

ARTHROPODS ASSOCIATED WITH VARIOUS AGE STANDS
OF DOUGLAS-FIR FROM FOLIAR, GROUND, AND AERIAL STRATA

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ABSTRACT

The arthropod community in a Douglas-fir forest near Blue River, Lane County, Oregon was investigated from March through August 1973. Five stands of Douglas-fir were sampled: two clearcuttings, a young-growth (20 years old), a midgrowth (125 years old), and an old-growth (450 years old) plot. A pole pruner was used to sample the foliar fauna on the trees in the young-growth plot. The ground fauna was sampled by pitfall traps, and the aerial fauna by rotary nets in each of the stand types. Relative abundances and general trophic categories and species lists for each strata are presented with comment. Trapping method efficiencies are discussed in light of the results of this study and data presented in the literature.

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INTRODUCTION

The terrestrial arthropod investigation of the Coniferous Forest Biome in 1973 was a generalized survey of the major arthropod groups associated with various age stands of Douglas-fir, Pseudotsuga menziesii (Mirb.) Franco. Surveys in the clearcut stands, 20, 125, and 450 years old, were standardized to obtain relative abundances.

Three methods were used to sample numbers of macroarthropods in selected forest strata.

1. Branches of young Douglas-fir trees were captured and pruned, and their attendant fauna collected.
2. Pitfall traps sampled the ground macroarthropod fauna.
3. Rotary nets sampled the aerial fauna 1.83 m aboveground.

Sampling was carried out weekly from March 27 to August 10, 1973. Each arthropod collected by the various sampling methods was stored in alcohol and identified later in the laboratory. Each sample received both a standardized sample number designating sampling method and plot, and a standardized week number following the scheme of Lewis and Taylor (1967). Original identifications were usually to family level, but generic or species determinations were obtained for most of the Coleoptera, Homoptera, and Hemiptera. The arthropods collected during this study are now located in the Entomology Collection, Department of Entomology, Oregon State University and work is presently underway to curate and further identify the arthropods.

Trophic Categories

The general trophic categories are based on the presumed mode of feeding of the life stage in which an arthropod was captured. An exception to this classification occurred with many wasp families, such as Ichneumonidae and Chalcididae. These were captured in the adult stage and classified as parasitic, although the adult was likely a nonfeeder, a nectar feeder, or perhaps a partial feeder on the juices of the insect on which it oviposits. Among particular groups of flies whose feeding habits differ according to sex, a 1:1 sex ratio was assumed for purposes of trophic tabulation. Thus, half the numbers might be assigned to parasites (as in blood feeders) and the other half to phytophages (nectar feeders) or unknown feeders. The trophic tabulation, then, is preliminary and often based on presumed feeding habits of whole families. The general categories recognized are:

Phytophages: Defoliators, sapsucking forms, and nectarivores feed chiefly on any portion of a plant.

Carnivores: Feeders on live animal material. This category was subdivided into predators and parasites. Parasitoids, ectoparasites, and bloodsuckers were considered parasites.

Scavengers: Fungivores, omnivores, and opportunistic feeders, feeding on dead or decaying plants or animals.

Ants: Many species of ants are specific to a particular food source. Because of their often unique community position and

inadequate species indentifications, however, they are treated separately.

Nonfeeders or Unknown Feeders: This includes arthropods that are characterized by one or more of the following conditions: They appear not to feed in the life stage sampled; their feeding habits are unknown; they belong to families with such diverse feeding habits (depending on the actual species involved) that no attempt was made to place it in an arbitrary category.

Site Description

Sampling was conducted in the H. J. Andrews Experimental Forest (recently established as an Experimental Ecological Reserve) in Oregon's western Cascade Mountains (lat. 44°13' N; long. 122°10' W.). Five plots, representing four stand ages of Douglas-fir, were sampled.

Plot 1, a 20-year-old stand at 610-m elevation, was the young-growth plot in this study. It is a low, flat, parkland area that gradually sloped northward from 0 to 25°, along the southern bank of Lookout Creek. The site was clearcut in 1951 and reforested in 1953. When sampled, tree density was estimated as 720 trees/ha. Trees were about 10 m tall. Tree density on steeper slopes averaged 916/ha, and density on the lower, more open area was about 532/ha. This plot was crisscrossed with burned slash, and the understory was dominated by Ribes sanguineum Pursh and Epilobium angustifolium L.

Plot 2, a 125-year-old stand of pole-size second-growth Douglas-fir (Dyrness and Hawk 1972), was the midgrowth plot. It was located on a hill at a higher elevation (975 m) than the other plots and was not affected by climate associated with creekside proximity and valley-floor air movement. Trees were about 30 m tall with an open understory, except for patches of Rhododendron macrophyllum G. Don, Vaccinium parvifolium Smith, Rubus ursinus Cham. and Schlecht., and Berberis nervosa Pursh. The litter layer on the forest floor was thin and composed primarily of needles, small twigs, and branches broken from the trees. Snow cover was more persistent because of the higher elevation. The site was clearcut in 1974, the year after sampling.

Plot 3, a 450-year-old stand along the southern bank of Lookout Creek at 474-m elevation, was the old-growth plot. Tsuga heterophylla (Raf.) Sarg. trees were scattered in the clearings. Acer circinatum Pursh, Gaultheria shallon Pursh, Berberis nervosa Pursh, and Rhododendron macrophyllum G. Don were the dominant shrubs and Linnaea borealis L., Achlys triphylla (Smith) DC., Oxalis oregana Nutt., and Polystichum munitum (Kaulf.) Presl the dominant herbs. Much moisture was retained by the thick litter and moss layer in this plot.

Plot 4, along the southern bank of Lookout Creek, and Plot 5, along the northern bank of McRae Creek, at 610-m elevation, were level clearcut plots. Despite reforestation, few replanted Douglas-fir trees survived. Acer circinatum Pursh, Epilobium angustifolium L., and Rubus ursinus Cham. and Schlecht., trailed over burned slash, bareground, and protruding rocks.

The open and exposed nature of these plots resulted in higher temperatures and lower humidity than in the more forested plots.

Air temperatures were recorded in Plots 1, 2, and 4 with a sheltered Yellow Springs hygrothermograph located 1 m aboveground. Plot 4 showed the greatest extremes in mean weekly maximum and minimum temperatures, followed by Plot 1 and Plot 2 (Figure 1). Lack of cover in Plot 4 allowed a great deal of reradiation. The low maximum temperatures in Plot 2 were due to the higher elevation and greater cover from the tall midgrowth trees. The minimum temperatures at the lower elevation of Plots 1 and 4 were affected by cover (or lack of it) and the effects of cold-air drainage associated with the location in the Lookout Creek canyon bottom. Thus, the minimum temperatures experienced in the young-growth and clearcut plots were usually lower than those of the midgrowth plot at a higher elevation. The temperatures recorded in the three plots responded similarly to cloud cover and precipitation (Figures 1 and 2).

