walter from western Oregon and California), the notogastral suture is ventrolateral and visible in ventral view (fig. 76) forming a V-shaped plate between the genital and anal sclerites and the wrap-around notogaster.

75



Figure 75-77; 75: Protoplophora sp. with transversely divided notogaster; 76: Mesoplophora sp. (Stinson Beach, CA.) with separate additional pair of peripheral ventral longitudinal sclerites (A) and separate complete rounded genital (GS) and anal plates (AS); notice that this specimen is immature evidenced by the incomplete development of genital sclerites; picture height = 225 μ ; 77: Mesoplophora sp. in lateral view; picture height = 275 μ . 2A Genital and anal plates broad, separated by a transverse furrow (figs. 78 to 80); both plates visible in profile as a convex belly crossed by a prominent belt-line (fig. 78, 80) (Phthiracaroidea) ...



Figure 78-80; 78: Hoplophorella sp. with distinctly pitted notogaster and venter with transverse suture (TS); 79-80: Phthiracarus sp. with protuberant ventral belly crossed by complete transverse suture (TS); picture height $79 = 925 \ \mu$, $80 = 825 \ \mu$.

All genera of Phthiracaroidea will key to this couplet. Balogh (1972) differentiates genera by the pattern of ventral setae, a character apparent only to the expert. *Hoplophorella* (fig. 78) and *Atropacarus* are distinguished by a distinctly reticulated notogaster; they have been collected by Walter (unpub.) in western Oregon. *Steganacarus* (in the strict sense) as cited in the catalog is probably erroneously recorded as widespread in North America. Of the smooth genera, *Hoplophthiracarus* (eastern half of the United States) has five pairs of anal setae and erect long lamellar setae; *Phthiracarus* (*Archiphthiracarus*) has five pairs of anal setae and decumbent lamellar setae; *Phthiracarus* (transcontinental) has three pairs of anal setae (with the other two pairs of sockets present) and decumbent lamellar setae.

Species distinctions in *Phthiracarus* (figs. 79 and 80) are difficult; the sizes of our native taxa are apparently highly variable. We have distinguished two groups: a large tan species (750 to 950 μ) and a smaller pinkish group (ca. 600 μ).

Both "species" of *Phthiracarus* were widespread and abundant within the research design; the large species occurred in 50 of the 52 sites, and the smaller group was found on 51. Both displayed the typical bimodal seasonal abundance curve, but in qboth the spring peak lasted into early summer (a season with few oribatids in the Andrews Forest). The larger species was found more in old growth than in tree seres and more in shrub than clearcut seres; it preferred dry conditions in tree and old-growth seres and moist sites in clearcut and shrub seres. The smaller taxon prefered old-growth and tree seres over shrub and clearcut, but demonstrated no moisture preference.

- 3A Large (>800 μ) and orange-tan; ventral region with two pairs (figs. 81 and 83) of longitudinal genito-anal plates (Oribotritiidae)

Oribotritia (=Plesiotritia) megale (Walker) (figs. 81 and 82; 1000 to 1500 μ) was widespread and abundant in Andrews Forest (48 of 52 sites) with constancy values less than 0.60 only in two clearcuts. It was not reported by Walter and Topic from the Salmon Creek drainage. Seasonal occurrence followed the typical bimodal pattern. This species preferred the shrub sere and avoided clearcuts. Density was twice as high under dry and midmoisture as under mesic conditions.



Figure 81-82; 81, 82: Oribotritia megale with double set of elongate genito-anal plates (GAP); oval body outline; prominent lateral carina (LC) on prodorsum; and complete genital/anal suture (GAS); picture height of 81 = 1300 μ , picture height of 82 = 1300 μ .

4B Size 400 to 1250 μ ; body elongate-oval (compare figs. 81 and 83) in dorsal view, length nearly twice width; with three pairs of long setae on prosoma; genito-anal plates with transverse suture extending less than half-way to lateral margin Maerkelotritia



Figure 83-84: 83, 84: *Maerkelotritia* with double pair of longitudinal genito-anal plates (GAP), an insignificant transverse genital/anal suture (A), inconspicuous lateral carina on prodorsum and long setae (ventral and lateral views); picture height 83 = 1300 μ , picture height 84 = 1600 μ .

Maerkelotritia (figs. 83 and 84) is represented by at least two species in this region: a larger species M. gibbera (Walker) (900 to 1300 μ) and at least one smaller one. Maerkelotritia gibbera was widespread but not very abundant, occurring in 47 of 52 sites but only in one of the 12 sites with a constancy of >0.75. Its sparse abundance concealed any distribution pattern.

Seasonally, the typical bimodal abundance pattern apparently held, but abundance was low in late winter. The other smaller (400 to 600 μ) species of *Maerkelotritia* occurred in only 16 of 52 sites; however, 5 of 12 times it had a constancy value greater than 0.75. It was twice as abundant as *M. gibbera*. Seasonal abundance was typically bimodal, with a high spring peak. The smaller species preferred shrub and tree seres of midmoisture and the tree sere of mesic sites. *Mesotritia* is distinguished by the lack of a carina on the prodorsum; it probably occurs in the Northwest.

5A Orange-tan, 275 to 350 μ ; prosoma without long erect dorsal setae; sensillum somewhat strap-like, broadened apically Microtritia

Microtritia (=Rhysotritia part) paeniminima (Walker) (fig. 85) was generally distributed through Andrews Forest. It was never abundant and therefore no distributional preference can be discerned. In only one of the 12 community types did it exceed a constancy value of 0.50. Seasonal abundance followed the typical bimodal pattern, but the spring peak and early summer trough were exaggerated.

5B White or off-white; size variable 6

6A Lateral borders of genito-anal plates outlined with melanization (not visible externally in scanning electron micrographs; fig. 86), appearing to be dark arcs highlighting the genito-anal plates in ventral view; genital and anal components of the longitudinal plates separated by a medial transverse fissure and adjacent triangular darkening; setae on prodorsum long; surface of dorsum often with distinct stippling Euphthiracarus

Synichotritia caroli Walker (400 to 800 μ) is probably abundant in the Coast Range of the Pacific Northwest (Walker 1964). It is very similar to Euphthiracarus but lacks the ventral triangular darkenings of the genito-anal plates. Many of the individuals seem to combine characteristics of both genera; this group needs further work to distinguish the species inhabiting Andrews Forest. Euphthiracarus tanythrix sierrensis Walker (500 to 600 μ) was widespread (51 of 52 sites), second only to the small Phthiracarus among ptyctimines in abundance, but more consistent (eighth of all oribatids--low only in clearcuts). Seasonality fit the typical bimodal pattern closely. Euphthiracarus tanythrix was nearly absent from clearcuts; it preferred tree seral stages over shrub or old growth and mesic habitats to xeric or midmoisture. At least two other species of Euphthiracarus are found in western Oregon.



Figure 85-87; 85: Microtritia paeniminima with single short pair of genito-anal plates (1GAP); picture height = 675 μ ; 86-87: Euphthiracarus sp. with a single pair of elongate longitudinal genito-anal plates, stippled notogaster and conspicuous genital/anal fissure (TS) ventral and lateral views); picture height 86 = 600 μ , picture height 87 = 1200 μ .

A second species of *Microtritia* is apparently abundant in the interior of rotted logs in the Andrews Forest (Cline & Seastedt, unpublished). *Rhysotritia* with 8 to 9 pairs of genital setae (four pairs in *Microtritia*) and trochanters III and IV bisetose (single in *Microtritia*) may also be expected in the region.



Figure 88: Euphthiracarus sp. showing broad flat rutella and the tips of robust pincher-like chelicerae; picture height = 240 μ .

SECTION E: galumnoid pterogasterines, winged mites with bomb-bay doors

These large, heavily pigmented mites with movable pteromorphs covering the entire leg region are a distinctive and rather uniform group. Genera are distinguished by ridges on the prodorsum and the condition of the dorsosejugal suture. These shapes are often difficult to see because of the dark pigmentation of most specimens.

1A Prosoma and opisthosoma separated by an uninterrupted dorsosejugal suture (fig. 89) Neoribates

Keying with *Neoribates* (cited by Walter from western Oregon) is the North American genus *Parakalumna*. The glabrous *Parakalumna* has pointed pteromorphs with curved anterior margins; *Neoribates* has pteromorphs with rounded anterior margins.

1B Prosoma and opisthosoma appear nearly continuous, without a continuous dorsosejugal suture fully separating them (fig. 90, 91)



Figure 89-91; 89: Neoribates with uninterrupted anterior border of the notogaster; 90: Pilogalumna without trace of dorsosejugal suture; 91: Acrogalumna with hint of dorsosejugal suture and carine posterolaterally on prodorsum.

2A With traces of an interrupted dorsosejugal suture between the prodorsum and notogaster; with a mental tectum, a shield covering the head region and obscuring the chelicerae in ventral view (fig. 94) Pergalumna/Galumna

Both *Pergalumna* and *Galumna* have been taken in the xeric shrub sere. Both are rare. *Pergalumna* is abundant arboreally on oak in the Willamette Valley. *Pergalumna* has been implicated in nematode predation, and may require a diet of nematodes for successful maturation (Rockett 1976).

2B Without traces of the dorsosejugal suture separating prodorsum and notogaster; without mental tectum Pilogalumna

Keying with *Pilogalumna* (transcontinental) is *Acrogalumna* (eastern United States). *Acrogalumna* has a distinct longitudinal line in the posterolateral region of the prosoma (fig. 91).



Figure 92-94; 92: *Pergalumna* sp. (Oregon Desert) in lateral view; picture height = 650 μ ; 93: *Pergalumna* sp. (Andrews Forest) with 2 pair of faint longitudinal lines on prosoma; picture height = 925 μ ; 94: *Pergalumna* second sp. ventral view showing mental tectum (MT); picture height = 650 μ .

SECTION F: spine-shouldered pterogasterines (Tenuialidae--Gustavioidea; Achipteriidae [without Anachipteria]--Achipterioidea)

This is an artificial grouping of two unrelated families of mites that have prominent, anteriorly directed flat broad spines arising from the shoulder region. The tenuialids are a uniform, species-poor group with genera differing primarily in the shape of the lamellae. The achipteriids are abundant and species rich; specific and even generic distinctions are usually difficult because most species are heavily melanized, and the arrangement and number of sacculi or pores is diagnostic.



Figure 95-97; 95: Achipteria sp. with large forward-pointing ("oxypters" - OX) pteromorphs (P), lateral outline of body not a smooth curve; picture height = 425 μ ; 96: Tenuiala nuda with slender forward-pointing pteromorphs (OX), lateral margin of body a smooth curve; picture height = 800 μ ; 97: Peltenuiala pacifica with wide forward-projecting pteromorphs (OX); picture height = 825 μ .

- 1A Without fully developed pteromorphs, anterolateral border of notogaster unmodified and a smooth curve; shoulder extension in the anterior direction only (figs. 99 and 100)...... 2
- 1B With fully developed pteromorphs, extending in part ventrally (fig. 98) Achipteria/Parachipteria

Achipteria, Pseudachipteria, and Parachipteria are distinguished by type and shapes of notogastral pores. These regions of lightened pigmentation are visible only through microscope slide mounts. Achipteria and Parachipteria are known to occur in western Oregon. Parachipteria has clearly visible compound "pores" (each apparently stippled, known as "areae porosae". Achipteria and Pseudachipteria have only minute pore-like structures, those of Achipteria are connected to tubular sacs. Walter (unpub.) records two species of Achipteria, A. oregonensis Ewing and A. borealis Ewing, in western Oregon.

Achipteria (figs. 95, 98 and 103; 460 to 500 μ) were found in all 52 sites (a feature shared only with Anachipteria, Oppiella, Ceratozetes, Quadroppia, Rhinosuctobelba, Suctobelbella, and Scheloribates). It was eighth in overall constancy and relative

abundance. Ubiquitous and numerous, it tended to avoid old growth and tree seres (nearly absent from xeric old growth), but remained independent of the moisture gradient. Several of the sites exhibited erratic year-to-year variability, but Achipteria usually paralleled Anachipteria abundance. It was most abundant in late winter and infrequent in early spring.

2A Body dorsoventrally flattened (fig. 99), outline nearly circular; aspect of mite similar to a discus; notogaster with prominent large setae (fig. 97) Peltenuiala

Peltenuiala pacifica Norton (figs. 97, 99, and 102; 730 to 800 μ) was common in Andrews Forest, occurred in 39 of 52 sites, but exceeded a constancy value of 0.50 only in midmoisture old growth. Seasonal abundance was clearly atypical, but without robust data from the spring, difficult to interpret. It preferred old growth to tree and tree to shrub seres; it was least abundant in drier communities.



