Facilitating Reforestation Through the Plant Microbiome: Perspectives from the Phyllosphere

Posy E. Busby,1 George Newcombe,2 Abigail S. Neat,1 and Colin Averill3

1Department of Botany and Plant Pathology, Oregon State University, Corvallis, Oregon, USA; email: posy.busby@oregonstate.edu
2Department of Forest, Rangeland and Fire Sciences, University of Idaho, Moscow, Idaho, USA
3Department of Environmental Systems Science, Institute of Integrative Biology, ETH Zürich, Zürich, Switzerland

Abstract

Tree planting and natural regeneration contribute to the ongoing effort to restore Earth's forests. Our review addresses how the plant microbiome can enhance the survival of planted and naturally regenerating seedlings and serve in long-term forest carbon capture and the conservation of biodiversity. We focus on fungal leaf endophytes, ubiquitous defensive symbionts that protect against pathogens. We first show that fungal and oomycetous pathogen richness varies greatly for tree species native to the United States (\(n = 0–876\) known pathogens per US tree species), with nearly half of tree species either without pathogens in these major groups or with unknown pathogens. Endophytes are insurance against the poorly known and changing threat of tree pathogens. Next, we review studies of plant phyllosphere feedback, but knowledge gaps prevent us from evaluating whether adding conspecific leaf litter to planted seedlings promotes defensive symbiosis, analogous to adding soil to promote positive feedback. Finally, we discuss research priorities for integrating the plant microbiome into efforts to expand Earth’s forests.

Keywords
plant pathogens, endophytes, biodiversity, plant phyllosphere feedback, climate change, tree planting
To keep every cog and wheel is the first precaution of intelligent tinkering.

—Aldo Leopold (75)

INTRODUCTION

Forests, particularly those found in the tropics, have long been known to be hotspots of biodiversity, inhabited by a wide variety of plants, insects, birds, amphibians, and mammals. More recently, we have come to appreciate the multitude of tree-associated microbes (fungi, bacteria, and archaea inhabiting various tree habitats), their complex interactions, and the mechanism by which they can promote forest biodiversity (8). The best understood and longest studied part of the tree microbiome is its most ecologically important symbiosis: mycorrhizal fungi (63). The ectomycorrhizal (EM) symbiosis is limited to approximately 2% of all plant species (although 60% of Earth’s 3.1 trillion trees), whereas the more ancient, arbuscular mycorrhizal (AM) symbiosis is much more common across Earth’s 60,065 tree species (122). Mycorrhizal symbioses are recognized as being fundamental to plant nutrition and the structure and function of forest soils (7, 63). In contrast, the organismal groups that make up the majority of the tree microbiome, mostly bacteria and fungi, are often characterized either as pathogens or as nonpathogens of uncertain role. Moreover, their identities are frequently represented simply by sequence-based approximations at the genus level or higher. Plant disease protection, conferred by many of these members of the plant microbiome, is increasingly documented, with communities of endophytic microbes in soil, roots, stems, leaves, flowers, and seeds combating or competing with pathogens and priming host immunity (34, 100, 130, 135). Together, this body of work suggests that the functional influence of the microbiome on its tree host is beneficial and protective (82). In this review, we address strategies for integrating diverse tree microbiomes into a massive effort to expand the global forest.

Ongoing tree planting and natural forest regeneration efforts are components of a larger strategy to address interrelated crises aggravated by deforestation. Over the course of human history, we have reduced the number of trees on Earth by half, from six trillion to three trillion (36). In many regions of the world, losses are continuing (121). The loss of forest habitat caused by land-use conversions is a major contributor to Earth’s sixth mass extinction (69), most acutely seen in the tropics (57, 58). These large-scale losses are one crisis. A second crisis exacerbated by deforestation is climate change (96). Deforestation returns carbon stored in tree stems and soil to the atmosphere, contributing to warming (87). Half of all terrestrial carbon stored in vegetation is stored in tropical forests alone (23). Earth’s largest tropical forests are in Amazonia, where a 17% loss in forest is associated with profound, system-wide effects, including regional shifts from carbon sinks to sources (53).

Reforestation is one way to address these interrelated crises. This includes conserving existing forests, promoting natural forest regeneration, and actively planting trees. These efforts are of special urgency for people living near forests, totaling 1.6 billion, or 21% of the current human population (93). Most of these people are in the tropics where the shade of trees offers critical protection against heat waves (91, 119). Many groups are contributing to the massive effort to restore Earth’s forests; examples include the United Nation’s Trillion Tree Campaign, Plant-for-the-Planet, One Tree Planted, Trees for the Future, the Arbor Day Foundation, Trees for Cities, International Tree Foundation, The Woodland Trust, Tree Canada, Casey Trees, Trees for Life, and Trees, Water & People. New groups emerge frequently. In the Netherlands, for example, one group collects volunteer tree seedlings where they are not wanted (e.g., a flower bed or vegetable patch) and gives them away to those who will plant them. To coordinate the many tree-planting efforts, the World Economic Forum established 1t.org (https://www.1t.org/) to “serve the global reforestation community by mobilizing private sector ambition and engagement, accelerating restoration in
priority regions, and empowering a new generation of ecopreneurs to take action on climate change.

In the most basic sense, success in global reforestation hinges on the survival of planted and naturally regenerating tree seedlings. The seedling stage is the most vulnerable in a tree’s life cycle (12, 46, 59), and plants depend heavily on their symbionts to tolerate both abiotic and biotic threats to their existence (139, 147). Plant disease has always been a concerning threat to the establishment of tree plantings (111); this threat is now exacerbated by climate change and the movement of pathogens by human activities (21, 123). Yet with few exceptions, restoration efforts are planting trees without knowledge of, or consideration for, their microbial symbionts, including pathogens. This is problematic given that historically, large-scale tree-planting projects often failed for reasons that involved microbes.