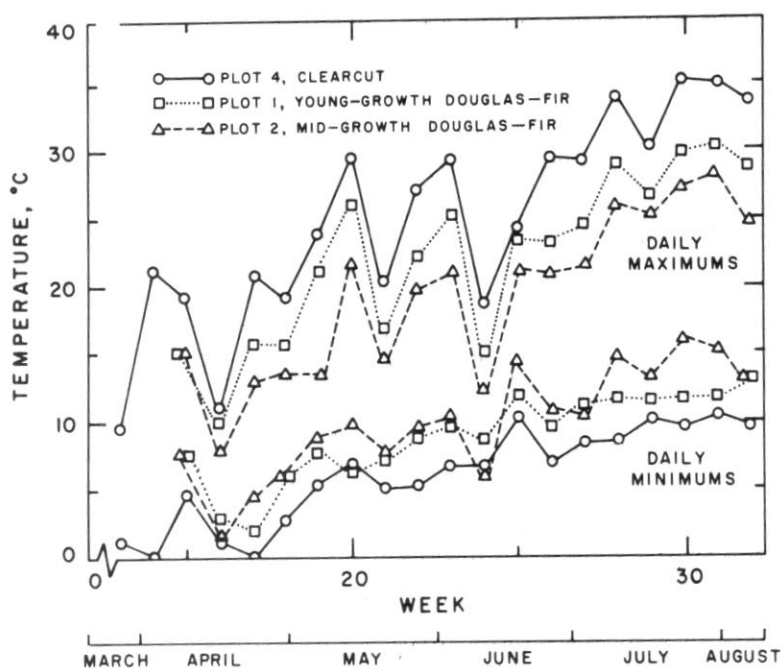


Figure 1. Mean maximum and mean minimum air temperatures 1 m aboveground in Plots 1, 2, and 4 from March through August 1973 in the H. J. Andrews Experimental Forest, Oregon.

Precipitation was recorded by the U.S. Forest Service at a station adjacent to Plot 3 above Lookout Creek. The amount of precipitation in 1973 was lower than normal for this area. Only 167 cm of precipitation were recorded from October 1972 to September 1973 compared to 307 cm the previous year and 271 cm the year before. Nevertheless, the month of June 1973 received twice as much precipitation as usual (8.6 cm compared to an average of 4.3 cm).

Methods

Three branch samples about 1 m long were sampled each from five different trees each week. Weekly densities of arthropods captured were estimated from the 15 branch samples. Each branch was weighed and measured as an

indication of live weight and swept or surface area of the foliage (Mason 1970). Sample sites on each tree were located 3.5-6.0 m aboveground, a height limitation imposed by the operating capabilities of the pruner assembly. Lower branches were sampled first to prevent disturbance of the foliage and arthropods higher in the tree. The three samples from each tree were representative of all sides of the tree and were assumed to be characteristic of the arthropod populations at the midcanopy level (I. W. Varty, personal communication). Once the selected branch had been caught in the net, the net was closed, the branch clipped, and the device lowered to the ground. The enclosed branch was vigorously beaten and shaken inside the net to dislodge clinging arthropods before the branch was removed, weighed, and measured. No search was made for internal branch or needle arthropods. Many of the smaller arthropods, such as Collembola, mites, and smaller adelgids, probably fell through the 0.5-mm² mesh, causing underestimated densities of smaller arthropods.

Densities of arthropod populations on the Douglas-fir branches were estimated by determining the number of arthropods collected per weight and per swept or surface area of sampled branches. Wet weight was recorded in grams, but foliage area was determined by multiplication of length by maximum width of each branch sampled. Based on the destructive sampling of two trees in Plot 1 and the estimated density of 720 trees/ha, the live weight of foliage in this plot was estimated at 50.34×10^6 g/ha, and the total foliage area was estimated at 1056.35×10^6 cm²/ha. The ratio of the total foliage weight/ha to the total weight of the fifteen sampled branches each week yielded a weekly foliage weight multiplier. This multiplier times the number of arthropods of each species or taxon collected each week gave the population density estimates. Likewise, a weekly foliage area multiplier was derived, which was multiplied by the number of arthropods collected, to yield a population density estimated from foliage area sampled rather than weight. The mean of the two estimates based on foliage weight and foliage area was used as the density estimate in this study.

Results and Discussion

Species Composition. Appendix 1 lists the number of specimens of each species or taxon collected on Douglas-fir. The total of 3,767 arthropods collected over the 20-week sampling period was composed mainly of Hemiptera (Heteroptera and Homoptera), which made up 57.5% of the arthropods (Table 1). Coleoptera made up nearly 11%, and spiders (Araneida) 13.5%. The composition of captures on young Douglas-fir are compared with those from a red pine plantation in Ontario (Martin 1966) (Table 1). Considering that the pruning method did not effectively sample smaller arthropods and that Martin's insecticidal technique possibly overestimated the Diptera actually on the trees, the faunal compositions are relatively similar. Exceptions are the low number of Psocoptera and higher relative numbers of Hemiptera on Douglas-fir as compared to red pine.

Table 1. Comparison of arthropods collected on 20-year-old Douglas-fir by pole pruner, and arthropods collected on red pine by Martin (1966) with an insecticidal spray technique.

	Douglas-fir		Red pine	
	No.	% of total	% of total	% of total ^a
Noninsecta				
Araneida	510	13.5	4.3	6.9
Acarina	16	0.4	30.8	-
Insecta				
Collembola	7	0.2	10.1	-
Hemiptera	2,165	57.5	22.7	36.1
Heteroptera	74	2.0		
Homoptera	2,091	55.5		
Psocoptera	174	4.6	9.9	15.8
Coleoptera	399	10.6	2.3	3.7
Neuroptera	63	1.7	1.1	1.8
Lepidoptera	19	0.5	0.5	0.8
Diptera	232	6.2	14.2	22.6
Hymenoptera	172	4.6	3.5	5.6
Others	10	0.3	0.9	1.4
Total Arthropods	3,767			

^aPercentage without inclusion of Acarina and Collembola.

Table 2 shows a preliminary trophic composition for the arthropods found on Douglas-fir. Phytophagous species accounted for 67% of the arthropods captured, and most of these were sapsucking Homoptera. The Cooley spruce gall aphid, Adelges cooleyi (Gillette) accounted for about 81% of the phytophagous numbers. Population numbers of adelgids may reach outbreak proportions in particular Douglas-fir stands, but this did not happen in Plot 1 during 1973, even though the estimated abundance was at least 2.5 million/ha during late July (Figure 3). This number is underestimated because of the insect's tenacity in the early instars and its small size in relation to the capture equipment method.