Figure 98-100; 98: Achipteria with fully formed ventrally-encircling pteromorphs (P); picture height = 550 μ ; 99: Peltenuiala pacifica with a flattened body and broad forward-directed humeral projections (OX), but without pteromorphs; picture height = 1000 μ ; 100: Tenuiala nuda with convex body shape and broad forward-directed humeral projections (OX); picture height = 750 μ .

2B Notogaster normally convex (fig. 100); outline oblong-cylindric; notogaster without setae Tenuiala

Keying with *Tenuiala* is *Tenuialoides*, distinct in having narrow translamellae (*Tenuiala* has broad, acutely pointed lamellar teeth). *Tenuiala* nuda Ewing (figs. 96, 100, 101 and 104; 710 to 800 μ) was encountered in three of 52 community sites; it apparently preferred tree and old-growth sites.



Figure 101-103; 101: Tenuiala nuda with narrow pointed lamellae (L); picture height = 325 μ ; 102: Peltenuiala pacifica with narrow lamellae, widely separated (L); picture height = 325 μ ; 103: Achipteria with broad lamellae separated by only a line (L); picture height = 125 μ .



Figure 104: Tenuiala nuda ventral view to show the spined shoulder projections dorsal to the legs); picture height = 725 μ .

NOTE: Confusable with this group of spine-shouldered oribatids is the genus Anachipteria (in part); one abundant species has a well-developed pair of anterodorsal coverings of the legs ("pedotecta") in a similar position as the spined shoulder projections. These are separate structures and not to be confused with the humeral angle of the notogaster (fig. 121).

SECTION G: typical pterogasterines (Oribatelloidea, Ceratozetoidea, Phenopelopoidea, Achipterioidea)

This visually coherent group consists of genera that are difficult to distinguish once Sections E and F are removed. The Phenopelopoidea are easily recognized by their cerotegumentary patterning, though technical distinction is based on the modified shape of the chelicerae. The Microzetoidea though not expected in the Pacific Northwest, would key to this group. The Oribatelloidea (+Achipterioidea) and the Ceratozetoidea are distinguished by the relative development of their lamellae (covering entire prodorsum or thin carinate ridges, respectively) and general coloring (dark in oribatelloids, more translucent in most ceratozetoids).

The "eupterous" condition of the pteromorphs is diagnostic of several genera; this condition may cause confusion to the beginner. Eupterous oribatids are able to flex their pteromorphs to enclose their legs. This ability is readily apparent under the dissecting microscope; a prominent light-colored seam shows at the base of the pteromorph. The seam seldom shows in an electron micrograph unless the pteromorph is flexed.



Figure 105-107; 105: Anachipteria sp. with conspicuous messy cerotegument, broad lamellae (L) obscuring all of prosoma; picture height = 550 μ ; 106: Propelops canadensis with patterned cerotegument and broad anterior translamella (L); picture height = 400 μ ; 107: Eupelops sp. with patterned cerotegument, short translamella (L) and immense interlamellar setae (ILS); picture height = 725 μ .

Anachipteria sp. "CT" (in the Oribatelloidea; figs. 105 and 109; 570 μ) was occasionally encountered throughout the grid, but only once exceeded a constancy value of 0.50 (mesic old growth). It inhabits primarily clearcut and shrub seres, preferring the midmoisture to mesic communities. Seasonal abundance is bimodal, with a high peak in spring and a broader, dry summer-fall peak. It is uncommon in late winter. Topic and Walter (unpub.) did not encounter it in the Salmon Creek watershed. Two other species of Anachipteria were found in Andrews Forest; they key out on page 53.

- 3A Brown; cerotegument conspicuously warty, forming a uniform pattern; translamella anteriorly placed, obscuring nearly all of prodorsum in dorsal view (fig. 106); dorsum without modified setae; dorsosejugal margin of notogaster unmodified...... Propelops

Propelops canadensis (Hammer) (figs. 106, 110 and 113; 340 μ) was one of the most widespread taxa in the Andrews Forest, found at 49 of the 52 sites. It was not abundant (most common in the midmoisture shrub sere), and only twice exceeded a constancy value of 0.75. Habitat preference was clearcut and shrub relative to tree and old growth seres; it was more common and constant in midmoisture than either dry or mesic sites. Seasonal abundance paralleled the bimodal pattern for total oribatids; however, the fall peak began during late dry summer.

3B Black; lenticulus prominent; cerotegument variable, not forming a uniform pattern; translamellar bridge over median portion of prosoma, leaving anterior portion of prodorsum visible; anterior border of notogaster with small anteriorly projecting lobes covering interlamellar setal bases (fig. 112); interlamellar setae modified into lamella-like structures (fig. 112); notogaster often with modified setae Eupelops



Figure 108: Peloptulus sp. with translamella, unmodified interlamellar setae and long spatulate sensilla; picture height = 500 μ .



Figure 109-111; 109: Anachipteria sp. with cerotegument (CT) but without flexible pteromorphs (P); picture height = 425 μ ; 110: Propelops canadensis with cerotegument (CT) but without flexible pteromorphs; picture height = 325 μ ; 111: Eupelops sp. with cerotegument (CT) and flexible pteromorphs (PS); picture height = 525 μ .



Figure 112-115; 112: Eupelops sp. with translamella (T) close to notogaster and huge interlamellar setae (ILS) arising from basal tubercles; picture height = $350 \ \mu$; 113: Propelops canadensis in ventral view, note the typically formed chelicerae; picture height = $425 \ \mu$; 114: Eupelops sp. with attenuate chelicerae; picture height = $150 \ \mu$; 115: Liacarus sp. with typically formed chelate chelicerae of most oribatids; picture height = $400 \ \mu$.

Eupelops (figs. 107, 111 and 112; 560 μ) was infrequently encountered in Andrews Forest. It was found in 13 of 52 sites, most abundantly in the midmoisture shrub sere. It occurred too infrequently to discern any seasonal abundance pattern.

Peloptulus has been collected in the Andrews Forest by Parsons, but not in the prime research grid (one specimen only; fig. 108).



Figure 116-117; 116: Ceratozetes sp. lateral view, note presence of lamellae (L); picture height = 475 μ ; 117: Scheloribates sp. lateral view, lamella visible only as linear ridge (LR); picture height = 325 μ .



Figure 118-120; 118: Scheloribates arboreal species, note the thin lamellar ridge (LR); picture height = 300 μ ; 119: Anachipteria large species with broad lamellae (L) and prominent pedotecta; picture height = 250 μ ; 120: Oribatella large species, lateral view of prosoma showing lamellae (L) projecting antero-dorsally over head region; picture height = 350 μ .



Figure 121-123; 121: Anachipteria large species with wide pointed lamellae and prominent pedotecta (PT); picture height = 400 μ ; 122: Anachipteria smaller species with broadly pointed lamellae and inconspicuous pedotecta (P = pteromorph); picture height = 550 μ ; 123: Anachipteria smaller species with inconspicuous pedotecta in lateral view (P = pteromorph); picture height = 550 μ .



Figure 124-126; 124: Oribatella medium species showing the wide bidentate lamellar tips (LT) and the smooth outline of the lateral portions of the notogaster; picture height = 450 μ ; 125: Oribatella small species; note emarginate dorsosejugal suture (DSJ), basally divergent lamellae and filiform sensillum; picture height = 300 μ ; 126: Oribatella small species (small morph) with smaller pteromorphs, parallel interior borders of the lamellae and uninterrupted dorsosejugal suture; picture height = 125 μ .

Oribatella is a genus represented by at least five species of different sizes in western Oregon, with three or four species occurring in Andrews Forest. The most widespread and abundant was found at 51 of 52 sites but was only 18th in total abundance (fig. 126). Its size varied, with peaks at 340 μ and 290 μ ; whether these two size classes represent two cryptic species or sexual differences is not known. Seasonal abundance was constant throughout the year, with a small peak in fall and a small trough in early winter. This composite size-class was generally distributed, preferred dry to midmoisture or mesic conditions and evidenced a preference for old growth and tree seres over either shrub or clearcut. The second, midsized species (390 μ ; fig. 125) was encountered infrequently (20 of 52 sites). It is nearly restricted to shrub seral stages and preferred mesic to midmoisture conditions (absent in dry sites). The third and largest species (figs. 120 and 124; 570 μ) was rare and encountered primarily in the mesic shrub sere. It was exclusively a shrub species, preferring mesic to either midmoisture or xeric gradients. Seasonality data for the two larger species were similar, with a high peak in early summer but remaining infrequent from fall through late winter. (Parsons' samples were not taken in spring or dry summer in relevant sites.)

Adoribatella (recorded from Colorado) differs from Oribatella in having the lamellar emargination shallow and therefore the bidentate lamellar teeth tiny.

- 6A Lamellae consisting of distinct separate flaps, not covering whole prodorsum, not appearing shelf-like in lateral view; black with lightened lenticular region near middle of dorsosejugal suture; posterior border of pteromorphs not discernable in dorsal view, profile nearly a smooth curve Anachipteria (part)

Anachipteria was represented in Andrews Forest by at least two species; the larger was easily confusable with Achipteria (treated in the spine-winged pterogasterine section). This larger species (figs. 119 and 121; 420 μ), present in all 52 sites, had a mean abundance greater than 20 in every site. It was the eighth most abundant and the eighth most constant species. It showed a slight avoidance of old growth and mesic sites. The seasonal distribution pattern of both fit the typical pattern. The larger species can be distinguished by the pointed plate protecting the legs and arising from them (and therefore not part of the true pteromorph). The smaller species (figs. 122 and 123; 350 to 380 μ), present in 50 of 52 sites, was characterized by a mean abundance greater than 20 in 11 of the 12 communities, and is 11th in total abundance. It was 15th in constancy, preferred the drier end of the gradient and tree and old growth seral stages (nearly absent in clearcuts). The smaller species exhibited the same erratic abundance pattern as Achipteria. Achipteria was slightly more abundant than the large Anachipteria and nearly twice as abundant as the small Anachipteria. See page 48-49 for another species of Anachipteria.

The genus *Tectoribates* (=*Anoribatella*), distinguished by having the medial lamellar tooth the longest, is reported in the catalog from North America but has not been encountered in western Oregon.

7A Lamellae forming a continuous flat shelf obscuring entire prosoma (fig. 127), without medial split; posterior border of pteromorphs discernable in dorsal profile, interrupting a smoothly curved outline (fig. 127) Lepidozetes

Lepidozetes is known from western Oregon, but has yet to be collected in the central Cascade Mountains. Scutozetes, a genus found boreally across North America but not yet found in Oregon, may be distinguished by the height of the lamellae, which obscure only the medial half of the prosoma.

Keying in couplet 7B might be two additional genera with an elongate split between the translamellate lamellae: *Tegoribates* (Alaska to Colorado and Indiana) has broad, smoothly transverse lamellar tips and a typical dorsosejugal suture; *Dentizetes rudentiger* Hammer (Alberta) has a transverse dorsosejugal suture and broad multitoothed lamellar tips (fig. 130). All three genera are expected in the forested Cascade Mountains.





Figure 127-130; 127: Lepidozetes sp. with completely fused synlamellae; 128-129: Mycobates sp, note the translamella (TL); picture height $129 = 125 \ \mu$, picture height $130 = 250 \ \mu$; 130: Dentizetes with translamella (TL) and multiple-toothed lamellar cusps;

Jugatala tuberosa Ewing is an abundant arboreal species inhabiting the upper crown of old growth Douglas-fir. Voegtlin (1982) reports it only on bare branches, with an additional unidentified species inhabiting epiphytic growth and trunk surfaces; the second species lacks the posterior tubercules of J. tuberosa.



Figure 131-132; 131: Anachipteria small species with close-up of lamellae (L) and inconspicuous pedotecta; picture height = $325 \ \mu$; 132: Jugatala sp. showing near absence of lamellar structures (LR) in contrast to Figure 131; note the clubbed sensillum (S); picture height = $275 \ \mu$.



Figure 133-136; 133-134; Jugatala tuberosa, note posteromesal tubercular humps (TUB), very small pteromorphs (P) and clubbed sensillum (S); picture height 133 = 625 μ , picture height 134 = 525 μ ; 135-136: Scheloribates large species, dorsal and lateral views (LR = lamellar ridge, P = pteromorph); picture height 135 = 400 μ , picture height 136 = 450 μ .

Scheloribates (figs. 135 to 142) is a diverse and abundant genus throughout Oregon. In Andrews Forest, this genus is represented by at least four species.



Figure 137-142; 137-138: Scheloribates (Ft. Rock, Oregon) in dorsal and lateral views; picture height $137 = 275 \ \mu$, picture height $138 = 200 \ \mu$; 139: Scheloribates sp. (470 μ) ventral view of a typical oribatuloid; picture height = 475 μ ; 140: high magnification of same species showing lamellar ridge (LR) and sensillum (S) interlamellar seta broken off and partially covered by dirt); picture height = 225 μ ; 141-142: Scheloribates tear-shaped species in dorsal and lateral views; picture height 141 = 200 μ , picture height 142 = 300 μ .