Tree-planting practices are typically borrowed from plantation forestry, which is rooted in tree genetics, silviculture, pathology, and entomology, a worldview in which productivity is prioritized, and microbes are narrowly viewed as pathogens to be avoided. These views persist today, where the microbiome is not considered relevant to forest management (129). For plantation methods that have been developed for a few of the world’s 60,065 tree species (14), success has been achieved by planting trees outside their native range, where their native pathogens do not occur. For example, disease has been avoided by introducing pines of the Northern Hemisphere into plantations of the Southern Hemisphere, Amazonian rubber into southeast Asia, or African oil palm into the neotropics. Alternatively, when trees have been planted within native ranges, plantations can fail due to native pathogens and unnaturally close spacing of the trees; Henry Ford’s failed rubber plantation in the Amazon (i.e., native range of *Hevea brasiliensis*) in the 1920s is one infamous example. Plantations under attack by pathogens are often managed with specific remedies (e.g., genes for resistance) (141) that apply only to a particular tree species, pathogen, and environment. Silvicultural methods of disease control are effective in some cases (20), but here we focus on a more general approach involving microbial symbionts to combat the wide array of pathogens (e.g., parasitic plants, fungi, bacteria, nematodes, phytoplasmas, and viruses) that affect trees globally.

At the other end of the spectrum, an absence of beneficial microbes can also lead to plantation failure. This is because when we plant forests, we rarely think to simultaneously “plant” a tree’s symbionts. Although a tree’s pathogens can be found in its leaves and soil, so are its key mutualists. Early efforts to establish pine plantations in the Southern Hemisphere failed until scientists thought to co-introduce associated mycorrhizal fungi, critical members of the forest microbiome (81). More generally, the importance of actively introducing symbiotic soil organisms to facilitate plant growth in novel landscapes has been known for more than a century (24, 62), and recent work has discovered the importance of soil microbiome restoration for facilitating ecosystem restoration. Native grasses in restored American Midwest prairies grow more vigorously and are more likely to survive when inoculated with the soil microbiome inhabiting intact prairie remnants (17, 71). Grassland restoration on abandoned Estonian mining landscapes could only be achieved by coinoculating native AM fungi (133). Restoration of high-latitude grassland communities could only achieve plant communities similar to those of intact remnants if soil microbial communities were coinoculated at the time of planting (142). Taken together, this body of work suggests an incredible, and largely unrealized, opportunity to enable and accelerate the restoration of Earth’s forests by reintroducing tree microbiomes.

The potential for belowground microbiome restoration to facilitate ecosystem restoration suggests an analogous opportunity within the aboveground forest microbiome given the potential for dispersal of immigrant inoculum from plants or litter nearby. Indeed, beneficial tree symbionts are also found in the phyllosphere but have received far less attention than their belowground
counterparts (86). In particular, fungal leaf endophytes found in all plants can contribute to plant defense against pathogens (29, 107), in addition to influencing various other aspects of plant growth, development, and resilience (102, 113). The mechanisms by which endophytes increase or decrease disease severity in their hosts are complex (e.g., parasitism, antibiosis, host-triggered immunity), but perhaps most simply, by occupying space and competing for resources, these commensal symbionts can exclude pathogens. Once leaves senesce and drop, they form a layer of dead leaves (litter) containing live endophytes beneath the tree. Endophytes complete their life cycle in the leaf litter; they sporulate and send their spores upward to reinfect newly emerging leaves. Thus, leaf litter, like soil, is an important reservoir of beneficial microbes (97, 131).

Given the ambitious goal of restoring Earth’s forests, and the importance of microbial symbionts for tree seedling defense and survival, a key question is what knowledge is needed to integrate the tree microbiome into forest restoration efforts. In this review, we focus on whether our current understanding of interactions between tree pathogens and leaf endophytes can inform low-cost, low-tech ways to promote aboveground defensive symbiosis in reforestation. We also discuss burgeoning efforts to integrate belowground symbionts into planting efforts as well as what success means in terms of carbon and diversity for the long-term. For example, by tolerating slow, initial growth we may successfully reduce the magnitude of the otherwise climate-unfriendly tradeoff between fast initial tree growth and tree longevity (101). Thus, our approach is distinct from the plantation forestry paradigm, which selects for fast, initial growth of single tree species. And our approach is distinct from the agricultural paradigm, which seeks to eliminate pathogens from crop monocultures. Instead, we explore approaches that integrate diverse tree microbiomes, which include pathogens, thereby supporting forest biodiversity, carbon capture, and long-term forest resilience.

CHARACTERIZING PATHOGEN COMMUNITIES OF TREE SPECIES

Understanding the pathogen communities of tree species used in tree-planting campaigns is a first step in managing disease during the vulnerable seedling stage. A common assumption is that all plants are affected by pathogens (79). In fact, a general introduction to plant pathology states that each species of plant appears to be attacked by 50–100 pathogens (3). If this is true, characterizing the pathogens of the world’s 60,065 tree species is a daunting place for reforestation efforts to begin. To probe our knowledge of tree species affected by pathogens, we used the databases of the US National Fungus Collections (BPI) to search for the fungal and oomycetous pathogens of the 1,312 tree species in the United States, given that fungi and oomycetes are the two major classes of plant pathogens (13). These trees were identified using a geographically constrained search of the 60,065 tree species in GlobalTreeSearch (14), where a tree species is defined as “a woody plant with usually a single stem growing to a height of at least two meters, or if multi-stemmed, then at least one vertical stem five centimeters in diameter at breast height.”