Table 2. General trophic composition of arthropods collected on Douglas-fir.^a

Category	No.	% of total	Major taxa and % of trophic category
Phytophages	2,537	67.3	<u>Adelges cooleyi</u> (Adelgidae) 81% <u>Scythropus ferrugineus</u> (Curculionidae) 4.6% Diprionidae 1.9%
Carnivores	842	22.4	
Predators	827	22.0	Araneida (spiders) 61.6% Syrphid larvae 10.4% <u>Podabrus</u> sp. (Cantharidae) 9.3% Neuroptera 7.6%
Parasites	15	0.4	
Scavengers	223	5.9	Psocoptera 78.0%
Ants	115	3.1	
Unknown and nonfeeding	50	1.3	
Total	3,767	100.0	

^aTrophic categories explained in Introduction.

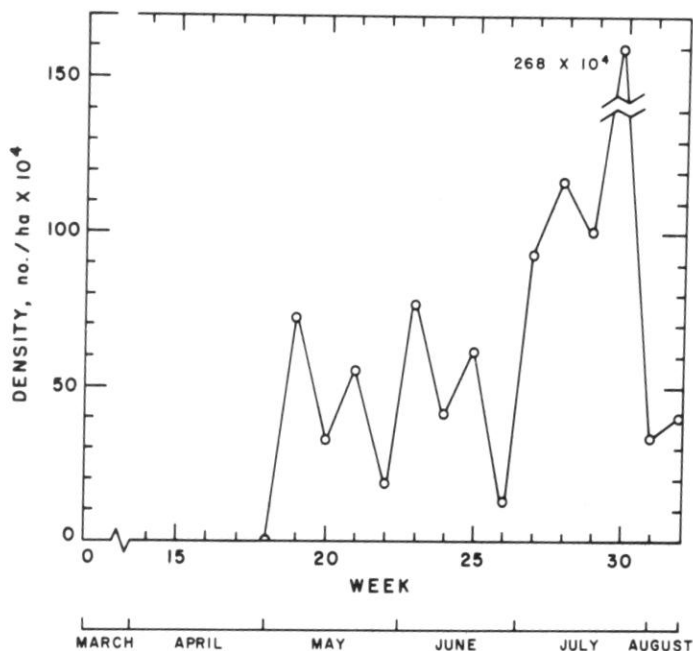


Figure 3. Weekly density estimates of the Cooley spruce gall aphid, Adelges cooleyi (Gillette), on 20-year-old Douglas-fir from pole-pruning samples, March through August 1973, H. J. Andrews Experimental Forest, Oregon.

The phytophagous weevil, Scythropus ferrugineus Casey, was most abundant in early spring when sampling began and the population was already on the decline (Figure 4). These weevils comprised about 5% of the phytophagous forms present on the trees and attained a relative abundance of at least 166,000/ha in late March.

Sawfly larvae of the Diprionidae accounted for about 2% of the phytophagous insects on Douglas-fir. Infestation was minimal, and no diprionid adults were collected by this method or by rotary net sampling. Most larvae were collected in late May and late June, and numbers declined thereafter. The maximum abundance was estimated to be about 70,000/ha in late May (Figure 4).

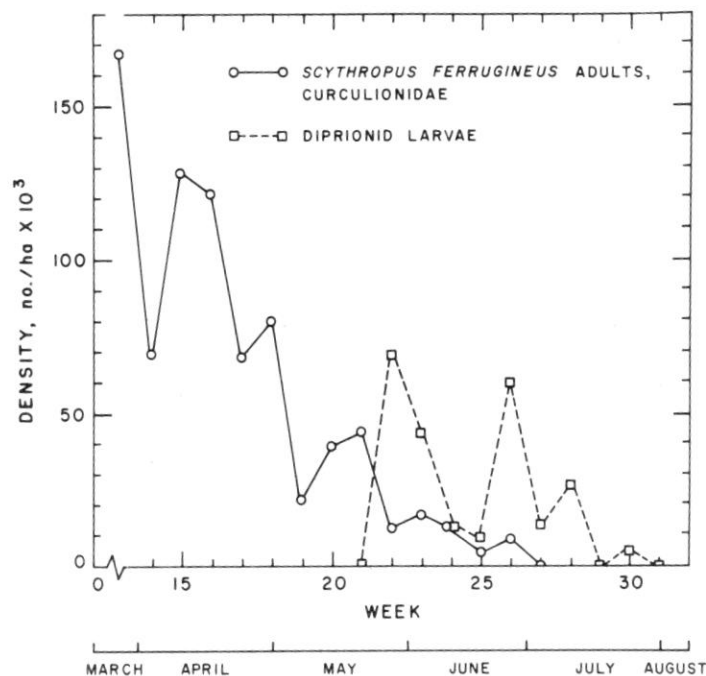


Figure 4. Weekly density estimates of the weevil, Scythropus ferrugineus Casey and diprionid larvae on 20-year-old Douglas-fir from pole-pruning samples, March through August 1973, H. J. Andrews Experimental Forest, Oregon.

Predaceous species made up about 22% of the arthropods collected (Table 2). The 3:1 ratio of phytophages to predators on the trees is similar to the ratio of (2.8-3.2):1 observed by Martin (1966) on red pine. Hunting spiders of the Salticidae and Thomisidae were the most abundant spiders. Predaceous cantharid beetles, syrphid fly larvae, and Neuroptera also were prominent. Adelgids were probably the most abundant and accessible prey, but to what extent predators utilized these prey is unknown. Cumming (1959) stated

that, in spite of high mortality in adelgids, predatory effects seemed to be minimal in the population she studied. In our study, spiders reached combined densities of over 250,000/ha in mid-May (Figure 5), and syrphid larvae and cantharid adults (*Podabrus* sp.) reached densities of 92,000/ha and 136,000/ha in early June (Figure 6).

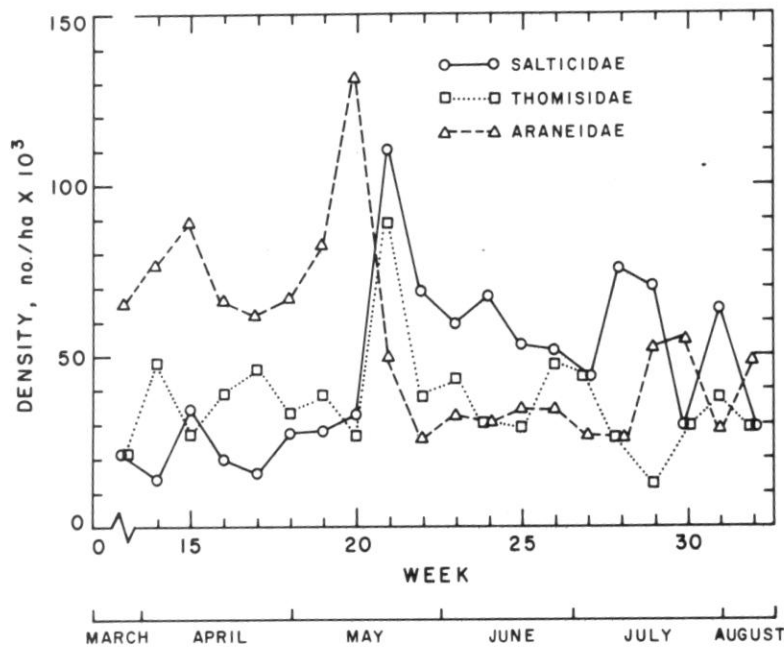


Figure 5. Weekly density estimates of spiders of the Araneidae, Salticidae, and Thomisidae on 20-year-old Douglas-fir from pole-pruning samples, March through August 1973, H. J. Andrews Experimental Forest, Oregon