One species is strictly arboreal, inhabiting the canopy of old growth Douglas-fir; Voegtlin (1982) reports it abundantly in epiphytic growth and on the bare bark of branchlets. At least two species were found in litter. The smaller (380 to 400 μ) ranked sixth in abundance and ninth in constancy, being found on all 52 sites. The smaller species preferred clearcut and shrub seres; it was almost entirely replaced by *Scheloribates* sp. "S" in old growth. It preferred dry sites. Seasonal abundance closely corresponded to that for total oribatids. The larger species (460 to 480 μ) was almost as abundant, found on all but one of 52 sites, and ranked seventh in abundance. Constancy values exceeded 0.80 in 8 of the 12 plant communities. It preferred the shrub sere (avoided clearcuts less than tree or old growth communities) in mesic or midmoisture conditions. Seasonal abundance seemed rather uniform (with one trough in early winter) rather than bimodal. A fourth species (large, tear-drop shaped) (figs. 141 and 142; 280 μ) occurred only in the three old growth communities and the midmoisture tree site.

Also keying in couplet 10A is *Oripoda*, which is known from western Oregon (Walter unpub.). *Oripoda* is distinguished by a cylindric body (fig. 226) (width less than half the length) with the posterior portion of the parallel-sided prosoma only half to one third the width of the anterior portion of the notogaster.

- 10B Pteromorphs separated from notogaster by a lightly colored sulcus; lamellae present as linear ridges on sides of prodorsum; dorsosejugal suture strongly convex 11
- 11A Anterior margin of notogaster nearly transverse; poronotic; monodactylous Xylobates

Xylobates has been collected by Walter (unpub.) in western Oregon.

Keying with *Haplozetes* (fig. 143) is *Peloribates* (fig. 146), a genus reported transcontinentally from North America; it may be distinguished by 14 pairs of long notogastral setae (10 pairs in *Haplozetes* and *Xylobates*). Both genera have been collected in western Oregon by Walter (unpub.).

12A Pteromorphs separated from notogaster by true sulcus of flexure (a lightly colored line at the base of the pteromorph) 13 12B Pteromorphs not separated from notogaster by sulcus of flexure...14

13A Translamellate (figs. 128 and 129) Mycobates

Mycobates has been collected by Walter (unpub.) in western Oregon and coastal central California (Walter 1985), but has not been encountered in Andrews Forest. Minuthozetes, reported from central California by Walter (1985), is distinguished by a single tarsal claw, long spindle-shaped sensilla, and a nearly straight anterior margin of the notogaster.

13B Lamellate, without translamellar ridge Diapterobates

Though not recorded from Andrews Forest, a species apparently keying to this genus has been found by Walter (unpub.) in western Oregon; the catalog reports it transcontinentally across Canada and the northern United States. Also keying here are two other distinctive genera recorded from North America by Balogh (1972) but not yet seen in Oregon. Neither genus has notogastral setae and hence can be distinguished from *Diapterobates*. *Pelopsis* (*=Parapelops*) has highly modified, flattened interlamellar setae that are apically bifurcate; it is widely distributed from the Northwest Territories to Texas. In *Punctoribates*, the dorsosejugal suture is modified and projects over nearly all of the prodorsum as a two-toothed shelf; this genus is also widely distributed from Alaska to Virginia.



Figure 143-145; 143: Haplozetes without translamella, note that the pale line of flexion at the base of the pteromorph is not visible in SEM; picture height = 500 μ ; 144-145: Sphaerozetes sp. in dorsal view, notice the translamella (TL) with apically notched teeth; picture height 144 = 225 μ , picture height 145 = 375 μ .



Figure 146: *Peloribates* without translamella but with prominent notogastral setae; 147: *Limnozetes* with a partial translamella and integumental patterning.

15A Translamella much wider than its flanking teeth Sphaerozetes

Sphaerozetes occurred frequently in Andrews Forest. It has three teeth on the tip of the rostrum, which distinguishes it from most similar ceratozetoid genera. Sphaerozetes (figs. 144, 145 and 148) preferred the mesic gradient (shrub, tree, and old growth) and xeric and midmoisture old growth.

Keying to this couplet is the aquatic genus *Limnozetes*. Unlike the smooth notogaster of *Sphaerozetes*, it has a stippled dorsum (fig. 147).

15B Translamella shorter than or equal to flanking teeth 16

Both genera occurred in Andrews Forest. *Ceratozetes* is very diverse and will be discussed below, where most of the species will appear to key out. *Trichoribates lamellata* (Ewing) is similar to *Ceratozetes*, but has an additional pair of notogastral setae (the subapical median pair), lamellae with two teeth at the tips and a minute capitate sensillum. *Trichoribates* was found only twice on the research grid.

Zachvatkinibates (coastal northern California) has dentate lamellar cusps and an elongate clavate sensillum; it is missing the subapical median notogastral seta but has the anteromedial setae. *Ceratozetes* has a filiform sensillum. *Podoribates*, widespread through North America is apparently glabrous and has a very short clavate sensillum.

16B Humeral region not formed into a small, yet conspicuous, obtuse angle, forming a gentle curve throughout; two anterior pairs of long dorsomedial notogastral setae present Melanozetes/Fuscozetes

These similar genera are reported from western Oregon by Walter (unpub.), but only one has so far been found in the Andrews Forest. The translamella in *Fuscozetes* is ribbon-shaped and short; in *Melanozetes*, it is linear. *Melanozetes* (figs. 149 and 150) was an infrequent species; it occurred throughout all tree stages and generally along the xeric gradient.

17A Notogaster apparently without setae; obtusely angled humeral regions visible in dorsal view Ceratozetes (part)

1.4	Sensillum clubbed2
1B	Sensillum fusiform or filiform setulose3
2A	Lamellar setae arising actually on prodorsum; rostral setae
	setulose; bothridium visible; transcontinentalChamobates
2B	Lamellar setae arising at tip of lamellae; rostral setae
	simple; bothridia hidden from dorsal view; transcontinental,
	reported by Walter from western Oregon
3A	Sensillum short-fusiform; rostrum pointed; from Florida and
	QuebecEuzetes
3B	Sensillum elongate filiform setulose4
4A	Rostrum with three tiny teeth anteriorly; legs tridactylous;
	notogaster poronotic; transcontinental, abundant in Oregon
	Ceratozetes
4B	Rostrum singly pointed anteriorly; first pair of legs
	monodactyl, others tridactyl; notogaster without pores;
	northeastern United StatesHeterozetes



Figure 148-150; 148: Sphaerozetes sp. with translamella (TL); picture height = 300 μ ; 149: Melanozetes sp. with translamella (TL) and squared off humeral region; picture height = 550 μ ; 150: Melanozetes sp. with translamella (TL); (DSJ = dorsosejugal suture); pw = 225 μ .

17B Notogaster with setae Ceratozetes/Hypozetes

These genera are superficially similar. *Hypozetes* (fig. 156) is sacculonotic; it has 10 pairs of notogastral setae and a clubbed sensillum. Voegtlin (1982) reports that it is abundant on lichens and moss in the high crown of old-growth Douglas-fir. *Ceratozetes* is poronotic (fig. 155); it has up to 11 pairs of notogastral setae (varying to glabrous) and a filiform sensillum. *Ceratozetes* is diverse in western Oregon; at least five species were found in these samples.

The most abundant *Ceratozetes* (380 μ ; figs. 151 and 152) is present in all 52 samples, and was sixth or seventh in relative abundance and constancy. Constancy exceeded 0.80 in all communities except the depauperate xeric clearcut. The seasonal abundance pattern followed the norm. The species preferred mesic more than midmoisture more than xeric sites and showed only a slight avoidance of clearcuts. A second species (310 μ ; fig. 153) was also widespread and abundant; it was found in 42 of 52 sites and was 14th in total abundance. This species preferred xeric over either midmoisture or mesic sites. In clearcut and shrub seres, it occurred only in dry areas; in the tree sere, it inhabited the dry and midmoisture sequence. In old growth, it occurred abundantly in all three moisture regimes. Seasonal abundance was atypical; it was trimodally high in early summer, fall, and late winter, and least abundant in dry summer.

A third species of *Ceratozetes* (460 μ) occurred infrequently in these samples. It was abundant in xeric clearcut sites and occasional in other dry or shrub sites. It was abundant in late winter and early summer (no data available for spring), with the usual troughs in dry summer and early winter. A fourth species (perhaps two) (530-630 μ) occurred infrequently (in 23 of 52 sites), but never in a constancy over 0.50. It also was found principally on xeric clearcut sites, and occasionally in other dry sites or tree seres. The seasonal abundance was apparently normal (exceptionally low in late winter), but no data are available from fall. A fourth species of *Ceratozetes* (tear-drop shaped; fig. 154; 330 μ) occurred only in shrub communities of midmoisture and mesic sequences. A fifth species (minute; fig. 155; 160 μ) was collected only twice in the research grid. The electron micrographs show prominent areae porosae on the notogaster but this may be an artifact of a newly moulted specimen.

The similar genus from boreal North America, *Iugoribates*, may be distinguished by a nearly transverse dorsosejugal suture. Similar to *Ceratozetes* in having a strongly convex dorsosejugal suture but with clubbed sensilla is the genus *Zetomimus* (known from New York).



Figure 151-156; 151-152: common 380 μ Ceratozetes species, note the elongate shape and long lamellar teeth (LT); picture height 151 = 350 μ , picture height 152 = 175 μ ; 153: smaller 310 μ species with the short lamellar teeth (LT); picture height = 200 μ ; 154: the tear-shaped species; picture height = 250 μ ; 155: tiny species with prominent areae porosae (AP); picture height = 275 μ ; 156: Hypozetes sp. high magnification of prodorsum showing the lamellar teeth (LT) and the sensillum (S); picture height = 250 μ .

SECTION H: Plateremaeoidea (=Gymnodamaeoidea), the flat-backed stilt mites



Figure 157-159; 157: Odontodamaeus veriornatus with elongate sensillum, extremely long legs and honeycombed notogaster; picture width = 1560 μ ; 158: Odontodamaeus veriornatus highly magnified to show pattern of prodorsal ridges (circular central ridge and semi-circular ridge between bothridium and interlamellar seta) and tubercle bearing the interlamellar seta (S = sensillum); picture height = 425 μ ; 159: Licnodamaeus sp. with leaf-like sensillum (S) and uniformly pitted notogaster.

- 2A Sensillum leaf-like; notogaster with surface evenly pitted; often elongate, with length more than twice the width; California (fig. 159) Licnodamaeus
 2B Sensillum clavate; widespread Plateremaeus

- 4A With prominent folds along posterior margin of notogaster and tubercles on the anterior portion of notogaster Pleodamaeus
 4B Without prominent folds on posterior margin of notogaster 5



Figure 160-161; 160: Gymnodamaeus sp. from the eastern deserts of Oregon with curved interlamellar ridges, interlamellar tubercles and extremely long legs; picture height = 1075 μ ; 161: Odontodamaeus veriornatus in ventral close-up showing the toothed border of the genital plates (GS) and the independent genital and anal apertures; picture height = 325 μ .

Odontodamaeus veriornatus Hammer is a dark brown species with a honeycombed notogaster that appears prominently under a dissecting microscope (780 μ). This is probably an arboreal species, recorded by Voegtlin (1982) in high frequency in lichens and mosses of the canopy of old-growth Douglas-fir. It occurred in 38 of 52 sample sites but in very low abundance; the fact that it occurs on the ground at all makes it unusual among the taxa studied by Voegtlin. On the ground, it was most abundant in midmoisture shrub and tree communities and mesic old growth. Only one seasonal abundance peak occurred in dry summer and fall; populations were lowest in early winter. The partial advance of the fall peak is a feature shared with Eremaeus and Oribotritia.

7A Notogaster longitudinally folded; six pairs of genital setae; 280 μ; Washington to Mojave Desert (fig. 163) Joshuella

We have found *Joshuella striata* in eastern Oregon and on a rotted log in the Coast Range. It has not yet been found in the Cascades.

Jacotella sp. is a small yellowish species (fig. 162; 280 μ) with no discernable pattern on the dorsum under the dissecting microscope (though one was readily apparent in the electron micrograph). It was widespread and abundant, occuring in 45 of 52 sites but most abundantly in the xeric shrub community. It demonstrated a strong preference for xeric over midmoisture over mesic conditions. It was primarily a species of the clearcut, which it preferred to shrub sites, while almost completely avoiding tree and old-growth sites. Seasonal abundance followed the same pattern outlined for Odontodamaeus.