Our search of the BPI databases for fungal and oomycetous pathogens of the 1,312 tree species found in the United States showed that pathogen richness varied greatly among tree species, ranging from 0 to 876 known fungal and oomycetous pathogens per US tree species, with nearly half (48%) of tree species lacking known pathogens (Figure 1). In other words, the assumption that all plants, including trees, have pathogens may not be true. Of course, there are some tree species for which pathogens have simply not been studied. But for many of the US tree species without fungal/oomycetous pathogens, evolutionary escape following long-distance dispersal is a plausible explanation (27). For example, most species lacking pathogens are Hawaiian endemics, whose mainland relatives host a diverse array of pathogens. The Hawaiian flora has long been understood as the product of many long-distance dispersal events (103). Those founding events, leading to the
Results of a US National Fungus Collections database search for known pathogens of all tree species found in the United States. Almost half of tree species in the United States lack known fungal and oomycete pathogens, and a significant portion of these are endemic to Hawaii. *Pseudotsuga menziesii* (Douglas fir) has 876 pathogens, which is the most reported of any US tree species. The x-axis indicates the number of known pathogens for each tree species, and the y-axis indicates the quantity of species per pathogen count. The frequency (y-axis) has been log transformed and the bars are overlaid. The blue bars represent species found in the United States overall, except for the Hawaiian endemics. The green bars represent species endemic to Hawaii.

speciation of Hawaiian endemics, have similarities to anthropogenic enemy release expected for introduced plants and their pathogens (88).

Of the 363 tree species native or endemic to Hawaii, 312 (i.e., 86%) may have lost their pathogens, as there are no BPI pathogen records. Many Hawaiian endemics have been studied for pathogens because the diseases they cause threaten small populations. For example, *Cyanea* comprises 85 species endemic to Kauai (114). One such endangered species, *Cyanea kubizewa*, is closely monitored and traps are currently being used to control rats that might damage the remaining two mature individuals, two juveniles, and 11 seedlings (114); we found only one pathogen of *Cyanea* species overall in our BPI search, and it was not affecting *C. kubizewa*. Of the many studies of the ecology and genetics of five species of *Metrosideros* endemic to Hawaii (98), pathology has usually been included (11). In BPI records for *Metrosideros*, we found pathogen richness varied from 0 pathogens for *Metrosideros rugosa* and *Metrosideros waiialealae* to 150 pathogens for *Metrosideros polymorpha*, which occurs across the Hawaiian Islands (11).

A further 324 (of 949, or 34%) non-Hawaiian tree species in the United States are also unaccompanied by pathogen records, suggesting that the phenomenon of natural enemy release is not restricted to Hawaii. Although an extensive analysis of range size relative to pathogen diversity is needed to clarify why some tree species may have escaped or lost their pathogens, it appears that the most widely distributed tree species are accompanied by many pathogens: 180 for *Prunus serotina*, which ranges through eastern North America, 108 for the wide-ranging *Prunus americana*, and 876 for *Pseudotsuga menziesii*, the dominant conifer of the Pacific Northwest. Such widely distributed trees are likely to be included in reforestation efforts, and thus defense against pathogens is crucial for their survival. In contrast, the most isolated tree species with the smallest populations are without pathogens: *Prunus eremophila*, a critically endangered species of perhaps
2,000 individuals native to the Mojave Desert (104), and Prunus fremontii, desert apricot of California and contiguous Mexico.

Disease-protective fungal leaf endophytes might seem to be of little value or concern for trees without pathogens, but pathogen reunions following the introduction of exotic pathogens are always a possibility. Some threatened Hawaiian endemics have been rescued from introduced, but perhaps ancient, pathogens, with endophytes from related plants with more robust populations (144). Indeed, endophyte-mediated approaches in conservation have become a frontier in endophyte ecology, with additional examples from Hawaii (32, 41). Hawaii has also pledged to add 100 million trees by 2030 through a combination of conservation, restoration, and tree planting. Thus, even in an environment where many tree species appear to have escaped their pathogens, endophytes could serve as insurance against pathogen reunions or encounters with novel pathogens (67).

FOLIAR ENDOPHYTES AS DEFENSIVE SYMBIONTS

Low-cost, low-tech solutions are needed in global tree-planting efforts to boost seedling defense against pathogens (21, 111, 123). Here, we explore whether defensive symbionts of the phyllosphere can fill this need, yet we recognize that a plant’s primary mechanisms of defense are genetic. Nonhost resistance, or complete resistance, is conferred by several genetic factors (48). Pathogens cannot infect nonhosts. For known host–pathogen combinations, the next best defense against pathogens is major gene resistance (i.e., gene-for-gene relationship) (47). Plants containing R genes are often selected for cultivation given their protection against disease. This practice has been successful for the few tree species that are cultivated in plantation forestry. But for most of the world’s tree species, breeding programs do not exist and are challenging and expensive to implement given long generation times. Seed for planting efforts could be selected from tree populations where R genes are likely to occur. Indeed, selecting genetically appropriate plant material (within a species) is one strategy for enhancing resilience to biotic and abiotic stressors in tree-planting efforts (125).

When pathogen infection does occur, ubiquitous fungal leaf endophytes can produce robust defensive responses (reviewed by 29, 107), rivaling the contributions of quantitative genetic resistance (108). This body of work suggests that harnessing these defensive symbionts could indeed bolster defense in regenerating forests. Diverse fungal leaf endophytes communities are found in all tree species; they are dominated by Ascomycetes, including ubiquitous taxa like Alternaria and Cladosporium, but also include basidiomycetous yeasts and many rare taxa (9, 60, 99, 132, 148). The incredible fungal diversity found within individual leaves was first discovered using culture-based methods (4, 50) and has been confirmed with culture-free methods (25). The consequences of hosting diverse endophytic communities are an active area of research in plant and fungal ecology. But an early (5) and commonly reported benefit of hosting endophytes is disease protection (reviewed by 29, 107).