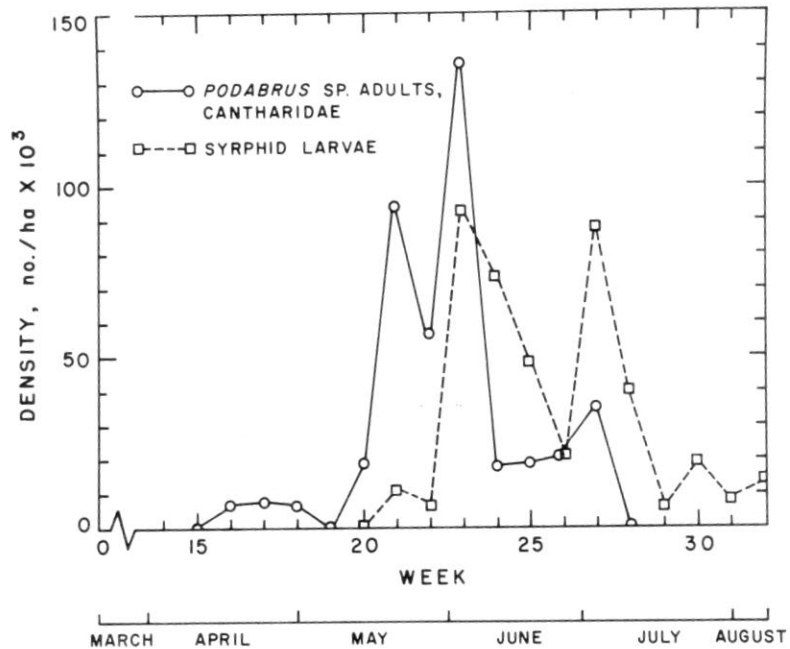


Figure 6. Weekly density estimates of cantharid beetles, *Podabrus* sp., and syrphid fly larvae on 20-year-old Douglas-fir from pole-pruning samples, March through August 1973, H. J. Andrews Experimental Forest, Oregon.

Scavengers were represented mainly by Psocoptera (Table 2). Depending on exact species present, however, some or all of these might actually be phytophagous. For lack of species identifications, whether ants collected were scavengers, phytophages, or predators was impossible to ascertain.

Capture Efficiency. Whether 15 branch samples per week from only five trees adequately sampled the arthropod fauna present on Douglas-fir branches is debatable. Varty (personal communication) mentioned, in connection with sampling techniques for the balsam twig aphid, that single branch samples from 40 trees per collection date would yield less than 20% standard error. Mason (1970) calculated that to sample larval populations of the tussock moth, three branch samples each from 11 or 12 trees would give a density estimate with a standard error within 20% for medium to heavy populations of larvae. Interpolation from his Table IV (p. 843) shows that the sampling technique followed in our study might be within 30% standard error if we were sampling for tussock moth larvae. The error factor for other taxa is not known.

GROUND-DWELLING ARTHROPODS

Average densities of ground-dwelling arthropods in forest soils have been enumerated elsewhere (Gill 1969, Huhta *et al.* 1967, Wallwork 1976). However,

little investigation has been conducted on the wandering macroarthropod fauna of forest floors as collected by pitfalls (Huhta 1971, Uetz 1975).

Methods

The pitfall traps were No. 10 steel cans, 15 cm in diameter and 18 cm deep. Each can was buried below the litter, flush with the soil surface. Square pieces of 1/3-inch hardware cloth, placed about 5-8 cm below the rim of the can, prevented both the capture of small mammals and reptiles and their consumption of the trapped arthropods. An aspirator was used to collect the smaller arthropods. Rainwater and debris, which had accumulated between sampling periods, was removed with a sponge before and after each collecting period.

Pitfall locations sampled the previous year were re-used in Plots 2, 3, and 5. Each plot contained 15 pitfalls, spaced 10 m apart and radiating from a central trap along the four compass coordinates. Plots 1 and 4 each contained 25 traps arranged in a 5 x 5 grid, each trap 10 m apart. The increased number of traps in these plots was an attempt to more intensively trap the ground-dwelling arthropods in these specific stands.

All pitfall traps were left open for 48 consecutive hours each week, resulting in 30 trap nights per week (1 trap night = 24 hours) for Plots 2, 3, and 5, and 50 trap nights per week for Plots 1 and 4. As thorough collection of Collembola and Acarina was not possible with this trapping method, no attempt was made to assess these populations. However, preliminary counts indicated that these two groups would have been the most numerous arthropods in most of the sampling periods and plots.

To compare the numbers of arthropods collected in each of the stands and to compensate for the greater number of traps in plots 1 and 4, the numbers collected in these two plots had to be reduced by multiplying by a factor of 0.6. For purposes of this analysis, we assumed that use of either the grid pattern or radiating pattern in the plots did not significantly affect number or pattern of arthropod captures.

Results and Discussion

Species Composition. Appendix 2 lists the number of specimens of each species or taxon collected in the pitfall traps. Table 3 summarizes the numbers of arthropods collected by taxonomic order. The greatest numbers of arthropods were collected from the clearcut Plots 4 and 5, where large populations of wandering spiders and ants were encountered. Young-growth Plot 1 retained much of the clearcut fauna, but had additional species inhabiting the young understory. Despite the more intensive sampling in Plots 1 and 4, data suggest that clearcut Plots 4 and 5 and young-growth Plot 1 had a greater species diversity than mid- and old-growth Plots 2 and 3 (Appendix 2).