Nortonella is a closely related genus known from the western United States and Canada. It differs from *Jacotella* in having only two pairs of adamal setae (three pairs in *Jacotella*).



Figure 162-163; 162: Jacotella sp. with honeycomb notogastral pattern, presence of bothridial ridge (BR), and bow-tie posterior setae; picture height = 425 μ ; 163: Joshuella striata with longitudinal folds on the notogaster, presence of a bothridial ridge (BR) and bow-tie posterior setae; picture height = 250 μ .

SECTION I: Perlohmannoidea and Epilohamnnioidea

Two species of *Perlohmannia* (figs. 164 and 166) are found in Andrews Forest (neither reported by Walter and Topic from Salmon Creek). One species (860 μ) occurred primarily in scattered samples of localized abundance. It was found in 37 of 52 sites, with a preference for mesic conditions. Though widely distributed, it avoided clearcut sites, preferring shrub and tree seres. Samples were not available from fall or late winter, but abundance was apparently low from early summer to fall and high in early winter and spring. The smaller species of *Perlohmannia* (650 μ) was found in 15 of 52 sites, but is essentially restricted to xeric and mesic shrub sites. Seasonal abundance was high from early winter to spring, low in early through dry summer.



Figure 164-166: Perlohmannia sp.: note the pectinate sensillum (S), dorsoventrally flattened aspect in lateral view, contiguous genital (GS) and anal plates (AS), peri-anal plates (AAS) and latero-abdominal gland (LAG) pore; picture height $164 = 1175 \ \mu$, picture height $165 = 950 \ \mu$, picture height $166 = 525 \ \mu$.

Epilohmannia n.sp. (figs. 167 to 169; 775 to 900 μ) was widespread and abundant in these samples, but exceeded a constancy value of 0.75 only in mesic tree and old-growth sites. Cline and Seastedt (unpub.) have identified numerous immatures in rotted wood samples from the Andrews Forest. It was primarily distributed in shrub, tree, and old-growth seres of mesic conditions; it also occurred in mesic clearcut and other tree and old-growth communities. Seasonal abundance peaked in fall and late winter with a low in spring (atypical!).



Figure 167-169: Epilohmannia sp.: notice the elongate notogaster, transverse ventral fold (TVF) between the genital and anal sclerites, the closely adjacent genital and anal plates, absence of adamal plates; picture height 167 = 1150 μ , picture height 168 = 975 μ , picture height 169 = 600 μ .
SECTION J: Hermannielloidea

The only genus inhabiting northwestern North America is *Hermanniella* (figs. 170 to 172). The erect cannon-like pores of the latero-abdominal glands are a character shared only with the diminutive transparent *Parhypochthonius*.

Though not abundant enough to rank among the top 15 most common oribatids of Andrews Forest, the commonest species of *Hermanniella* was found at 51 of 52 sites with a constancy value over 0.75 in all tree and old-growth sites. This species (525 μ) preferred old-growth to tree seres, which it favored over shrub and clearcut sites. It showed no preference relative to moisture gradient. The pattern of seasonal abundance closely paralleled that of total oribatids.

Several other species have been found in Oregon, often rather abundant where they occur (*H. occidentalis* Ewing; *H. robusta* Ewing cited by Walter in unpublished data). Two other species have been found in Andrews Forest: 1) a very infrequent, smaller taxon (450 μ) favoring mesic shrub and tree stages, and 2) a widespread, infrequent, larger species (700 μ), which apparently preferred xeric old growth, tree and old-growth seres in the midmoisture regime, and mesic shrub sites.



Figure 170-172: Hermanniella, note the protruding latero-abdominal gland (LAG) and large genital (GS) and anal sclerites (AS) in close proximity. The partial remains of the tritonymphal exuvia is visible in Fig. 172 under SEM; under a dissecting microscope this shed skin is not noticeable though it is carried by all species; picture height $170 = 700 \ \mu$, picture height $171 = 625 \ \mu$, picture height $172 = 550 \ \mu$.

SECTION K: Crotonioidea (=Nothroidea), Liodoidea

Most of the species in this group can be recognized by their large size, unusual body profile, and intricate surface sculpturing.

1A Adults carrying shed skins of immatures plastered onto dorsal surface (fig. 173); genital plate fully transversely divided (fig. 174); dorsum flat (fig. 175)(Liodoidea) Platyliodes

Other liodid genera recorded from North America include: *Telioliodes* (Florida) with exuviae with large, apical, posteriorly directed setae (nearly half as long as notogaster) and with lamellar setae; *Liodes* (all eastern United States) without lamellar setae or apical exuvial setae but with an irregularly wrinkled notogastral pattern; and *Poroliodes* (New York) without lamellar setae or apical exuvial setae but with a uniformly punctate dorsum.

Platyliodes macroprionus Woolley & Higgins (figs. 173 to 175; 750 μ) is a strictly arboreal species encountered only once in these samples. Voegtlin (1982) found all stages in high abundance on bare branches, lichens, and mosses in the high canopy of old growth.



Figure 173-175: Platyliodes macroprionus in dorsal, ventral and lateral views, note the transversely divided genital plates (GS) and the nymphal exuviae (NSS); picture height 173 = 1125 μ , picture height 174 = 1100 μ , picture height 175 = 1100 μ .



Figure 176-178: 176: *Liodes* sp. showing the integumentary pattern; 177-178: *Camisia spiniger* in dorsal and lateral views, note the concave posterior margin of the opisthosoma between the large posterolateral setae arising from prominent tubercles (TUB); picture height 177 = 1525 μ , picture height 178 = 1500 μ .

2A Entire body covered with dense cerotegument and dirt (figs. 177 to 178) Camisia (part)

Camisia spiniger (Hermann) (figs. 177 and 178; 875 to 900 μ) is apparently not primarily a soil litter species, though it was infrequently encountered there (22 of 52 sites, but never with a within-site constancy over 0.50); haphazard collecting has found it on tree trunks on several occasions. It was too infrequent in our samples to yield clear seasonality curves or plant-succession associations. Additional species of *Camisia* key out in Section C, page 36.

2B Body not covered with dense cerotegument and adhering dirt..... 3

3A Dorsal surface densely pitted; posterolateral setae much more elongate than rest of notogastral setae (fig. 179) Nothrus

Nothrus silvestris Nicolet (Fig. 179-182; 1000 μ) was a common mite in these samples. Though present in only 32 sites, and only in 4 of 12 communities with a constancy value exceeding 0.50, its large size and distinctive appearance render it one of the most familiar oribatids of the Andrews. Seasonal abundance followed the curve for total oribatids. Though no moisture preference was apparent, it showed a strong preference for tree and shrub stages (nearly absent in clearcut and old growth).

3B Bothridium present; without transverse ridge posteriorly (900 to 950 μ) Platynothrus



Figure 179-181: Nothrus silvestris in dorsal, lateral and high resolution views to show surface ornamentation, the convex posterior border of the opisthosoma and the bothridium; picture height $179 = 1525 \ \mu$, picture height $180 = 1000 \ \mu$, picture height $181 = 525 \ \mu$.



Figure 182: Nothrus silvestris in ventral view to show the large adjacent genital plates (GS) and anal plates (AS); picture height = 600 μ .

Platynothrus banksi (Michael) and P. peltifer (Koch) are common species in the mountains of western Oregon, but neither has been found in Andrews Forest. Neonothrus, known from Alaska, has prominent bothridia and an interrupted transverse V-shaped ridge; the sides of the notogaster are more nearly parallel and the dorsal setae more prominently blade-shaped. Heminothrus (s. str.) is known throughout the eastern United States and Canada; the median portion of the notogaster is somewhat concave and the posteromarginal setae arise on distinct apophyses (merely tuberculate in Platynothrus).



Figure 183-185: *Platynothrus banksi* in dorsal, lateral and ventral views, note that the genital plate (GS) is not divided as it is in the Liodoidea; picture height $183 = 875 \ \mu$, picture height $184 = 875 \ \mu$, picture height $185 = 875 \ \mu$. Note the extended ovipositor in Fig. 184.



Figure 186-189: Feeding appendages with typical pincher-like chelicerae (compare to *Eupelops* and *Rhinosuctobelba*) but divergently shaped rutella (chisel-like structures forming floor of mouth region); 186: *Nothrus* sp.; picture height = 150 μ ; 187: *Epilohmannia* sp.; picture height = 150 μ ; 188: *Perlohmannia* sp.; picture height = 250 μ ; 189: *Scapheremaeus* sp.; picture height = 70 μ .

SECTION L: Damaeoidea (=Belboidea)

The hemispheric notogaster, the beaded legs and eight pairs of longitudinally arranged notogastral setae are diagnostic for this superfamily. Generic distinctions are not often reflected in overall body form; published keys rely on the setal arrangement on the genu and tibial leg segments. *Quatrobelba* is not included in the key; it is definitely found in the Pacific Northwest but the authors do not know how to distinguish it.



Figure 190-192; 190: New genus #1 (near Damaeus) in dorsal view (Olympic Mts., Washington); picture height = 3500 μ ; 191: same species in lateral view with extremely long legs (A) and nymphal shed skins (NSS) glued to notogaster; picture height = 525 μ ; 192: Caenobelba sp. with cerotegument and dirt removed to show absence of spinae adnatae; picture height = 1000 μ .

This species has been collected occasionally in the mountains of Washington, but has not yet been found in the Oregon Cascades. It differs in setal characters from the closely related *Damaeus*, expected from this region.

1B Legs not much exceeding body length 2

3A With a distinct thoracic protuberance projecting between the base of the first two pairs of legs Epidamaeus(Akrodamaeus)(part)

Epidamaeus (Akrodamaeus) has cerotegument produced in filamentous columns on the notogaster (fig. 201); *Dyobelba* has a granular cerotegument that does not impair the surface sheen of the notogaster. The rostrum is prolonged into a snout in *Dyobelba*, which Walter (unpub.) has collected in western Oregon.



Figure 193-195: Belba californica displaying spinae adnatae (SA), filiform sensillum (S) and usual presence of organic debris on the notogaster; picture height 193 = 1750 μ , picture height 194 = 1400 μ , picture height 195 = 480 μ .

Caenobelba probably comprises three different species in Andrews Forest. The midsized species (figs. 192 and 204; 500 to 525 μ) was by far the most abundant and widespread; a larger (625 to 765 μ) and a smaller (300 μ) species were also found throughout the grid. No preference for successional sere or for a moisture regime was apparent for the genus as a whole or for any of its constituent size classes. These oribatids were distributed in all 52 sites, with a constancy value exceeding 0.90 in all but five. Seasonal abundance followed the oribatid norm, with low density in late winter.

Belba californica Norton (figs. 193 to 195; 800 μ) has not been encountered in these samples, but occurred in great numbers in a midsummer sample from a dry streambottom from MacRae Creek in Andrews Forest. It is frequent in litter samples from coastal Marys Peak (Oregon) and the coastal mountains of Washington.

Hungarobelba sp. (fig. 196; 300 to 325 μ) occurred in 50 sample sites but was never abundant (constancy value never exceeded 0.75). Seasonal occurrence followed that for total oribatids, but the peaks and troughs were of greater amplitude. This species apparently preferred shrub seres of any moisture regime and the tree sere under midmoisture conditions.

Dasybelba (figs. 197 and 198) known from the Great Basin of Oregon and Colorado is distinguished by the densely plumose setae on the legs. Different species apparently may or may not carry nymphal shed skins as adults.



Figure 196-198; 196: Hungarobelba sp. with ovoid body outline, large spinae adnatae (SA), beaded legs emphasized by dense covering of cerotegument on setae and lack of nymphal shed skins; picture height = 450 μ ; 197-198: Dasybelba sp. with clavate sensillum and densely plumose leg setae (Fort Rock, Oregon); dorsal view and high resolution of dorsosejugal region; picture height 197 = 475 μ , picture height 198 = 50 μ .

A large $(370 \ \mu)$ species was infrequent in the samples; it occurred in only 18 sites. Though it apparently preferred old-growth and tree seres to shrub (which it preferred to clearcuts), it was completely absent from the midmoisture old-growth site. Seasonal abundance seemed unimodal in spring and reduced through winter. This species apparently represents an undescribed genus.

Basilobelba (in the Amerobelboidea) may also key to this couplet (fig. 269). As in Epidamaeus, Basilobelba carries nymphal exuviae on its dorsum, but differs in having much less conspicuous cerotegument, no spiny adnatae, inconspicuous notogastral setae, and a ventral plate with 17 pairs of setae (four pairs in damaeoids).

6B Adult without covering of nymphal shed skins

••••••• Epidamaeus(Epidamaeus)

Epidamaeus (fig. 199-201) occurred in all but one site. The possible two or three species were never abundant, with a constancy value exceeding 0.75 only twice. They revealed no habitat or moisture preference. Seasonal abundance was low in early through late winter.