Plant disease modification is a central function of many fungal leaf endophytes. These defensive symbionts occur in diverse fungal lineages (145) and have been studied in several tree species (5, 28, 33, 52). Populus has become a model in the study of plant–endophyte–pathogen interactions, with several studies showing that common fungal leaf endophytes reduce the severity of the tree’s major leaf rust pathogen (28, 76, 108). Several of the taxa identified as reducing leaf rust disease, like fungi such as Eudarluca, Cladosporium, and Trichoderma, or bacteria such as Bacillus, are known to antagonize pathogens via mycoparasitism (10, 95) and/or by inducing plant genetic resistance (43). Communities are also protective, with the arrival order of endophytes into the community modulating the level of disease protection (76). At face value, priority effects in community
assembly may seem to suggest that early integration of endophytes into tree-planting efforts will boost their abundance and their efficacy in reducing disease. However, we urge caution before drawing general conclusions, as much remains to be learned about how endophyte interactions impact the functional influence of the microbiome on plant health (61). For example, Leopold & Busby (76) found that downstream differences in plant disease modification resulting from varying endophyte arrival order were not the result of the differential relative abundance of endophytes or shifts in community composition. Instead, early arriving endophytes may have interacted with the plant’s immune system to reduce disease. In another study, simply changing the arrival order of endophytes into a community resulted in a switch from a protective interaction to a pathogenic interaction (1). Together, this work highlights how the composition of the plant microbiome can be decoupled from its functional influence on plant disease, and the challenge of precisely predicting the impact of the tree microbiome in reforestation.

The complex interactions between plants and endophyte communities that result in disease protection are not known for the vast majority of global tree species. However, a variety of mechanisms have been identified. Endophytes can exclude pathogens via competition for space and resources (82). These interactions are likely to occur between endophytes and necrotrophic pathogens given their competition for space and nutrients in the apoplast. However, direct pathogen antagonism via parasitism or antibiosis may be more likely between endophytes and biotrophic pathogens. Endophytes can also trigger host immunity and thereby reduce disease (42, 83). Given the complexity of mechanistic interactions underlying endophyte-mediated disease modification, one might conclude that designing applications that are generalizable across the world’s 60,065 tree species is futile. To the contrary, given that endophyte communities are taxonomically diverse and that endophytes capable of modifying disease occur across diverse fungal lineages, we expect that promoting endophyte symbiosis will result in a diversity of interactions that protect tree seedlings from their pathogens. Such a boost by inoculation may be needed to eliminate a deficit in endophyte diversity in nursery-grown seedlings compared to their naturally regenerated counterparts (51). A positive relationship between endophyte diversity and disease protection could result from complementarity among the effects of many species or a greater chance that the community includes a particularly strong pathogen antagonist (72, 127). Indeed, diversity within phyllosphere bacteria communities was previously shown to be associated with greater disease protection in Arabidopsis (31). Beyond diverse fungal leaf endophyte communities, plants deploy a variety of other above- and belowground defensive symbionts to aid in pathogen defense—e.g., bacterial endophytes (34, 106, 138), viruses (115), mycorrhizae (66, 80), and nematodes (70)—further suggesting that conserving diversity in the microbiome will protect this associated function.

**PHYLLOSHERE FEEDBACK CAN BE POSITIVE**

To explore the potential for fungal leaf endophyte communities to serve as defensive symbionts in regenerating forests, we searched the literature for studies of plant phyllosphere feedback. This is an emerging area of research that adapts the experimental framework from plant soil feedback, builds on considerable work with nurse plants and mulch in forest restoration (19), and asks questions about how foliar microbes may either support or hinder the performance (growth, defense, etc.) of conspecific seedlings (140). These feedbacks can be tested by inoculating aboveground plant tissues with microbes sourced from the phyllosphere (synthetic communities or bulked from litter) and comparing plant performance relative to either uninoculated controls or those inoculated with microbes sourced from heterospecific litter. Likewise, plant soil feedback experiments investigate these interactions with soil-inhabiting microbial communities by
examining the extent to which belowground communities facilitate or inhibit growth of con-
specific seedlings (18). Past research has shown that host-specific soil pathogens can accumulate
and suppress seedling growth in conspecific soils (16, 35, 68, 78), whereas mycorrhizae facilitate
positive feedbacks in conspecific soils (35). In some cases, endophytic bacteria (Pseudomonas spp.)
in roots directly antagonize root pathogens of forest trees linked by the mycorrhizal network
(39). However, rarely are phyllosphere microbes considered in feedback studies.

In a strict sense, a plant phyllosphere feedback study is one in which microbes sourced from the
phyllosphere, rather than the leaf litter in its entirety, are applied to experimental plants. This dis-
tinction ensures that any treatment effect on plant performance can be attributed to the microbes
rather than to other factors associated with leaf litter that have been shown to influence seedling
fitness, such as metabolites or shading (136, 137). Unfortunately, our literature search identified
very few published plant phyllosphere feedback studies that fit our definition, and no studies fo-
cusing on tree species or pathogen protection (Table 1). All experiments used Asteraceae plant
hosts and found that microbes sourced from conspecific litter reduced seedling fitness relative to
heterospecific litter (45, 140, 146). Plants that are phylogenetically or physiologically different
from those used in these studies, like tree species, may exhibit a different feedback strength or
type.