Table 3. Arthropods collected in pitfall traps.^a

Order	Plot ^b									
	1		2		3		4		5	
	Young growth No. No. ^c total		Midgrowth No. No. ^c total		Old growth No. No. ^c total		Clearcut No. No. ^c total		Clearcut No. No. ^c total	
Noninsecta										
Isopoda	52	31	0	-	2	0.4	0	-	0	-
Chilopoda (Class)	25	15	5	1.0	12	2.2	25	0.9	11	0.7
Diplopoda (Class)	77	46	59	11.6	51	9.4	20	0.7	66	3.9
Chelonethida	17	10	3	0.6	18	3.3	2	0.0	5	0.3
Phalangida	14	8	2	0.4	13	2.4	11	0.4	3	0.2
Araneida	103	62	51	10.1	36	6.7	645	22.2	353	20.9
Acarina	p	p	p		p		p		p	
Insecta										
Collembola	p	p	p		p		p		p	
Heteroptera	47	28	3	0.6	0	-	87	3.0	84	5.0
Homoptera	16	10	2	0.4	13	2.4	61	2.1	156	9.2
Coleoptera	614	368	299	58.7	220	40.7	339	11.7	357	21.2
Diptera	47	28	22	4.3	37	6.9	46	1.6	14	0.8
Hymenoptera	625	375	56	11.0	115	21.3	1,647	56.7	611	36.2
Others	19	11	7	1.4	23	4.3	23	0.8	28	1.7
Total arthropods	1,656	992	509		540		2,906		1,688	

^aData for Plots 1 and 4 include columns listing the numbers reduced (times 0.6) for interplot comparisons because Plots 1 and 4 were more intensively sampled than Plots 2 and 3 (1,000 compared to 600 trap nights).

^b"p" = present in great numbers, but not counted.

^cNumbers in previous column times 0.6.

Clearcut Plots 4 and 5, with their high temperatures and burned slash, were ideally suited to wandering spiders of the Lycosidae and Gnaphosidae. Large populations of ants, mostly Camponotus spp. (carpenter ants), occurred within the slash. The carabid beetles, Harpalus sp., were relatively common ground dwellers in these clearcuttings and were not found in the forested stands.

Young-growth Plot 1 had a thick growth of secondary shrubs and trees, and concomitant with this undergrowth was a large phytophagous arthropod population. Large numbers of the phytophages, such as chrysomelid beetles, Pyrrhalta carbo LeConte, and the weevils, Dyslobus sp., were collected in this plot. Ants were common, but not as abundant as in the clearcuttings. Isopods were found almost exclusively in this plot. The carabids, Promecognathus crassus LeConte and Pterostichus herculeanus Mann., also were common in Plot 1.

Midgrowth Plot 2 had increased numbers of curculionids, mostly Steremnius carinatus Boh., and decreased numbers of Heteroptera and Homoptera, especially ants, compared to the other plots. The carabid, Pterostichus herculeanus Mann., was especially numerous in this plot.

Old-growth Plot 3 had a large unestimated population of Collembola and Acarina in its thick, damp layer of litter and mosses. The predaceous carabid, Promecognathus crassus LeConte, which feeds on slugs and snails, was abundant in this habitat. The number of wandering spiders, however, was less than in the other habitats sampled.

When the sampled populations are divided into trophic categories, habitat variation is expressed by the faunal composition. The composition of the ground-dwelling arthropods is related to the herbaceous vegetation and available litter and, at least indirectly, to the climate that produced that habitat. Table 4 describes the trophic composition of the various habitats as a percentage of the collected population. This trophic classification is preliminary and generalized, often based at the family level. Presentation of accurate Collembola and Acarina counts would have changed Table 4 substantially.

The greatest numbers of predators were collected in clearcut Plots 4 and 5. They composed only 30%-36% of the epigeal arthropods captured in these two plots, compared to 48% and 40% in Plots 2 and 3 (Table 4). The heavy litter and moss layer in Plot 3 supported large populations of scavengers and detritivores, which composed about 24% of the arthropods in this plot, compared to 4%-15% for the other four plots. As light intensity, temperature, and humidity changed in the litter layer with increased stand age (Figure 1), the population of detritivores and scavengers inhabiting that niche increased proportionately (Table 4) (Gill 1969, Pedigo 1970).

Table 4. General trophic composition of arthropods collected in pitfall traps.^a

Category	Plot									
	1		2		3		4		5	
	Young growth % of total		Midgrowth % of total		Old growth % of total		Clearcut % of total		Clearcut % of total	
	No.		No.		No.		No.		No.	
Phytophages	435	26.3	118	23.2	68	12.6	231	7.9	334	19.8
Carnivores	367	22.2	254	49.9	217	40.2	925	31.8	655	38.8
Predators	356	21.5	247	48.5	214	39.6	876	30.1	612	36.3
Parasites	11	0.7	7	1.4	3	0.6	49	1.7	43	2.5
Scavengers	211	12.7	76	14.9	128	23.7	124	4.3	126	7.5
Ants	610	36.8	51	10.0	112	20.7	1,600	55.1	558	33.1
Unknown and nonfeeding	33	2.0	10	2.0	15	2.8	26	0.9	15	0.9
Total	1,656	100.0	509	100.0	540	100.0	2,906	100.0	1,688	100.1

^aTrophic categories explained in Introduction.

Table 3 indicates some large differences in the composition by Order in clearcut Plots 4 and 5. For example, Hymenoptera (mostly ants, Appendix 2) were 56.7% and 36.2%, and Coleoptera were 11.7% and 21.2% in Plots 4 and 5. It is difficult to ascertain if these numbers reflect physical variation within stand type, such as vegetational or slope variability or both, or variation in pitfall trap spacing, i.e., grid pattern compared to radiating pattern. Acceptance of the second possibility would only justify comparisons between the grid traps on Plots 1 and 4 and, separately, among the radiating traps of Plots, 2, 3, and 5.

Martin's (1965) 4-year pitfall study of the ground-dwelling arthropods in a red pine plantation showed an increase in the number of spiders collected with increasing stand age. His mean values ranged from 31% to 54% of the fauna, excluding Acarina and Collembola, in the "establishment" to "young forest" stage. In our study, however, the greatest number of spiders comprised about 22% and 21% of the fauna in clearcut Plots 4 and 5, but forested Plots 1, 2, and 3 had only about 6%, 10%, and 7% spiders (Table 3).

Midgrowth Plot 2 showed the greatest percentage of Coleoptera, 59%, and clearcut Plots 4 and 5 showed the least, 12%-21% (Table 3). The large numbers of Carabidae and Curculionidae in most plots, and Chrysomelidae in Plot 1, resulted in larger populations of Coleoptera than the 22%-27% reported by Martin (1965) in red pine stands. Ants also were more abundant in our study sites than in plantations Martin investigated.

Capture Efficiency. The effectiveness of pitfall traps has been debated in the literature. Greenslade (1964) and Southwood (1966) criticized this method to quantify populations, and Luff (1975) discussed some factors that make pitfall traps unacceptable for population quantification. Banerjee (1970) concluded that "a direct relationship does not exist between densities and number trapped...for sample surveys designed to assess relative population densities in different habitats". When practical, mark-recapture techniques, used in conjunction with pitfall trapping, appear to be useful, but a small number of recaptures and the problems Luff (1975) describes present sources of error. Thomas and Sleeper (1977) discussed this problem and the equations used for estimating densities of tenebrionid beetle populations in a desert community.