200 199 201 203 204 202 NSS

Figure 199-204; 199-200: Epidamaeus desert species (Ft. Rock, OR) in dorsal and lateral view (SA= spinae adnatae); picture height 199 = 275 μ , picture height 200 = 325 μ ; 201: Epidamaeus Andrews Forest species with some cerotegument removed to show dorsosejugal region; picture height = 510 μ ; 202-203: New genus #2 showing nymphal exuvial pattern (NSS) and dense cerotegument on all leg surfaces, dorsal and lateral views; picture height 202 = 500 μ , picture height 203 = 475 μ ; 204: Caenobelba sp. in lateral view showing position and extent of dirt and cerotegument (A); picture height = 600 μ .

SECTION M: Gustavioidea (=Liacaroidea)

This group is rich in number of genera, but the uniformity of body shape combined with the prominent lamellae prevent confusion with any other section of the key. Genera and species are distinguished largely by the shape of the lamellar teeth, easily visible on all but the most melanized species.

1A Lamellae thin and projecting all the way from their bases in an anterodorsal plane, like horns of a ceratopsian dinosaur (figs. 205 to 207) Ceratoppia

Ceratoppia was represented in these samples by three species of different sizes. The midsized species (500 to 600 μ) was most abundant, present in 48 sites. Though its constancy value exceeded 0.50 in all communities except the xeric clearcut, it was never abundant relative to total oribatids. Its seasonal abundance was rather uniform because of a high value during early summer, but it retained the typical early winter trough. It preferred tree and old-growth stages independent of moisture regime.

The large species (650 to 800 μ) was infrequent, present in only 18 sites. It showed a strong preference for old growth and was absent from clearcuts. It preferred mesic to midmoisture conditions and was nearly absent in the xeric seres. The smallest species (380 to 480 μ) was infrequent, present in only 37 sites, and exceeded constancy values of 0.50 in only three communities. Seasonal abundance was typical but for a higher than normal value in spring. Habitat choice was unclear; it seemed scattered, avoiding clearcuts and preferring xeric over mid- or mesic moisture conditions.



Figure 205-207: Ceratoppia sp. with the slender long raised lamellae (L) extending antero-dorsally far over the plane of the prosoma, note the filiform sensillum; picture height 205 = 350 μ , picture height 206 = 750 μ , picture height 207 = 750 μ .

The genus *Parapyroppia*, recorded by Walter (unpub.) from western Oregon, and *Pyroppia*, cited in the catalog from Alaska and Quebec, will also key to this couplet; they differ in the presence of a short dilated sensillum (S).

1B Lamellae not projecting freely anterodorsally like dinosaur horns



Figure 208-213; 208 & 210: Metrioppia oregonensis dorsal views, note convergent thin lamellae (L) and sensillum (S); picture height $208 = 675 \ \mu$, picture height $210 = 300 \ \mu$; 209, 211 & 213: Gustavia sp. sagittal, dorsal and lateral views, note prodorsum is mostly tucked in under notogaster and is not visible in dorsal view (R = rostrum; L= lamella); picture height $209 = 575 \ \mu$, picture height $211 = 1000 \ \mu$, picture height $213 = 750 \ \mu$; 212: Kodiakella sp. with thin sigmoid lamellae (L).

Metrioppia oregonensis Woolley & Higgins (figs. 208 and 210; 270 μ) was widespread and abundant in the grid. It was 11th in total density and 7th in constancy. Seasonally, its abundance followed that for total oribatids. This species strongly prefers tree and shrub successional seres over either clearcut or old growth. Abundance was much greater in mesic than in xeric or midmoisture sites. Metrioppia has an elongate pectinate sensillum.

With a tear-drop profile--and thus keying with *Metrioppia*--are the North American genera *Conoppia* and *Metapyroppia*, not yet collected in Oregon. *Metapyroppia* (Tennessee) is extremely robust, with the width equaling or exceeding body length; sensillum is fusiform. *Conoppia* (Northwest Territories and West Virginia) is an unrelated but similar-appearing cepheid; it is translamellate (teeth much shorter than interlamellar distance) and has prominent long interlamellar setae (reduced in *Metrioppia*).

Stonyxenillus is translamellate, with broad bidentate cusps; lamellae obscuring entire prodorsum (fig. 278). Adoristes and Kodiakella (fig. 212) have slender, simple lamellae not obscuring much of the prodorsum; the lamellae of Kodiakella are weakly S-shaped. Kodiakella is definitely known from Alaska through northern California.

2B Lamellae with a conspicuous translamellar ridge or synlamellate



Figure 214-216; 214-215: *Liacarus* near *robustus* with very long translamellar tooth (TT) and slender double-pointed cusps in dorsal and lateral views; picture height 214 = 1800 μ , picture height 215 = 850 μ ; 216: *Liacarus* small tan species with broad lamellar cusps (LT) and no medial tooth; picture height = 750 μ .

3A Tear-drop shaped (fig. 211); notogastral setae absent; prosoma directed ventrally, nearly hidden in dorsal view (fig. 211); black without a light-colored lenticulus; chelicerae not the typical lobster-claw shape, but elongate and tapering Gustavia

Gustavia sp. (figs. 209, 211 and 213; 600 μ) was widespread in these samples; it occurred in 49 of 52 sites. Though never abundant, it reached 2-percent relative abundance in the xeric shrub sere. Its seasonal abundance was typical of the total oribatid pattern. It preferred shrub (strongly avoiding clearcuts) and was found scattered through tree and old-growth sites. It avoided mesic conditions.

- 4A With broad lamellar teeth, lamellate or with translamellae not Ashaped; three tarsal claws Liacarus complex

	Sensillum	Lamellar teeth	Median tooth	Range
Rhaphidosus Liacarus Dorycranosus Procorynetes Opsioristes Adoristes Leuroxenillus Stonyxenillus Xenillus	Filiform Spindle Lanceolate Truncate Spindle Clavate Setiform Spindle Clavate	Short/large Short/large Unequal Short Tiny Long Long Short/long	- + + +/- (+) - + + +/-	transcontinental transcontinental transcontinental transcontinental western United States eastern United States transcontinental transcontinental
Paenoppia	Filiform	Long	-3	Colorado

Several ill-defined genera will key to this couplet (table 5). Generic distinctions are based primarily on the shape of the sensillum (Wooley 1969).

One group of species (*Liacarus bidentatus*, fig. 220; *L. spiniger*, fig. 221; and *L. robustus*, fig. 214, 215 and 217) are so darkly pigmented and the sizes and shapes so variable that definite species distinctions based on lamellar shape and translamellar teeth are not possible. This group was widespread (44 sites) and abundant through the sample grid, but the distribution pattern was confused; highest numbers and constancy appeared to be in the tree seres.

The smaller (270 to 310 μ) and light tan group of specimens (figs. 216 and 217) may represent more than one species. As a group, they were present in most sites (49) and followed the typical bimodal seasonal abundance pattern. The distribution pattern was confused, but did show an avoidance of xeric seres. This group's abundance was highest in midmoisture clearcut, midmoisture old-growth, and mesic oldgrowth sites, with constancy values in old growth always greater than 0.80.

The related xenillid genera *Leuroxenillus*, *Stonyxenillus* (with spindle-shaped sensilla), and *Xenillus* (with clavate sensilla) are also difficult to distinguish; they usually have prominent notogastral stippling and are treated in section Q (p. 100).



Figure 217-220; 217: *Liacarus* near *robustus* with long translamellar tooth; picture height = 225 μ ; 218: *Liacarus* sp. with typical spindle-shaped sensillum, tiny cusps and no medial translamellar tooth (TT); picture height = 275 μ ; 219: *Liacarus* small tan species with broad lamellar cusps and no medial tooth; picture height = 175 μ ; 220: *Liacarus* near *bidentatus* with elongate double-pointed lamellar cusps (LT) with no medial tooth; picture height = 300 μ .

5A Translamella minute, lamellae strongly converging and thin; tridactylous (310 μ) Furcoribula

Furcoribula was present only in midmoisture old-growth and mesic clearcut samples; in both, it was rare.

5B Lamellae broad, synlamellate for most of their length; monodactylous (200 μ) Cultroribula

The *Cultroribula biculturata* complex was rare throughout the grid, present in only 28 sites and never with a constancy value exceeding 0.50. It was abundant in the midmoisture clearcut, nearly absent in all mesic seres, and scattered without obvious pattern otherwise. It seemed rare in early summer (no data for spring or dry summer) and uniformly abundant from fall to late winter (most common in late winter).



Figure 221-222: 221: Liacarus near spiniger with 2 broad points on lamellar cusps (LT) and two minute translamellar teeth (TT); picture height = 200 μ ; 222: Furcoribula sp. high resolution of dorsum showing the converging, thin lamellae (L) which are fused to the surface for most of their length and the minute translamella; sensillum (S) is not forked, lower line is an artifact; picture height = 275 μ .

SECTION N: non-lamellate; smooth; largely the Oripodoidea (=Oribatuloidea); Oppioidea; and Eremaeoidea

This artificial grouping encompasses primarily oribatids with smooth surfaces and few prominent diagnostic characters. These species are often the most abundant in samples. The lamellar ridges, easily visible in electron micrographs, are difficult to distinguish under the dissecting microscope.



Figure 223-225; 223: Zygoribatula with conspicuous translamellar line (TL); picture height = 200 μ ; 224: Eremaeus sp. with raised and parallel lamellar ridges (LR); picture height = 550 μ ; 225: Oppiella species with costula (C) on dorsosejugal border, near absence of lamellar ridges (LR); picture height = 550 μ .

Zygoribatula (figs. 223, 227 and 228), represented in Andrews Forest by one infrequent species, is distinguished from species of similar size and shape (450 to 475 μ) by the translamellar ridge. It was present in 19 of 52 sites, but attained a constancy value of greater than 0.6 only in the xeric shrub sere. It occured sparingly in other clearcut sites and in xeric tree stages. Its seasonal abundance fit the normal bimodal pattern, with all peaks and troughs more pronounced than typical.

Lucoppia, also known from western Oregon, is distinguished by linear lamellae (ribbon-shaped in Zygoribatula) and five pairs of very reduced genital setae (four pairs in Zygoribatula).

1B Lamellar ridges, if present, then not joined by translamellar ridge (figs. 224 to 225) 2



Figure 226-228; 226: Oripoda with shortened prodorsum and prominent humeral region; 227-228: Zygoribatula sp. showing translamellar ridge (TL); picture height 227 = 550 μ , picture height 228 = 550 μ .

- 2B Anterior margin of dorsosejugal border modified, with strong obtuse angles and anteriorly projecting over at least half of prosoma as a shelf (occasionally incomplete mesally); dorsal view of prosoma (less than one-fifth of body length) covered with nearly parallelsided plate; sensillum and bothridium not visible in dorsal view (fig. 226) Oripoda

Several North American oribatuloid genera will key to this couplet, but none have been seen in Andrews Forest. Gymnobates, Neogymnobates, and Gymnobatoides have a notogaster widest at half its length and the dorsosejugal suture more than one-half the width of the mite. Oripoda (New York to New Mexico) and Parapirnodus have the notogaster widest at the humeri; in Oripoda, the dorsosejugal suture is only onethird the width of the mite and complete, but in Parapirnodus (with one tarsal claw) it is about one-half the width of the mite and much reduced.

3A Dorsosejugal suture prominent and often with cristae 4 3B Dorsosejugal suture reduced (fig. 229) or mesally interrupted; body elongate, length more than twice width; sensillum capitate Eporibatula/Gerloubia (SEE KEY NEXT PAGE)

1A Bothridia near middorsal axis, highly visible; sensillum
long filiform; posterior pronotal setae minute; Sierran
CaliforniaAmerus
1B Bothridia in the usual lateral position2
2A Elongate, length greater than twice the width; sensillum
capitate to clavate
2B Robust, length 1.5 times width; sensillum fusiform; Tennes-
see, Virginia, and AlaskaLiebstadia
3A One notogastral seta in dorsal humeral region (10 pairs
of notogastral setae)4
3B Two notogastral setae in dorsal humeral region (14 pairs
of notogastral setae; sensillum capitate; 4 pairs of genital
setae Eporibatula
4A TridactylousParaphauloppia
4B MonodactylousGerloubia
o to in the set of the

This is a very diverse taxonomic assemblage of genera. Though several genera are reported from the west coast, only one species has thus far been found in Andrews Forest. *Eporibatula* sp. (fig. 229; 130 μ) was infrequently collected in the research grid, primarily in mesic and xeric tree sites. The characteristics that separate the genera must be seen under a compound microscope.