Other plant phyllosphere feedback studies, although not strictly fitting our definition as stated
above, used low-tech protocols for applying phyllosphere microbes and could thus serve as models
for reforestation efforts. Such studies diverged from our definition in that the inoculum source was
litter in its entirety (rather than microbes sourced from litter), or the experiment did not include
conspecific inoculum. Although several of these studies have demonstrated conspecific litter in-
hibiting the growth of seedlings (6, 77), others suggest circumstances in which plant phyllosphere
feedback is positive. For example, a recent study found that conspecific litter of the common
ragwort, Jacobaea vulgaris, reduced seed germination rates yet increased growth for established
seedlings (90). These findings imply that the way plants interact with microbial communities varies
by life stage. Litter microbes have also been found to provide plant hosts with disease protection.
In one example, the host was a conspecific tree seedling that had been inoculated with conspecific
litter (33), whereas, in another, the host was an agricultural crop and the litter inoculum had been
sourced from a nearby pine forest (110). Overall, these studies highlight context dependency in
plant phyllosphere feedback. Experiments have the potential to play an integral role in informing
how to best harness the aboveground microbiome in planting efforts. However, to do so effect-
ively, these experiments will need to replicate the context in which the litter microbes are to be
applied.

Douglas fir (P. menziesii), a dominant conifer of the Pacific Northwest of the United States
(109), will be a focal species used in a reforestation project in Willamette National Forest, Ore-
gon. This ready application makes P. menziesii an ideal candidate for a plant phyllosphere feedback
experimental case study that could inform the use of leaf litter as a source of microbial inoculum
for seedlings in this tree-planting project. In a greenhouse inoculation experiment, we found that
coastal Douglas fir (P. menziesii var, menziesii) seedlings inoculated with microbes sourced from
Douglas fir litter (moist-incubated to enhance fungal sporulation) and applied directly to needles
displayed greater growth than uninoculated control plants (A.S. Neat, F.A. Jones, J. LaManna,
K.A. Gervers, P.E. Busby, unpublished data). However, feedback varied between two tree popula-
tions: the upper-elevation population benefited from phyllosphere microbes, whereas the lower-
elevation population was unaffected. Similarly, the forestry literature reports coastal Douglas fir
(P. menziesii var. menziesii) seedlings perform best on bare mineral soils, whereas Rocky Mountain
Douglas fir (P. menziesii var. glauca) seedlings benefit from a litter layer (65). Although these ex-
amples illustrate the benefit of litter microbes for seedling growth and their inexpensive and easy
Table 1  Summary of methods and findings for published phyllosphere feedback studies

<table>
<thead>
<tr>
<th>Plant species</th>
<th>Plant type</th>
<th>Life stage</th>
<th>Experimental design</th>
<th>Inoculum</th>
<th>Inoculum source</th>
<th>Response to inoculum</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Triticum aestivum</em></td>
<td>Grass (crop)</td>
<td>Seedling</td>
<td>Greenhouse</td>
<td>Foliar microbes</td>
<td>Nearby pine forest</td>
<td>Pathogen protection</td>
<td>110</td>
</tr>
<tr>
<td><em>Ageratina adenophora</em></td>
<td>Herbaceous</td>
<td>Seed, seedling</td>
<td>Laboratory</td>
<td>Foliar microbes</td>
<td>Conspecific plant material</td>
<td>Reduced germination rate, reduced growth</td>
<td>45</td>
</tr>
<tr>
<td><em>Jacobaea vulgaris</em></td>
<td>Herbaceous</td>
<td>Seed, seedling</td>
<td>Greenhouse</td>
<td>Direct litter exposure</td>
<td>Conspecific or heterospecific plant material</td>
<td>Reduced germination rate, increased growth from conspecific exposure</td>
<td>90</td>
</tr>
<tr>
<td>Asteraceae spp.</td>
<td>Herbaceous</td>
<td>Seedling</td>
<td>Greenhouse</td>
<td>Foliar microbes</td>
<td>Conspecific or heterospecific plant material</td>
<td>Reduced growth from conspecific exposure</td>
<td>140</td>
</tr>
<tr>
<td>Asteraceae spp.</td>
<td>Herbaceous</td>
<td>Seedling</td>
<td>Greenhouse</td>
<td>Foliar microbes</td>
<td>Conspecific or heterospecific plant material</td>
<td>Reduced growth from conspecific exposure</td>
<td>146</td>
</tr>
<tr>
<td><em>Theobroma cacao</em></td>
<td>Tree</td>
<td>Seedling</td>
<td>Field</td>
<td>Direct litter exposure</td>
<td>Conspecific plant material</td>
<td>Pathogen protection</td>
<td>33</td>
</tr>
<tr>
<td><em>Fagus sylvatica, Picea abies</em></td>
<td>Tree</td>
<td>Seed</td>
<td>Field</td>
<td>Direct litter exposure</td>
<td>Conspecific or heterospecific plant material</td>
<td>Reduced germination rate from conspecific or heterospecific exposure</td>
<td>6</td>
</tr>
<tr>
<td><em>Machilus leptophylla, Litsea elongata, Acer pubinerve, Distylium myricoides</em></td>
<td>Tree</td>
<td>Seedling</td>
<td>Field survey</td>
<td>Direct litter exposure</td>
<td>Conspecific or heterospecific plant material</td>
<td>Reduced seedling survival from conspecific exposure</td>
<td>77</td>
</tr>
<tr>
<td><em>Pseudotsuga menziesii</em></td>
<td>Tree</td>
<td>Seedling</td>
<td>Greenhouse</td>
<td>Foliar microbes</td>
<td>Conspecific or heterospecific plant material</td>
<td>Increased growth from conspecific exposure</td>
<td>A.S. Neat, F.A. Jones, J. LaManna, K.A. Gervers, P. Busby, unpublished data</td>
</tr>
</tbody>
</table>

*These studies were selected because the host plants were inoculated with microbes sourced from litter or directly exposed to litter in an experiment designed specifically to focus on the influence of litter microbes on host fitness (rather than other qualities of litter, such as shading).
application, they also highlight complexities in outcomes that are not currently understood. Thus, even for one of the most well-studied tree species in forestry, knowledge gaps prevent us from answering the question of when, where, and how adding conspecific leaf litter to planted seedlings promotes defensive symbiosis. Below, we highlight research priorities that can accelerate our understanding of the role of diverse microbiomes in tree performance, defense, and resilience as well as our ability to optimize these benefits in global forest restoration.