Gist and Crossley (1973) described the use of fenced extinction plots and the quantifications possible. Mispagel (1977) extensively followed the extinction plot concept in a desert community for larger beetles with excellent results. Nevertheless, to totally eradicate all ground-dwelling arthropods in a 100-m² plot within a single year's time, was extremely difficult, if not impossible. With adequate maintenance and the use of drifts or attractants for quick capture, extinction plot trapping was considered an adequate method to quantify epigeal arthropods.

In this study, pitfalls were widely scattered in Plots 2, 3, and 5 and concentrated in a grid pattern in Plots 1 and 4. Among other factors, captures can be influenced by slope exposure (Tolbert 1975) and the low probability of sampling an adequate proportion of the area utilized by the existing populations because of the number or location of traps, or both. Assuming the same error factors in each study plot, a knowledge of the

number of individuals and the biologies of the species groups captured in each plot can be used to describe the differences in vegetation, cover, and climate of those plots as reflected by the arthropod fauna adapted to those conditions.

AERIAL ARTHROPODS

Little work has been done to describe aerial populations or their diversity through various strata of a community (Duviard and Pollet 1973). We used a rotary net to describe the aerial component of the forest arthropod fauna at a single height in different age stands of Douglas-fir. This method has been followed primarily to estimate the population structure and flight patterns of scolytid bark beetles (Daterman et al. 1965, Gara and Vite 1962).

Methods

Arthropods were sampled by a rotary net device developed by Gara and Vite (1962). It consisted of a nylon mesh net, 38 cm in diameter and 70 cm long, rotated in a horizontal plane 1.83 m aboveground. The net was rotated at 60 rpm by a 0.25 horsepower electric motor, powered by a portable gasoline generator, located at least 15 m from the net assembly.

Sampling was continuous for a single 2-hour period each week in Plots 2 and 3, and for two 2-hour sampling periods on different days each week in Plots 1 and 4. The majority of samples were taken between 10:00 A.M. and 4:00 P.M. To standardize the sampling time, we excluded early morning and late evening crepuscular flying insects. Limited manpower and equipment made impossible standardization of sampling time to a particular 2-hour period of the day.

Results and Discussion

Species Composition. Appendix 3 lists the numbers of specimens of each species or taxon collected by rotary net during the sampling periods in each stand type. Table 5 summarizes the totals by taxonomic order. The greatest numbers were collected in clearcut Plot 4 followed by young-growth Plot 1. Although the sampling effort was twice as great in these two plots as in the older stands, the greater numbers of arthropods possibly reflect the greater diversity of the herbaceous understory and the vertical limitation of habitat. In other words, stands with tall trees, as Plots 2 and 3, probably have more vertically dispersed aerial fauna than a stand with short trees or a clearcut plot. Plots 2 and 3 contain arthropods in the canopy that might not be expected to be present at the lower 1.83-m level of sampling.

Diptera were the most commonly captured insects and composed 63%-82% of the aerial fauna captured in all stand types (Table 5). Flies of the families Mycetophilidae, Chironomidae, Empididae, and Cecidomyiidae were most numerous, comprising 56% of all flies and about 41% of all arthropods found in all stands (Appendix 3).

Table 5. Arthropods collected in rotary net traps over 20 weeks in 1973.

	Plot							
	1		2		3		4	
	Young growth		Midgrowth		Old growth		Clearcut	
	No. ^a	% of total	No.	% of total	No.	% of total	No. ^a	% of total
Noninsecta								
Araneida	34	0.7	1	0.0	4	0.1	16	0.3
Acarina	37	0.8	6	0.2	2	0.1	6	0.1
Others	0	-	1	0.0	0	-	1	0.0
Insecta								
Plecoptera	3	0.1	0	-	1	0.0	8	0.1
Thysanoptera	0	-	19	0.5	1	0.0	307	5.4
Heteroptera	12	0.2	10	0.3	4	0.1	33	0.6
Homoptera	122	2.5	334	9.5	208	6.6	801	14.0
Psocoptera	11	0.2	0	-	3	0.1	1	0.0
Coleoptera	192	4.0	298	8.5	270	8.6	166	2.9
Neuroptera	3	0.1	8	0.2	2	0.1	3	0.1
Lepidoptera	15	0.3	14	0.4	13	0.4	56	1.0
Diptera	3,926	81.9	2,625	74.9	2,456	78.3	3,611	63.0
Hymenoptera	438	9.1	189	5.4	160	5.1	713	12.4
Others	2	0.0	2	0.1	11	0.4	6	0.1
Total arthropods	4,795		3,507		3,135		5,728	

^aBecause of the increased sampling effort in Plots 1 and 4, the numbers collected are halved to simulate trapping intensity equal to Plots 2 and 3.

Coleoptera comprised 3%-9% of the aerial fauna and were more abundant in the forested areas (Table 5). Members of the Scolytidae, Scraptiidae, Elateridae, and Cantharidae were most numerous. In spite of the more intensive sampling in Plots 1 and 4, indications were that Coleoptera species composition was more diverse in clearcut Plot 4 than the other three plots (Appendix 3).

Homoptera were most plentiful in clearcut Plot 4 where over 65% were Adelges cooleyi and 24% were aphids (Table 5). In contrast, aphids composed about 70%-90% of the Homoptera collected in the three forested plots. Aerial Thysanoptera were collected almost exclusively in the clearcut plot.

The spiders collected by rotary net (Table 5, Appendix 3) were usually immature forms that may have dropped from the trees and were ballooning on the wind.

Hymenoptera were somewhat more abundant in Plots 1 and 4, 9%-12% of the totals, than the 5% in Plots 2 and 3 (Table 5). The parasitic Braconidae

and Ichneumonidae comprised 60%, 41%, and 49% of the Hymenoptera in Plots 1, 2, and 3. Miscellaneous Chalcidoidea and Proctotrupeoidea were the most abundant Hymenoptera in clearcut Plot 4, comprising about 41% of the total compared to only 2%-14% in the other three plots. The apoid bees, such as the Andrenidae, Apidae, and Megachilidae, comprised 20%-23% of the Hymenoptera found in the midgrowth and clearcut plots and only 11%-14% in the young- and old-growth plots. High numbers would have been expected in the clearcut and young-growth plots if one considers only the availability of nectar and pollen sources. If some of the bees wereinquilines, however, host and nest requirements must be considered when explaining the numbers of pollinators.

Ants collected in the clearcutting by aerial net further verifies the large formicid population previously indicated by pitfall trapping.