Figure 229-231; 229: Eporibatula sp. with nearly rudimentary dorsosejugal suture (no DSJ), lamellar ridges very reduced; picture height = 130 μ ; 230: Megeremaeus ditrichosus with cristae on anterior margin of notogaster (C); picture height = 1350 μ ; 231: Eremaeus sp. in dorsal view showing prominent costula, lack of spinae adnatae, and cerotegument; picture height = 975 μ .

5A Crista present (spiny tubercles arising from notogaster along dorsosejugal suture on either side of midline) Megeremaeus

Megeremaeus ditrichosus Woolley & Higgins (fig. 230; 925 to 950 μ) has been taken frequently by Walter (unpub.) in western Oregon. In Andrews Forest, it seemed to be restricted to subalpine forest litter.

5B Crista absent (figs. 231 and 232) Eremaeus



Figure 232-234; 232: Eremaeus arboreal species in dorsal view; picture height = 425 μ ; 233: Eremaeus sp. Andrews Forest litter species in dorsal view, with long notogastral setae; picture height = 475 μ ; 234: Eremaeus sp. (Marys Peak, OR.) in dorsal view (LR = lamellar ridges); picture height = 725 μ .

Many species of *Eremaeus* apparently occur in Oregon, but only four were in Andrews Forest. Voegtlin (1982) reports two species in high abundance from mosses (fig. 232) and lichens of old-growth Douglas-fir canopy. Neither species was taken on the ground. A third widespread and abundant species was found in 46 of these sites (figs. 231, 233 and 235; 750 to 780 μ). It occurred in old growth, regardless of moisture regime. Seasonal abundance followed the typical bimodal pattern, but the fall peak was advanced and declined primarily in dry summer (similar to the pattern observed for *Odontodamaeus veriornatus* and *Oribotritia megale*). A fourth species was abundant in the subalpine fir forest of Andrews Forest (fig. 270); this species has the most noticeably stippled dorsum and may key to Section P (*Kartoeremaeus*) page 96.

7A Anterodorsal tip of prosoma with linear ridge (fig. 239); sensillum linear Rhynchobelba

Rhynchobelba was frequent in Andrews Forest samples; it was nearly the same size, color, and shape as the largest species of *Oppiella* and has not been treated separately in our analysis.

Rhinosuctobelba cf. dicerosa Woolley & Higgins (figs. 236, 238 & 240; 500 to 575 μ) was widespread and abundant in Andrews Forest, occurring in all 52 sites. Though not abundant enough to be among the top 15 species of oribatids, constancy within the community replicates is high, dropping below 0.70 only in xeric clear-cut and old growth. It exhibits the typical seasonal pattern, but the spring peak is delayed through early summer. It is generally distributed throughout all moisture regimes and successional seres (with perhaps a slight preference for shrub and tree seres).







Figure 236-240; 236-237: Chelicerae of *Rhinosuctobelba dicerosa* emphasizing their elongate structure (lateral view and ventral views); picture height 236 = 225 μ , picture height 237 = 133 μ ; 238: *Rhinosuctobelba dicerosa* with elongate t-shaped snout(R = rostrum); picture height = 750 μ ; 239: *Rhynchobelba* with prominent anterodorsal snout (R = rostrum); 240: *Rhinosuctobelba dicerosa* with prominent ventral snout; picture height = 750 μ .

8A Lamellae expressed as a distinct, raised linear ridge (fig. 241)...

SEE KEY BELOW

		Dorsosejugal suture straight
		Dorsosejugal suture normally curved2
2	2A	Dorsosejugal suture medially interrupted; aquatic;
		with lateral cerotegumentary border; 16 to 17 pairs of
		notogastral setaeHydrozetes
2	2B	Dorsosejugal suture not interrupted; no cerotegumentary
		bands; non-aquatic3
-	ΒA	Sacculonotic; 10 pairs of notogastral setae5
0.1	BB	Poronotic; 14(12) pairs of notogastral setae4
L	ŧΑ	Lamellae linear, shortPhauloppia
L	ŧΒ	Lamellae raised, ribbon-likeOribatula
5	δA	Rostrum pointed
5	5B	Rostrum roundedDometorina/Hemileius
6	5A	Lamellar ridges parallel and closer to each other than
		to sides of prodorsum; sensillum with an elbow-bend;
		anterior margin of notogaster with two tubercles;
		transcontinentalDolicheremaeus
6	6B	Lamellar ridges converging, close to lateral margins
		of prodorsum; sensillum capitateDometorina

These genera are cited in the catalog from North America but only the two in couplet 4 are likely to be found in arboreal or litter samples from the Cascades. Topic and Walter (unpub.) encountered *Oribatula* in litter samples and Voegtlin (1982) found high numbers of *Phauloppia* in arboreal canopy studies. The genus *Dometorina* (Nova Scotia) is distinguished by bothridia covered by the notogaster; *Hemileius* (eastern half of the United States and Canada) by visible bothridia; *Metaleius* (North Carolina) by normal three-clawed legs; and *Paraleius* (Louisiana) by a modified posteriorly pointing median claw.



Figure 241-243; 241: Oribatula sp.; picture height = 400 μ ; 242-243: Phauloppia sp. with lamellar ridges (LR) and sensillum (S) indicated; picture height 242 = 225 μ , picture height 243 = 425 μ .



Figure 244-246; 244-246: Hydrozetes sp. in dorsal and lateral views with high magnification of dorsosejugal region; note the circumferential ring of cerotegument (CT) that serves as a plastron, the absence of sensilla and the broadly interrupted dorsosejugal suture; picture height 244 = 350 μ , picture height 245 = 350 μ , picture height 246 = 150 μ .

The marine ameronothroid Alaskozetes (Alaska to California) will key at couplet 9 (fig. 260), but it is quite unlike the Suctobelbidae or Oppiidae. The notogaster is nearly circular; the prosoma subequal in width to the notogaster and about half its length; without traces of lamellae; with very reduced capitate sensilla; interlamellar setae many times the length of the sensillum; and a dense cluster of elongate twisted setae between the genital and anal plates.

9B Minute species, largely transparent, without dark food boli in the gut; 150 to 175 μ ; with two transverse ridges across most of prosoma (figs. 250 to 251); prosoma nearly as wide as opisthosoma; anterior edge of notogaster with prominent saddle-like prominences (crista) Quadroppia

Quadroppia cf. quadricarinata (Michaels) (160 μ) was the second most abundant mite, occurring in all 52 sites. Its constancy value ranked fourth among total oribatids; however, its minute size renders it one of the most inconspicuous mites in the samples. Its seasonal abundance curve closely follows that for total Andrews oribatids. It prefers mesic to midmoisture to xeric seres. It appears independent of successional type, except that it is nearly absent in xeric clearcut. It is probable that a second infrequent and larger species also occurs in the Andrews Forest.

10A Prosoma nearly as long and wide as opisthosoma; never with dark food boli; bright red-brown; costula usually present; chelicerae not the usual lobster-claw shape; dorsal portion of prosoma with pair of flattened surfaces (fig. 247-249) Suctobelbidae (SEE KEY TO GENERA)

- 1A Dorsosejugal suture with one or two teeth or notogaster with saddle-like crista (fig. 248).....2 1B Dorsosejugal suture without teeth; notogaster without
- tubercles; dorsosejugal suture with two teeth (fig. 248)..... Suctobelbella 3A Rostrum with incision (fig. 247)..... Suctobelba 3B Rostrum without incision (Alaska to North Carolina)

.....Allosuctobelba

This group was represented in Andrews Forest by at least two species of Suctobelbella (figs. 248 and 253; ca. 200 to 250 μ). As a genus, Suctobelbella was found in all 52 sites, ranking first in consistency and third in abundance. It appeared uniformly distributed throughout the successional seres and moisture gradients but was lowest in the clearcut. Seasonal abundance closely fit the pattern for total oribatids.



Figure 247-249; 247: Suctobelba sp. with smooth patches (SP) on prosoma and dorsosejugal suture without teeth; picture height = 230 μ ; 248: Suctobelbella sp. with smooth patches (SP) on prodorsum and dorsejugal suture with a prominent pair of lateral teeth (C); picture height = 110 μ ; 249: Rhinosuctobelba dicerosa with smooth patches (SP) on prosoma and snouted rostrum (R; S = sensillum); picture height = 300 μ .

10B Prosoma distinctly narrower than opisthosoma and never half its length; food boli usually seen as contrasting dark spots in dorsal view; legs with a single tarsal claw Oppiidae(principally) (SEE TABLE OF GENERA)

Distinguishing features of the genera related to *Oppiella* are presented in tabular format for ease of reference.

CRISTA COSTULA	SENSILLUM	NOTES
Oribella - +	Fusiform	Epimeres III & IV not fused; setae and dorsum unmodified (New York, California)
<i>Veloppia</i> Pair - tubercles	Fusiform	Epimeres III & IV not fused; notogaster with longitudinal sulci; setae large 200 μ ; (California, Oregon)
Autogneta Long +	Fusiform	Rostral tip incised; 230 μ (transcontinental)
<i>Oppiella</i> Short +	Fusiform	Figs. 254, 256, 257 & 259; 175 to 375 μ (transcontinental)
Quadroppia Short +/-	Golf club	Prosoma with two transverse ridge edges appearing as lines (transcontinental)
Banksinoma Short/ - converging	Long setose	Genital and anal plates very large, touching; 340 μ (transcontinental)
Gemmazetes Short -	Clavate/ filiform	Genital and anal plates very large, touching; arboreal; 300 μ (Alaska, California)
Brachioppiella Short -	Pectinate	Normal rostral setae (California)
Ramusella Short -	Pectinate	Plumose rostral setae (transconti- nental)
Oppia	Long setose; fusiform	Figs. 255 & 258 (transcontinental)
Multioppia	Fusiform	Additional notogastral setae, (13 pairs not usual 10)(Quebec to Louisiana)
Ameroppia	Capitate fig. 261	Interlamellar setae absent (Illinois to tropics)
Aeroppia	Capitate/ ball	Posterior notogastral setae thick and hollow (eastern half USA)

Nearly all Andrews Forest oppiids fall into the genus Oppiella, a genus that probably contains at least three species locally. One species is noticeably larger (325 to 375 μ) and light reddish; it may be confused with *Rhynchobelba*, which is separable by the much deeper bright red color and the absence of all traces of food boli. Lumped in our analysis, these two species were 12th in total abundance, found in 51 of 52 sites, and have constancy values above 0.8 in all but the clearcut sites. Seasonally, they followed the typical abundance pattern, but are less frequent in dry summer than early summer. They were widespread and abundant in all successional seres and moisture types, perhaps with a slight aversion to clearcuts.

The other two species are nearly the same size (175 to 250 μ) but differ in color; one is a transparent reddish, and the other (*O. nova* (Oudemans)) is light tan. Together, they comprised the most abundant and the second most constant oribatid group in Andrews Forest. They were found in all 52 sites, and achieved a constancy value of 1.0 in all sites except the xeric clearcut. Seasonal abundance closely follows the norm. They preferred mesic to midmoisture and avoided xeric seres. Old-growth and tree seres were preferred to shrub stages, which were preferred to clearcuts.

Gemmazetes clavatus (300 μ) occurs arboreally on oak in the Willamette Valley (Walter, unpub.). The genera Cosmoppia (Florida), Microppia (transcontinental, including British Columbia) and Pantelozetes (Alaska) belong in this alliance, but we do not know their diagnostic characters.



Figure 250-252; 250: Quadroppia quadricarinata notice the promient costula and transverse ridge on the prodorsum in lateral view; picture height = 200 μ ; 251: Quadroppia quadricarinata with raised dorsum of prosoma highlighted by a transverse ridge (TR), with prominent costula (C); picture height = 150 μ ; 252: Banksinoma sp. with irregular denticulations on the translamellar ridge (TR; S = sensillum); picture height = 225 μ .



Figure 253-256; 253: Suctobelbella sp. with pair of smooth patches (SP) on the prodorsum and prominent crista (C); picture height = $325 \ \mu$; 254: Oppiella nova dorsal view with crista (C); picture height = $400 \ \mu$; 255: Oppia sp. dorsal view without crista; picture height = $333 \ \mu$; 256: Oppiella sp. in ventral view; picture height = $400 \ \mu$.



Figure 257-261; 257: Oppiella nova dorsosejugal region showing costula (LR) and crista (C); picture height = 150 μ ; 258: Oppia sp. dorsosejugal region without costula or crista; picture height = 100 μ ; 259: Oppiella sp. showing costula (LR) and crista (C); picture height = 150 μ ; 260: Alaskozetes sp. with very elongate setae (A) posterior to the genital sclerites; 261: species near Ameroppia with capitate sensillum; picture height = 350 μ .