THE IMPORTANCE OF INTEGRATING PLANT AND MICROBIAL DIVERSITY INTO GLOBAL FOREST RESTORATION

Given the urgency in responding to the climate and biodiversity crises, and the known problems associated with planting tree species outside their native ranges, it will be necessary to expand tree populations in situ where their pathogens commonly occur. An alternative to the plantation forestry paradigm is needed, one that integrates both local tree species diversity and microbial diversity. The importance of tree diversity (e.g., plantations of species mixtures) has received some attention in forestry and conservation (2, 68, 106). In this review, we have focused on how tree microbiome diversity can promote pathogen defense in regenerating forests. In addition, transplanting the tree microbiome should also promote forest biodiversity broadly (86). For example, at the scale of a forest ecosystem, tree pathogens are not necessarily negative. Specialized, host-specific pathogens diversify plant communities by attacking only their hosts, thus providing a local advantage to their hosts’ competitors (15). Pathogens can also diversify forest communities via habitat formation, e.g., by creating witches’ brooms and stem cavities for birds and mammals (126). Forest endophytes can promote plant diversity by acting as commensal symbionts on their host plants but pathogens on their hosts’ competitors (i.e., apparent competition) (56, 92). Thus, just as a tree seedling will suffer if it is not transplanted with its associated microbes, a forest cannot be truly restored without its microbiome.

Plantations are well known to lack the biodiversity found in nearby forests (22). Similarly, arboretums illustrate how tree planting without consideration for microbial diversity can fail to become forests. For example, the University of Idaho’s 100-year-old arboretum is superficially like the natural forests of Moscow Mountain 16 kilometers away—both contain a mixture of mature coniferous and deciduous tree species. The founding goal of the arboretum was to introduce and observe exotic tree species. The pervasive consequences of that goal are striking. The obvious difference is that there are non-native tree species in the arboretum and native trees in the natural forest; less obvious is the recapitulation of this pattern by understory flora (30). In addition, the mycofloral diversity of the arboretum is lacking compared to the natural forest. Common pathogens of native trees such as dwarf mistletoe of Douglas fir and decay fungi such as species of Fomitopsis are also absent from the arboretum. In contrast, non-native pathogens of non-native trees (e.g., powdery mildew of Acer platanoides) and herbaceous plants (e.g., an oomycetous pathogen of Lunaria annua) are present. The contrast between plantings and forests reminds us that tree plantings should not only remove carbon from the atmosphere but also conserve biodiversity. Symbionts, with their interactions and processes, make plantings into forests.

The idea of promoting biodiversity to benefit ecosystem health and productivity is not new. In the mid-nineteenth century, Darwin hypothesized that diverse plant communities were more productive than species individually (37). The relationship between plant species diversity and ecosystem productivity was later confirmed (128) and then shown to be driven by microbes (116). In forestry, mvuca de sementes (Portuguese for mixture of seeds) in the Amazon and the Japanese method developed by the late Akira Miyawaki are two examples in which tree diversity seeks to enhance forest biodiversity (89). Miyawaki was also an early advocate of integrating mycorrhizae...
A tree-planting site. Plant-for-the-Planet trial, Yucatán Peninsula of México. (a) 16,000 seedlings were planted in a randomized block design situated on an abandoned cattle pasture in a Mexican semi-evergreen tropical forest in 2020. The blocks were paired with natural regeneration control plots. Survival and growth rates are measured to track response. (b) Planting ~50-cm tall seedlings of eight native, arbuscular mycorrhizal fungi–associated tree species. Half of the planting holes were inoculated with 500 mL of soil from a nearby forest in which the same species frequently occur to accelerate restoration by reintroducing symbionts. The other half of the seedlings were planted without inoculum.

into forest restoration to enhance seedling growth and survival, a practice now recognized in grassland restoration (17, 71, 142). Also, ongoing, large-scale field trials in Wales, UK and the Yucatán Peninsula of México are testing the idea that restoration of both trees and a complex soil microbiome from nearby intact forests can facilitate tree growth and survival. For example, in a Plant-for-the-Planet trial in the Yucatán, seedlings of eight native, AM fungi–associated tree species are being planted with or without 500 mL of soil from a nearby forest in which the same species occur (Figure 2). This treatment aims to accelerate restoration by reintroducing fungal symbionts. If positive microbial restoration–ecosystem function outcomes are strong, they could form the foundation of a microbiome restoration industry built on top of emerging forest carbon and biodiversity markets. However, as we work to incorporate the microbiome into a more holistic picture of ecosystem restoration, it is critical that introduced microbes are complex, diverse, and sourced from local populations. Doing so represents the best chance to rebuild and maintain the incredible biodiversity of microbial life inhabiting Earth’s forests.

RESEARCH PRIORITIES FOR FACILITATING REFORESTATION THROUGH THE PLANT MICROBIOME

We have illustrated knowledge gaps in our understanding of tree pathogens and endophytes that raise questions about how to best promote microbiome-mediated defense against pathogens during a tree’s most vulnerable stage of development and beyond, as trees age and face an unpredictable array of abiotic and biotic challenges (101). These gaps exist because plant–microbiome interactions have traditionally been studied in model systems (e.g., Arabidopsis) and crop plants, with objectives that are about neither conservation of biodiversity nor carbon removals from the atmosphere. Thus, tree-planting and natural forest regeneration projects must pioneer new opportunities to study tree microbiomes as they broadly relate to host health across a diverse array of tree species. Our overarching hypothesis is that conserving the tree microbiome in planting
efforts will enhance tree seedling defense and survival and ultimately the success of forest restoration in terms of carbon capture, forest biodiversity, and long-term forest resilience. The research priorities described below provide a framework for testing this hypothesis.