Table 6 shows the habitat variation indicated by the trophic composition of the aerial arthropod fauna. Many of the most common flies do not feed in the adult stage, have unknown feeding habits, or belong to a family with variable feeding habits. Chironomids, mycetophilids, cecidomyiids, phorids, and sciarids are examples. Therefore, between 48%-67% of the arthropods in each plot were categorized as Unknown/Nonfeeding. Doubtless, many of these actually belong to the Scavenger/Detritivore or Phytophage categories. Because of the substantial numbers of rotary net arthropods classified as Unknown/Nonfeeding, we reserve any comments about the trophic composition of the aerial fauna. A shift of part or most of the Unknown/Nonfeeding category of Table 6 into the other categories would drastically change the numbers and the percentages relative to the different stand types, possibly contradicting any preliminary remarks that relate stand structure to faunal composition.

Table 6. General trophic composition of arthropods collected in rotary nets.^a

Category	Plot							
	1		2		3		4	
	Young growth		Midgrowth		Old growth		Clearcut	
	No.	% of total	No.	% of total	No.	% of total	No.	% of total
Phytophages	885	9.2	627	17.9	410	13.1	3,057	26.7
Carnivores	3,156	32.9	575	16.4	540	17.2	2,696	23.5
Predators	2,148	22.4	340	9.7	264	8.4	1,297	11.3
Parasites	1,008	10.5	235	6.7	276	8.8	1,399	12.2
Scavengers	75	0.8	63	1.8	87	2.8	98	0.9
Ants	3	0.0	3	0.1	2	0.1	70	0.6
Unknown and nonfeeding	5,462	57.0	2,239	63.8	2,096	66.9	5,529	48.3
Total	9,581	99.9	3,507	100.0	3,135	100.1	11,450	100.0

^aTrophic categories explained in Introduction.

Capture Efficiency. Although primarily limited to qualitative analyses, we feel that the rotary net sampling device can be effective in determining faunal composition and relative abundance of adult forms of flying insects if care is used in standardization of sample timing and equipment. Nevertheless, its effectiveness for particular species is debatable. For example, the motion of the net easily distracts adult Lepidoptera, which may avoid the swinging net. Furthermore, the mesh of the net may be too large to adequately sample the smallest arthropods. Therefore, a solid piece of material sewn in the bottom of the net is advisable. The assumption that the proportion of the fauna flying one day is the same as that flying the next day is not necessarily valid in detailed analyses of rotary net data. Insect flight activity is dependent upon prevailing climatic and microclimatic factors in addition to season and time of day. Wind velocity, cloud cover, precipitation, and barometric pressure can all affect the flight response of certain insects (Johnson 1969).

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APPENDIX 1. Arthropods of each species or taxon collected from 20-year-old Douglas-fir by pole-pruning sampling in young-growth Plot 1 in the H. J. Andrews Experimental Forest, Oregon, March 27 to August 7, 1973.

Species	Total No. a	Species	Total No. a
Arachnida ^b		Coleoptera	
Acarina		Cerambycidae	
Araneida		<u>Anoplopera crassipes</u> Lec.	1
Amaurobiidae	16	Chrysomelidae	
Anyphaenidae	4	<u>Pyrrhalta carbo</u> Lec.	3i, 60a
Araneidae	9	Cleridae	
Clubionidae	184	<u>Enoclerus sphegeus</u> F.	1
Gnaphosidae	5	<u>Hydnocera scaber</u> Lec.	1
Oxyopidae	1	Coccinellidae	
Salticidae	2	<u>Anatis rathvoni</u> Lec.	1
Thomisidae	172	<u>Cycloneda polita</u> Csy.	1
Insecta	133	<u>Hyperaspis</u> sp.	1
Coleoptera		<u>Mulsantina picta</u> Rand.	3
Alleculidae		<u>Psyllobora 20-maculata</u> Say	1
Hymenophorus sinuatus Fall	1	<u>Scymnillus aterrimus</u> Horn	1
Bruchidae		<u>Scymnus ardelio</u> Horn	2
<u>Acanthoscelides pauperculus</u> Lec.	6	<u>S. lacustris</u> Lec.	1
Buprestidae		Unknown	6i
<u>Anthaxia deleta</u> Lec.	3	Curculionidae	
Cantharidae		<u>Cylindrocryptus furnissi</u> Buch.	5
<u>Malthodes dorotheae</u> Fend.	7	<u>Dyslobus segnis</u> Lec.	5
<u>M. flexuosus</u> Fend.	1	<u>Dyslobus</u> sp.	1
<u>Malthodes</u> sp.	1	<u>Lechriops californica</u> Lec.	1
<u>Podabrus cavicollis</u> Lec.	66	<u>Pissodes fasciatus</u> Lec.	7
<u>P. conspuratus</u> Fall	1	<u>Rynchaenus parvicollis</u> Lec.	2
<u>P. piniphilus</u> Dej.	9	<u>Scythropus ferrugineus</u> Csy.	118
<u>Podabrus</u> sp.	1	Dascillidae	
<u>Silis insperata</u> Green	6	<u>Macropogon piceus</u> Lec.	5
Cephaloidea		Derodontidae	
<u>Cephaloon tenuicornis</u> Lec.	2	<u>Laricobius laticollis</u> Fall	1

APPENDIX 1. (Continued)

Species	Total No. ^a	Species	Total No. ^a
Coleoptera		Coleoptera	
Elateridae		Staphylinidae	
<u>Ampedus oregonus</u> Schaef.	1	<u>P. testaceum</u> Mann.	1
<u>Ctenicera columbiana</u> Brown	1	<u>Xylodromus concinnus</u> Marsh	1
<u>C. umbripennis</u> LeC.	1	Unknown family	1i
<u>Megapenthes caprellus</u> LeC.	14	Collemola ^b	
Unknown	5	Sminthuridae	7
Helodidae		Diptera	
<u>Elodes</u> sp.	1	Anthomyiidae	2
Hydrophilidae		Cecidomyiidae	7
<u>Cercyon</u> sp.	1	Ceratopogonidae	3
Melyridae		Chironomidae	62
<u>Anthocomus mixtus</u> Horn	12	Chloropidae	2
<u>Eurelymus atra</u> Csy.	1	Culicidae	2
Oedemeridae		Empididae	24
<u>Oxaxis bicolor</u> LeC.	1	Lauxaniidae	
<u>Xanthochroa testacea</u> Horn	1	<u>Minettia flaveola</u> (Coq.)	1
Ostomidae		Lonchaeidae	1
<u>Eronyxa pallidus</u> Mots.	3	Lonchopteridae	1
Scolytidae		Muscidae	5
<u>Hylastes nigrinus</u> Mann.	1	Mycetophilidae	4
<u>Pityophthorus</u> sp.	1	Phoridae	2
<u>Pseudohylesinus nebulosus</u> LeC.	4	Sciaridae	9
<u>Scolytus unispinosus</u> LeC.	2	Simuliidae	5
Scraptiidae		Sphaeroceridae	1
<u>Anaspis</u> sp.	1	Syrphidae	86i, 1a
Staphylinidae		Tephritidae	
<u>Amphiroum maculatum</u> Horn	3	<u>Pericantha</u> sp.	1
<u>Medon shastanicum</u> Csy.	1	Unknown family	13i
<u>Pelecomalium opaculum</u> Csy.	2	Ephemeroptera	
		Ephemereilidae	1