SECTION P: Mites with patterned backs; non-lamellate

This artificial category includes a wide variety of unrelated oribatids sharing only the prominent surface sculpturing and lack of lamellae. A wide variety of body shapes and sizes will key to this section.



Figure 262-265; 262-263: Scapheremaeus arboreal species of Voegtlin in lateral and dorsal views, note the circular groove (A) within the notogastral pattern of cerotegument, capitate sensillum (S) and flat dorsum; picture height $262 = 370 \ \mu$, picture height $263 = 400 \ \mu$; 264: Nanhermannia with uniform punctation, oblong-cylindric body and no submarginal groove; picture height = 550 μ ; 265: Nanhermannia sp. with large U-shaped area (A) occupying anterior half of opisthosomal venter encompassing the genital plates (GS) but anterior to the anal plates (AS); picture height = 450 μ .

1A Dorsum of opisthosoma flat (fig. 262), with complete circular groove emphasizing irregularly multiridged cerotegument (fig. 263); lamellae present, trans-lamellate, but inconspicuous amongst cerotegument; without prominent setae (Cymbaeremaeoidea) Scapheremaeus

Voegtlin (1982) found one species of *Scapheremaeus* (350 to 400 μ) abundant on bare branches of the canopy of Douglas-fir old growth; we have never encountered it in litter samples from under the same trees.

The related genus *Micreremus* differs in having four pairs of genital setae (six pairs in *Scapheremaeus*), a nonreticulate prosoma, and a uniformly honey-combed notogaster; it has not been collected in Oregon (recorded only from Massachusetts).

- 1B Notogaster not flat, without complete circular sulcus (at most with an indistinct pair of longitudinal folds extending three-quarters the length of notogaster); with prominent setae on notogaster ... 2

Nanhermannia (figs. 264 and 265; 450 to 500 μ) was widespread (45 sites) but never abundant in the grid. The seasonal abundance pattern was not clear because data from fall or late winter are absent, but a peak occurred in spring--as is typical. It was distributed nearly exclusively in tree and old-growth seres (preferring the tree stage) and preferred xeric over midmoisture over mesic conditions.

Veloppia (reported by Topic and Walter (unpub.) from Salmon Creek drainage just south of the Andrews Forest) has very long notogastral setae, conspicuous cerotegument, and two additional tubercles along the dorsosejugal suture (200 μ).

The eastern genus *Licneremaeus* may be encountered in the Pacific Northwest. *Licneremaeus* has expanded, clubbed sensilla and a pattern of large honeycombs on the anterior portion of the notogaster (fig. 266). The marine *Ameronothrus* (not illustrated) may key here; its habitat is sufficient to distinguish it.



Figure 266-269; 266: Licneremaeus with interrupted anterior portion of notogaster; 267-268: Hermannia displaying convex notogaster with thick layer of cerotegument and prominent setae; picture height $267 = 900 \ \mu$, picture height $268 = 900 \ \mu$; 269: Basilobelba with unique nymphal exuvia.

Hermannia (figs. 267 and 268; 750 to 900 μ) is frequently encountered in the Coast Range, but has not turned up in Andrews Forest samples.

5B Widest at the two-thirds to three-quarters distance; bothridium prominent; sensillum elongate Amerobelboidea (=Eremuloidea)

Several genera of the Amerobelboidea may occur in the conifer forests of the Pacific Northwest; the following are the most likely. *Basilobelba*, recorded by Walter (1981) from central California (fig. 269), carries nymphal shed skins (with an anterior cobra-like extension) on the notogaster; has a filiform, pectinate sensillum; and long, plumose, whip-like setae posteriorly. *Fosseremus* (from North Carolina and the District of Columbia) has four prominent depressions, which give it an H-shape, interrupting the notogastral reticulation; an elongate, capitate sensillum; and a curved dorsosejugal suture. *Eremobelba* (fig. 271), reported by Walter (1981) from California litter, has a notogaster with fine pits arranged in a large honeycomb design; very long, whip-like sensilla; and prominent notogastral setae. *Damaeolus*, reported from the District of Columbia, has a finely punctate notogaster and prodorsum; elongate, spindle-shaped sensilla; and 11 pairs of elongate, twisted notogastral setae. *Eremulus*, reported from the east coast of the United States, has a finely punctate notogastral dorsum, an elongate sensillum, and 11 pairs of elongate notogastral setae.

Kartoeremaeus (transcontinental; fig. 270) is an eremaeoid with a densely pitted cerotegument (450 to 600 μ); it differs from the amerobelboid genera in having three claws; only a single pair of adgenital setae (three in Amerobelboidea); four to five pairs of genital setae (two in Amerobelboidea), and more than three pairs of adanal setae.



Figure 270-271; 270: Kartoeremaeus subalpine species with pitted integument; picture height = 700 μ ; 271: Eremobelba sp. with patterned cerotegument and prominent notogastral setae; picture height = 450 μ .

SECTION Q: Mites with patterned notogasters and lamellae; Cepheoidea, Carabodoidea

This is an artificial category created for convenience of identification. Mites falling into this section are instantly distinguished from those appertaining to the other sections; however, such a wide variety of body shapes and cerotegumentary patterns are possible that there is no single type representative of the whole group.



Figure 272-274; 272: Scapheremaeus arboreal species with nearly obscured lamellae (L), a flat dorsum, a capitate sensillum (S), and a circular groove (A) in the cerotegument; picture height = 250 μ ; 273: Eupterotegaeus dorsal view with large lamellae (L), thickly patterned cerotegument and circular groove (A); picture height = 750 μ ; 274: Eupterotegaeus sp. with the broad lamellae (L) incurved at their tips (S = sensillum); picture height = 400 μ .

- a major part of the integumentary pattern; circular grooves, when present, not complete anteriorly 3
- 2A Notogaster convex, with exuviae forming one or two concentric circles; prosoma nearly one half length of opisthosoma, covered with dense cerotegument; lamellae projecting as prominent anterior horns (apically flexed mesally, though often obscured with cerotegument) Eupterotegaeus

Eupterotegaeus (figs. 273, 274 and 293; 675 to 750 μ) was represented by an infrequent species that occurred in 12 sites, but nowhere with a constancy value above 0.50. It was present primarily in midmoisture shrub and mesic old-growth stages. It was not abundant enough to reveal its habitat preference or seasonal pattern.

Voegtlin (1982) discovered one species (350 to 400 μ) abundant on bare branches of old-growth canopy; it has not been encountered in litter from under the same trees.





Figure 275-279; 275: Leuroxenillus, the form with notogastral setae in dorsal view; picture height = 1350 μ ; 276: Leuroxenillus with regular field of coarse punctation, convergent lamellae with a tooth-like projection from the translamella (TT); picture height = 900 μ ; 277: Stenoxenillus sp. with converging simple lamellae; 278: Stonyxenillus with broad translamellar cusps and a short translamellar ridge; 279: Scutovertex sp. with narrow translamellar cusps and a wide translamellar ridge, anterior border of notogaster interrupted.

3A Dorsum with irregular pattern of transverse ridges and grooves; lamellae prominent, covering all of prosoma, strongly converging (fig. 280) Ametroproctus/Coropoculia

Ametroproctus is abundant in old-growth canopy (500 to 520 μ) and has not been collected in litter (fig. 280).

Ametroproctus (Coropoculia) was collected six times in the research grid (each time represented by only a single individual (fig. 281). It has indistinct lateral notogastral setae and six pairs of genital setae (notogastral setae absent; four pairs of genital setae in Ametroproctus).

3B Dorsum without irregular pattern of transverse ridges and grooves

4A Dorsum with prominent pattern of equally spaced punctures (figs. 275 and 276); no cerotegument on body and lamellae; lamellae sharply pointed anteriorly Leuroxenillus

How many species are represented in Andrews Forest samples (900 to 1200 μ) is not clear. One group of specimens (=Leuroxenillus trichionus?) is characterized by long notogastral setae (fig. 275). It occurred in 15 sites, but was too infrequent to show seasonal patterns. It was encountered only in xeric sites, which it prefers to shrub and tree communities. The other group (=Stonyxenillus?; fig. 278) appeared to lack setae under the dissecting microscope; one or two species were present. Though infrequent, this group was widely distributed and occurred in 48 sites. It demonstrated the typical seasonality pattern and occurred in roughly equal abundance in shrub, tree, and old-growth communities.

The genus *Xenillus* (distinguished by its capitate sensillum) is also reported from North America (Balogh 1972). *Stenoxenillus* (fig. 277; Alabama and North Carolina) differs from both these genera in having narrow lamellae (weakly translamellate), which end in short pointed tips.

4B Dorsum and lamellae covered with patterned cerotegument, not shiny (figs. 282 and 287) 5 280 281



Figure 280-281; 280: Ametroproctus with a prominent pattern of transverse ridges and grooves, broad strongly converging lamellae (L), and capitate sensillum (S); picture height = 550 μ ; 281: Ametroproctus (Coropoculia) with prominent pattern of irregular ridges, equally prominent on convergent lamellae (L); picture height = 650 μ .



Figure 282-284; 282: Sphodrocepheus tridactylus with wide, strongly convergent lamellae (L), a thin cerotegument, 1 pair of humeral notogastral setae and nearly circular profile; picture height = 1100 μ ; 283: Sphodrocepheus anthelionus with broad converging lamellae (L) and 2 pairs of humeral notogastral setae (HS); picture height = 550 μ ; 284: Carabodes sp. in ventral view showing typical arrangement of genital and anal plates; picture height = 375 μ .

Ommatocepheus cf. *clavatus* Wooley is an abundant arboreal mite characteristic of Garry oak in the Willamette Valley; it was not encountered in Andrews Forest.

7A Three tarsal claws; notogaster strongly convex Sphodrocepheus

Two species occurred in Andrews Forest (figs. 282 and 283). Both had the fall abundance peak partially shifted into dry summer. Sphodrocepheus tridactylus Woolley & Hammer is slightly smaller and is distinguished by the shorter notogastral setae and one pair of setae at the humeral expansion; it was endemic to old growth, occurring in 28 sites. It appeared to have no moisture preference between different old-growth sites. Sphodrocepheus anthelionus Woolley & Hammer has longer notogastral setae, a distinct translamella, and two pairs of setae at the humeral expansion. The species was limited to the tree stage, exhibiting a preference for mesic over midmoisture over xeric regimes; this species, much more numerous, was found in 31 sites and along the Topic and Walter (unpub.) gradient in Salmon Creek.

7B With a single tarsal claw Cepheus/Oribatodes

Cepheus (fig. 288) is synlamellate (with diverging apices); Oribatodes has separate lamellae with converging apices. Walter (unpub.) has collected both genera in western Oregon, but as of yet neither has been found in the Andrews Forest.



Figure 285-287; 285: Carabodes with a thick cerotegument, nearly parallel lamellae, elongate body and distinct dorsosejugal suture; picture height = 375 μ ; 286: Carabodes large black species with prominent concentric groove (A), reticulated dorsum and parallel lamellae (L); picture height = 550 μ ; 287: Tectocepheus velatus with a thick cerotegument, parallel lamellae (L), elongate body and vestigial dorsosejugal suture (300x); picture height = 325 μ .

Tectocepheus (fig. 287) was encountered in all 52 sites and had a constancy value above 0.70 everywhere except the depauperate xeric clearcut. Though widespread and constant, it was not generally abundant. Seasonal abundance patterns were bimodal but notably low in early summer and high in early winter. No preferred successional sere was evident and the distribution was apparently independent of the moisture regimes as well. Abundance was high in the midmoisture clearcut site.

Also keying to this position is *Scutovertex* (fig. 279; Licneremaeoidea). Walter (1981) records it from central California litter. It has a fusiform sensillum, thin parallel lamellae connected by a transverse bridge, irregularly wrinkled notogaster, and a strongly curved and medially interrupted dorsosejugal suture.

9A Dorsum with pattern of longitudinal ridges and grooves in cerotegument; notogaster strongly convex, nearly hemispheric; color black Exochocepheus

Exochocepheus (fig. 292) has been collected only once in a subalpine meadow of Andrews Forest. A similar genus, Niphocepheus (fig. 290), also possibly occurs in the Pacific Northwest. Unlike Exochocepheus, which has simple lamellae with the teeth shorter than the lamellar bases, Niphocepheus is synlamellate with the length of the teeth subequal to the lamellar bases.

Carabodes probably is comprised of at least two small yellowish species locally (figs. 284-285; 250 to 350 μ). Carabodes was present in 45 of the sites; it ranked 6th in total density, 10th in relative abundance but less than 20th in constancy. Seasonal abundance fit the norm for Andrews oribatids. It was nearly absent from clearcut and shrub samples (constancy less than 0.5), and seemed to prefer tree and old-growth sites in all three moisture regimes equally (but is inexplicably absent from the mesic tree site).