**Identifying Causal Factors that Give Rise to Variation in Tree Microbiome Composition and Its Functional Influence on Tree Seedlings**

How do the benefits of hosting a diverse microbiome vary among tree species and environments? Many studies have documented variation in the plant microbiome across host species, guilds (e.g., EM versus AM, evergreen versus deciduous), developmental stages, trait values, and along complex environmental gradients (e.g., temperature, moisture, elevational, latitudinal gradients) (38). For example, variation among tree hosts (73, 134) and along climatic gradients (9, 55, 148) influences microbial composition in the phyllosphere. Yet how such variation, in turn, influences the functional influence of the microbiome on its host is poorly understood. Functional convergence among compositionally distinct soil microbial communities has been shown at continental scales in *Pinus* forests (124). However, plant–soil feedback can vary along environmental gradients (74), with particular microbes enabling plant survival in extreme environments (i.e., habitat-adapted symbiosis) (112). Given the urgency in global forest restoration, we must learn as we go, experimentally manipulating hosts and microbiomes across regenerating forests to determine optimal parameters for maximizing success. At the same time, we must also continue to learn from intact forests, asking questions such as whether endophytic communities of tree species vary along gradients of forest age and diversity. Formally recognizing thousands of undescribed species in this effort may be necessary for consideration in conservation. Do communities of endophytic bacteria and fungi constitute an extension of the tree defense system provided by mycorrhizal networks (120) or a separate system? Collaboration among forest ecologists, microbiologists, pathologists, and tree planters is essential for making progress on this and other research priorities.

**Determining How Planting and Transplanting Techniques for Both Seedlings and Associated Microbes Influence the Benefit of the Microbiome to its Host**

What planting and transplanting practices best promote above- and belowground tree symbionts? How well do transplanted microbes establish and persist in forest restoration sites? How do the age of the intact forest and its distance to planted or naturally regenerated tree seedlings influence benefits derived from the forest's mycorrhizal network? In native prairie restoration, the benefit of inoculating plants with AM fungi extended to uninoculated plants two meters away (84, 85). Does it make a difference if the network is EM or AM (118)? Comparisons of agricultural fields and intact forests have revealed little diversity of Glomales (AM forming) in arable land and more diversity in the forest soil (64). If older forest soil is used as inoculum, spores and other propagules must be present and viable; fortunately, fungal propagules remain viable for years (94). As with the first research priority, these questions can be addressed in the context of ongoing restoration efforts with effective collaboration between scientists and restorationists.

Transplanting the phyllosphere microbiome involves additional considerations. Can we transplant the phyllosphere microbiome as readily as we transplant the soil microbiome? Fungal leaf endophytes have been applied individually to bolster plant defense. For example, seedlings are inoculated with *Beauveria bassiana*, an endophyte of *Pinus monticola* (51) and an entomopathogenic member of Cordycipitaceae, to protect against insects. And seedlings are commonly inoculated with species of *Trichoderma*, common antagonistic endophytes of many tree species, to protect against a variety of pathogens (26). However, even in the few well-studied systems in which good candidates for inoculation are known, there are logistical obstacles to generating and applying
inoculum. Isolating and applying an individual endophyte require technical skills and access to specialized equipment, whereas diverse leaf litter can be applied by anyone given simple instructions. Some evidence for disease protection in tree seedlings treated with conspecific litter (33), in combination with the benefits of mulching (19), suggests that planting tree seedlings with conspecific litter is a tractable method for conserving phyllosphere microbial diversity in tree-planting efforts. Litter can be transplanted to naturally regenerating tree seedlings, which may help to accelerate a process that is often viewed as passive (143). Yet these are hypotheses that require testing in ongoing reforestation efforts.

Identifying Keystone Microbes in the Forest Microbiome

The tree microbiome is a part of the broader forest microbiome, i.e., microbes associated with trees but also with other forest habitats like soil, snags, rocks, insects, mammals, birds, lichens, etc. Successfully restoring forests, with their diverse organisms, processes, and interactions, hinges on restoring the forest microbiome as a whole, including pathogens. But how do we do this? The renowned forest restorationist Payeng, or “Forest Man” (54), swore by his practice of transplanting tree seedlings with worms, ants, termites, and other soil builders. Starting on barren ground and with no professional credentials, Payeng single-handedly built a forest on his island in the Brahmaputra River, India, that now attracts diverse megafauna like tigers and elephants. On the other side of the world, in the US Pacific Northwest, live Douglas fir trees and snags, and therefore their associated microbiomes and necrobiomes, are retained after cutting to accelerate forest development (49). Retaining live trees and their associated mycorrhizal networks is now well recognized as benefiting forest regeneration (117). But what are other keystone forest microbes? Identifying critical forest microbes and associated processes will allow us to prioritize elements of the forest microbiome for conservation.

Determining the Relationship Between Forest Microbiomes and Greenhouse Gasses Beyond Carbon Dioxide

Forest microbes play a critical role in carbon capture. Yet we know little about how forest microbes influence greenhouse gasses other than carbon dioxide, e.g., nitrous oxide (a greenhouse gas 300 times more potent than carbon dioxide). Urea is applied in agriculture to compensate for limited nitrogen, but this can lead to an increase in atmospheric nitrous oxide. An early study showed that excluding EM fungi increased nitrous oxide emissions (44). More recent work, with both EM and AM fungi, has shown that these symbionts can reduce emissions of nitrous oxide (123, 125). Additional studies on both nitrous oxide and methane emissions from forest trees are needed, with the contributions of specific microbes addressed (105).