An additional distinctive species of *Carabodes*, much larger and completely black, is present in the Andrews Forest (fig. 286; 500 μ). It was an infrequent species found in only 7 of 52 sites and restricted to mesic shrub.

Odontocepheus has a transverse anterior margin of the notogaster with two teeth and a row of four large, anteriorly directed setae just posteriad. Dolicheremaeus has a transverse anterior margin of the notogaster with two tubercles but without the row of anteriorly directed setae (fig. 291). Tegeocranellus resembles the profile of Carabodes, with a convex anterior notogastral margin; in Tegeocranellus the anterior portion of the notogaster has a pair of longitudinal folds.



Figure 288-291; 288: Cepheus sp. with converging synlamellae (L); 289: Ommatocepheus sp. with enlarged bothridium and capitate sensilla; 290: Niphocepheus sp. with pattern of many longitudinal lines in the cerotegument; 291: Dolicheremaeus sp. with transverse anterior notogastral margin.



Figure 292-293; 292: Exochocepheus sp. with three longitudinal ridges on the notogastral cerotegument; picture height = 750 μ ; 293: Eupterotegaeus sp. high resolution lateral view of head region showing the lamellae (L); picture height = 375 μ .

MAJOR TYPES OF IMMATURES

Figure 294-305; 294: Cepheoidea (*Sphodrocepheus*, protonymph); picture height = 1100 μ ; 295: Cepheoidea (*Sphodrocepheus*, tritonymph); picture height = 550 μ ; 296: Cepheoidea (*Ommatocepheus*); 297: Liodoidea (*Platyliodes*); picture height = 1000 μ ; 298: Plateremaeoidea (*Jacotella*, tritonymph with nymphal exuviae removed); picture height = 525 μ ; 299: Plateremaeoidea (*Jacotella*, tritonymph); picture height = 475 μ ; 300: Damaeoidea (*Caenobelba*); picture height = 850 μ ; 301: Damaeoidea (*Epidamaeus*); picture height = 600 μ ; 302: Damaeoidea (*Hungarobelba*); picture height = 425 μ ; 303: Eremaeoidea (*Megeremaeus*); picture height = 450 μ ; 304: Eremaeoidea (*Eremaeus*, common species); picture height = 450 μ ; 305: Hermanniellioidea (*Hermanniella*); picture height = 450 μ .



MAJOR TYPES OF IMMATURES

Figure 306-317; 306: Enarthronota (*Liochthonius*); picture height = 175 μ ; 307: Mesoplophoroidea (*Mesoplophora*); picture height = 275 μ ; 308: Crotonioidea (*Camisia*); picture height = 925 μ ; 309: Crotonioidea (*Platynothrus*); picture height = 775 μ ; 310: Crotonioidea (*Nothrus*); picture height = 675 μ ; 311: Nanhermannioidea (*Nanhermannia*); picture height = 425 μ ; 312: Gustavioidea (*Ceratoppia*); picture height = 1100 μ ; 313: Gustavioidea (*Liacarus*?); picture height = 700 μ ; 314: Gustavioidea (*Leuroxenillus*?); picture height = 775 μ ; 315: Oppioidea/Oripodoidea; picture height = 300 μ ; 316: Oppioidea/Oripodoidea; picture height = 350 μ ; 317: Hermannioidea (*Hermannia*); picture height = 500 μ . **306 307 308 309**



MAJOR TYPES OF IMMATURES

Figure 318-323; 318: Tectocepheoidea (*Tectocepheus*); picture height = 200 μ ; 319: Phenopelopoidea (*Eupelops*); picture height = 350 μ ; 320: Phenopelopoidea (*Propelops*?); picture height = 400 μ ; 321: Achipteriidae; picture height = 450 μ ; 322: Achipteriidae; picture height = 425 μ ; 323: Achipteriidae; picture height = 425













DIAGRAMMATIC KEY TO MAJOR TYPES OF IMMATURES





INDEX

Acarida	34	a	100
Acaronychus	32, 33	Coropoculia	
Achipteria	6-7, 19, 45-47,	Cosmochthonius	26-27
<u>-</u>	53	Cosmoppia	92 68
Achipteriidae	45, 107	Crotonioidea	
Achipterioidea	45	Ctenacarus Culturation la	32, 33
Acrogalumna	43	<i>Cultroribula</i>	6, 81 95
Adoribatella	53	Cymbaeremaeoidea Damaeoidea	72
Adoristes	78-79	Damaeus	72
Aeroppia	91	Dameolus	97
Akrodamaeus	73	Dasybelba	74
Alaskozetes	89, 94	Dentizetes	54
Allodamaeus	4	Diapterobates	58
Allosuctobelba	90	Dolicheremaeus	88, 103
Amerobelboidea	4, 74, 96-97	Dometorina	88
Ameronothrus	96	Dorycranosus	79
Ameroppia	91, 94	Dyobelba	73
Amerus	84	Enarthronota	25, 34
Ametroproctus	18, 99-100	Eniochthonius	28-29
Anachipteria	6-7, 19, 45-53,	Eobrachychthonius	29
	55	Eohypochthonius	6-7, 18, 27-28
Anoribatella	53	Epidamaeus	6-7, 18, 73-75,
Aphelacarus	32, 33	Lpidamacas	105
Atopochthonius	26-27	Epilohmannia	6-7, 18, 34,
Atropocarus	38	Sprionmanning	65-66, 71
Autogneta	91	Eporibatula	6-7, 18, 83-84
Banksinoma	91, 92	Eremaeoidea	82, 84
Basilobelba	74, 96-97	Eremaeus	6-7, 18, 63,82,
Beklemishevia	32, 33	<u>El ondoub</u>	84-86, 105
Belba	19, 73	<i>Eremobelba</i>	97
Belboidea	72	Eremuloidea	96
Bifemoratina	32, 34	Eremulus	97
Brachioppiella 44	91	Eulohmannia	18, 34-35
Brachychochthoniu	<i>us</i> 30	Eupelops	6-7, 13, 19,
Brachychthoniidae	4, 29, 31	200020000	48-50, 71,
Brachychthonius	6-7, 18, 29-30		107
Caeculus	19	Euphthiracaroidea	
Caenobelba	6-7, 18-19, 72-	Euphthiracarus	6-7, 18, 37,
	73, 75, 105	<u> </u>	41-42
Camisia	7, 19, 35-36,	Eupterotegaeus	19, 98, 104
	69, 106	Euzetes	59
Carabodes	6-7, 18-19,	Exochocepheus	103, 104
	101-103	Fosseremus	97
Carabodoidea	98, 101	Furcoribula	18, 81
Cepheoidea	98, 101	Fuscozetes	59
Cepheus	101-103	Galumna	43
Ceratoppia	6-7, 18, 76,	Gehypochthonius	34
	106	Gemmazetes	91-92
Ceratozetes	6-7, 18, 45,	Gerloubia	83-84
	51, 59-61	Grypoceramerus	4
Ceratozetoidea	48, 51, 55	Gustavia	6-7, 13, 77, 79
Chamobates	59	Gustavioidea	45, 76
Conoppia	78		 A second sec second second sec

Gymnobates 83 83 Gymnobatoides Gymnodamaeoidea 62 Gymnodamaeus 63 57-58 Haplozetes Hemileius 88 Heminothrus 71 Hermannia 19, 96, 106 6-7, 18-19, 67, Hermanniella 105 Hermannielloidea 67, 105 Heterozetes 59 Hoplophorella 38 Hoplophthiracarus 38 Humerobates 59 Hungarobelba 6-7, 18, 74, 105 88-89 Hydrozetes Hygroribates Hypochthoniella 28 Hypochthonius 6, 18, 25, 28 7, 60-61 Hypozetes Iugoribates 61 Jacotella 6-7, 18, 64, 105 *Johnstonella* 63 Joshuella 64 7, 18, 54-55 Jugatala Kartoeremaeus 85, 97 Kodiakella 77-78 Lepidozetes 54 Leuroxenillus 6-7, 19, 79, 99-100, 106 Liacaroidea 76 Liacarus 6-7, 18-19, 50, 79-81, 106 96, 102 Licneremaeoidea Licneremaeus 96 Licnodamaeus 62 Liebstadia 84 Limnozetes 58-59 Liochthonius 6-7, 29-30, 106 Liodes 68-69 Liodoidea 68, 71 Lucoppia 82 6-7, 18, 39-40 Maerkelotritia Malaconothrus 36 Megeremaeus 19, 84-85, 105 Melanozetes 59-60 Mesoplophora 37. 106 Mesoplophoroidea 37 Mesotritia 40 Metaleius 88 Metapyroppia 78 6-7, 18, 77-78 Metrioppia Micreremus 95 Microppia 92

18, 40-41 Microtritia Microzetoidea 47 57 Minuthozetes Mixochthonius 29 Mixonomata 37 55 Mochlorizetidae 36 Mucronothrus 91 Multioppia Mycobates 54, 57 Nanhermannia 6-7, 18, 95-96, 106 95 Nanhermannioidea Neobrachychthonius29 Neogymnobates 83 29 Neoliochthonius 71 Neonothrus 43 Neoribates Niphocepheus 103 26-27 Nipponiella Nortonella 64 Nothroidea 68 6-7, 19, 69-71, Nothrus 106 103 Odontocepheus Odontodamaeus 7, 19, 62-64, 85 101, 103, 105 Ommatocepheus Opiidae 4, 82, 89, 91 18, 91, 94 Oppia 6-7, 18, 45, Oppiella 82, 86, 91-94 4 Oppioidea Opsioristes 79 6-7, 18-19, 52-Oribatella 53 Oribatelloidea 48-49, 51 101-102 Oribatodes Oribatula 88 Oribatuloidea 51, 55, 82 Oribella 91 Oribotritia 6-7, 18, 39, 63, 85 57, 83 Oripoda 51, 55, 82 Oripodoidea 79 Paenoppia Palaeacarus 32, 33 32, 34 Palaeosomata Pantelozetes 92 45 Parachipteria 43 Parakalumna Paraleius 88 58 Parapelops 84 Paraphauloppia 76 Parapyroppia Parhypochthonius 34, 67 Passalozetes 4 Passalozetoidea 4, 96

_		~	<pre>< 7 10 / 5</pre>
Pelopsis	58	Scheloribates	6-7, 18, 45,
Peloptulus	49-50		51, 55-57
Peloribates	57-58	Scutovertex	99, 102
Peltenuiala	6-7, 19, 45-47	Scutozetes	54
Pergalumna	18-19, 43	Sellnickochthoniu	
Perlohmannia	6-7, 18, 35,	Sphaerochthonius	26-27
	65, 71	Sphaerozetes	6-7, 18, 58-60
Phauloppia	88	Sphodrocepheus	6-7, 19, 101,
Phenopelopoidea	48-49		105
Phthiracaroidea	38	Spinozetes	4
Phthiracarus	6-7, 18, 38, 41	Steganacarus	38
Phyllozetes	26	Stenoxenillus	99-100
Pilogalumna	43	Stonyxenillus	19, 78-79, 99-
Plateremaeoidea	4, 62		100
Plateremaeus	62	Suctobelba	90
Platyliodes	7, 18, 68, 105	Suctobelbella	7, 13, 18, 45,
Platynothrus	19, 69-71, 106		90, 93
Pleodamaeus	62	Suctobelbidae	89, 91
Plesiotritia	39	Suctobelbila	90
Podoribates	59	Synchthonius	29
Poecilochthonius	29	Synichotritia	41
Poroliodes	68	Tectocepheus	6-7, 18, 102,
Procorynetes	79	-	107
Propelops	6-7, 19, 48-50,	Tectoribates	53
	107	Tegeocranellus	103
Prostigmata	34	Tegoribates	54
Protobelba	73	Teleioliodes	68
Protoplophora	37	Tenuiala	7, 45-47
Provertex	4	Tenuialidae	45
Pseudachipteria	45	Tenuialoides	46
Pterochthonius	25	Traychetes	18
Ptyctimina	37	Trhypochthoniell	<i>us</i> 35
Punctoribates	58	Trhypochthonius	18, 35
Pyroppia	76	Trichoribates	59
Quadroppia	6-7, 18, 45,	Trimalaconothrus	36
	89-92	Veloppia	91, 96
Quatrobelba	72	Verachthonius	29
Ramusella	91	Xenillus	79, 100
Rhaphidosus	79	Xylobates	57
Rhinosuctobelba	6-7, 13, 18,	Zachvatkinella	18, 32, 33
	45, 71, 86-87,	Zachvatkinibates	59
	90	Zetomimus	61
<i>Rhynchobelba</i>	86-87, 92	Zygoribatula	6-7, 18, 82-83
Rhysotritia	40	-/6	., _, _, ,
Scapheremaeus	7, 18, 71, 95,		
	98-99		
	70-77		