CONCLUSION

A growing body of work supports the notion that the functional influence of the microbiome on its tree host is beneficial and protective. Yet the full extent of microbial interactions and their potential contributions to global reforestation are not clearly understood (86). We identified major knowledge gaps by reviewing the pathogen status of US tree species, and the functionality of phyllosphere microbes. Even for the world’s most simple forest—Pando, the 47,000-stem aspen monoculture in central Utah that is the world’s largest and oldest organism (40)—the tree microbiome is largely unstudied. Forest microbiologists and ecologists will play an important role in advancing the science of forest microbiomes as we begin the Decade of Ecosystem Restoration called for by the United Nations. With variation in new forests ranging from monoclonal
plantations to highly diverse Miyawaki forests and naturally regenerated or surviving old forests, there is ample variation to study and understand. From tree species without pathogens and trees whose pathogens have yet to be described to the functions of phyllosphere microbes in widely ranging species with hundreds of known pathogens, we have a great deal to learn. We will need to consider the presence or absence of genes for resistance, particular symbionts that protect their hosts, the functions of endophytes in trees lacking pathogens, and the overall contribution of the forest microbiome to ecosystem function. We may discover ways to better protect seedlings and young saplings with low-tech, cost-effective exposures to leaf litter. We may find that greater initial diversity shortens the period that new plantings spend as carbon sources before becoming carbon sinks. Above all, we must engage in this urgent effort to restore Earth's forests.

DISCLOSURE STATEMENT
The authors are not aware of any affiliations, memberships, funding, or financial holdings that might be perceived as affecting the objectivity of this review.

ACKNOWLEDGMENTS
We are grateful to Devin Leopold and Andy Jones for comments on an earlier version of this manuscript. This work was supported by the National Science Foundation under the LTER Grant LTER8 DEB-2025755 (2020–2026), the US Department of Energy's Office of Biological and Environmental Research (DE-SC0019435), and the US Department of Agriculture National Institute of Food and Agriculture (2020-67013-31797). Any opinions, findings, and conclusions or recommendations expressed in this material are those of the authors and do not necessarily reflect the views of the National Science Foundation.

LITERATURE CITED
82. McLaren MR, Callahan BJ. 2020. Pathogen resistance may be the principal evolutionary advantage provided by the microbiome. Philos. Trans. R. Soc. B 375(1808):20190592
Contents

Point-of-Care DNA Amplification for Disease Diagnosis and Management  
José R. Botella ................................................................. 1

Going Viral: Virus-Based Biological Control Agents for Plant Protection  
Jeroen Wagemans, Dominique Holtappels, Eeva Vainio, Mojgan Rabiei, Cristina Marzachi, Salvador Herrero, Mohammadhossein Ravanbakhsh, Christoph C. Tebbe, Mylène Ogliastro, María A. Ayllón, and Massimo Turina ........ 21

Rooting Out the Mechanisms of Root-Knot Nematode–Plant Interactions  
William B. Rutter, Jessica Franco, and Cynthia Gleason ................................................. 43

The Phloem as an Arena for Plant Pathogens  
Jennifer D. Lewis, Michael Knoblauch, and Robert Turgeon ........................................... 77

Peptide Effectors in Phytonematode Parasitism and Beyond  
Melissa G. Mitchum and Xunliang Liu ................................................................. 97

Yellow Dwarf Viruses of Cereals: Taxonomy and Molecular Mechanisms  
W. Allen Miller and Zachary Lozier ................................................................. 121

Recognition and Response in Plant–Nematode Interactions  
Shabid Siddique, Alison Coomer, Thomas Baum, and Valerie Moroz Williamson ........ 143

Pathogen Adaptation to the Xylem Environment  
Leonardo De La Fuente, Marcus V. Merfa, Paul A. Cobine, and Jeffrey J. Coleman .... 163

Exploring the Emergence and Evolution of Plant Pathogenic Microbes Using Historical and Paleontological Sources  
Carolyn M. Malmstrom, Michael D. Martin, and Lionel Gagnevin ..................................... 187

Diversity, Evolution, and Function of Pseudomonas syringae Effectoromes  
Cedoljub Bundalovic-Torma, Fabien Lonjon, Darrell Desveaux, and David S. Guttman ................................................................. 211
Molecular Interactions Between *Leptosphaeria maculans* and *Brassica* Species
*M. Hossein Borban, Angela P. Van de Wouw, and Nicholas J. Larkan* 

Future of Bacterial Disease Management in Crop Production
*Anuj Sharma, Peter Abrahamian, Renato Carvalho, Manoj Choudhary, Mathews L. Paret, Gary E. Vallad, and Jeffrey B. Jones* 

Ecology of Yellow Dwarf Viruses in Crops and Grasslands: Interactions in the Context of Climate Change
*Jasmine S. Peters, Beatriz A. Aguirre, Anna DiPaola, and Alison G. Power* 

Mycovirus Diversity and Evolution Revealed/Inferred from Recent Studies
*Hideki Kondo, Leticia Botella, and Nobuhiko Suzuki* 

Facilitating Reforestation Through the Plant Microbiome: Perspectives from the Phyllosphere
*Posy E. Busby, George Newcombe, Abigail S. Neat, and Colin Averill* 

Climate Change Effects on Pathogen Emergence: Artificial Intelligence to Translate Big Data for Mitigation
*K.A. Garrett, D.P. Bebber, B.A. Etherton, K.M. Gold, A.I. Plex Sulá, and M.G. Selvaraj* 

Exploring Soybean Resistance to Soybean Cyst Nematode
*Andrew F. Bent* 

Errata

An online log of corrections to *Annual Review of Phytopathology* articles may be found at http://www.annualreviews.org/errata/phyto