Species	Total No. ^a	Species	Total No. ^a
Ephemeroptera		Homoptera	
Unknown family	1	Cicadellidae	
Hemiptera		<u>Ballana</u> sp.	2
Lygaeidae		<u>Erythroneura</u> sp.	1
<u>Kleidocerys</u> sp.	1	<u>Scaphytopius</u> sp.	1
Miridae		Unknown	2i, 5a
<u>Deraeocoris</u> sp.	2	Psyllidae	6
<u>Dicyphus</u> sp.	1i	Hymenoptera	
<u>Phytocoris</u> sp.	1	Braconidae	3
<u>Pilophorus</u> sp.	2	Cynipidae	1
<u>Plagiognathus</u>	11i, 17a	Diprionidae	47i
<u>Psallus</u> sp.	15i, 14a	Eucharitidae	1
Unknown	4	Figitidae	1
Pentatomidae		Formicidae	115
<u>Euschistus</u> sp.	1	Ichneumonidae	2
<u>Peribalus</u> sp.	1	Misc. Chalcidoidea and Proctotrupoidea	2
<u>Podisus</u> sp.	1	Lepidoptera	
Reduviidae		Geometridae	3i
<u>Zelus</u> sp.	1i	Noctuidae	3i
Tingidae		Nymphalidae	
<u>Corythuca</u> sp.	1	<u>Limnitis archippus</u> (Cramer)	1i
Unknown family	1i	Misc. microlepidoptera	6
Homoptera		Unknown	6i
Achilidae		Neuroptera	
Adelgidae ^b	7	Chrysopidae	25i, 2a
<u>Adelges cooley</u> i (Gillette)	2056	Coniopterygidae	1
Aphididae ^b	8	Hemerobiidae	22i, 11a
Cercopidae		Raphidiidae	
<u>Aphrophora</u> sp.	2	<u>Agulla</u> sp.	2
<u>Philaenus</u> sp.	1		

APPENDIX 1. (Continued)

Species	Total No. ^a	Species	Total No. ^a
Orthoptera		Plecoptera	
Acrididae	1	Chloroperlidae	
Gryllidae	1i	Alloperla sp.	1
Tettigoniidae		Leuctridae	
Insara sp.	1	Leuctra sp.	1
Plecoptera		Psocoptera ^b	174
Capniidae		Thysanoptera ^b	1
Capnia sp.	1	Unknown Order	1i

^a "i" indicates immature form captured; all other numbers refer to adult captures (sometimes "a" is used to avoid confusion).

^b All numbers were recorded as adults; no attempt was made to differentiate between adult and immature life stages.

APPENDIX 2. Total numbers of arthropods of each species or taxon collected from 5 different Douglas-fir stands by pitfall trapping in the H. J. Andrews Experimental Forest, Oregon, March 27 to August 7, 1973.^a

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Isopoda ^c	52		2		
Diplopoda ^c					
Chordeumida	13	9	9	6	5
Polydesmida					
Chonaphe sp.	17		2		19
Harpaphe haydeniana	32	49	36	5	4
Unknown	8	1	3	3	38
Polyxeniida					
Polyxenus sp.				2	
Spirobolida	1				
Unknown	6		1	4	
Chilopoda ^c					
Geophilomorpha	2	1	1	1	2
Lithobiomorpha	12	2	7	10	5
Scolopendromorpha	7	2	4	13	4
Unknown	4			1	
Arachnida ^c					
Acarina	p	p	p	p	p
Araneida					
Agelenidae	4		1	2	
Amaurobiidae	8	2	2	1	1
Antrodiaetidae	7	4	4	6	6
Anyphaenidae	1	1	1		
Araneidae	28	9	8	4	2
Clubionidae					
Ctenizidae	2	2	1	1	

APPENDIX 2. (Continued)

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Arachnida ^c					
Araneida					
Dipluridae		1	1		1
Gnaphosidae	8	4		48	28
Hypochilidae				1	
Lycosidae					
Lycosa sp.	21	15	3	499	283
Unknown	1	3	4		
Oxyopidae				1	1
Salticidae				5	1
Thomisidae	2	2	3	5	1
Unknown	21	8	8	72	30
Chelonethida	17	3	18	2	5
Phalangida	14	2	13	11	3
Insecta					
Coleoptera					
Alleculidae				1	
Mycetochara malkini Hatch					
Byrrhidae				6	
Byrrhus stolidus Csy.				1	
Byrrhus sp.					
Lioon simplicipes Mann.			1		
Listemus formosus Csy.	7	1		9	4
Listemus sp.	3			1	
Morychus oblongus LeC.					5
Unknown				2	
Cantharidae					
Malthodes dorotheae Fend.	1				
M. oregonus Fend.					1
Malthodes sp.			1		

APPENDIX 2. (Continued)

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Coleoptera					
Cantharidae					
<i>Podabrus piniphilus</i> Dej.	1				
<i>Silis insperata</i> Green	1				
Unknown					
Carabidae				1i,4a	1i,2a
<i>Amara littoralis</i> Mann.					4
<i>A. sinuosa</i>				3	2
<i>Amara</i> sp.				1	2
<i>Apristus constrictus</i> Csy.				7	
<i>Bembidion osculans</i> Csy.	1				5
<i>Cychrus tuberculatus</i> Harr.	1	2	2	2	3
<i>Harpalus</i> sp.					95
<i>Microlestes</i> sp.				17	14
<i>Notiophilus sylvaticus</i> Esch.	2i		1i	1	1i
<i>Promecognathus crassus</i> LeC.	80	15	79	24	5
<i>Pterostichus amethystinus</i>	2		2		
<i>P. castaneus</i> Dej.	2		3	2	2
<i>P. herculeanus</i> Mann.	70	107	32	25	9
<i>P. inopinus</i> Csy.	5		1	2	3
<i>P. lama</i> Men.	2	6	4	7	4
<i>Scaphinotus angulatus</i> Harr.	1		5	1	
<i>S. marginatus</i> Fisch.	2		1	1	1
<i>S. rugiceps</i> Horn		5			
<i>Zacotus matthewsii</i> LeC.		3	1		
Unknown					1
Cephaloidea					
<i>Cephaloon tenuicornis</i> LeC.					1
Cerambycidae					
<i>Anoplodera crassipes</i> LeC.		1		1	
<i>Dicentrus bluthneri</i> LeC.		1			

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Coleoptera					
Chrysomelidae					
<u>Altica tombacina</u> Mann.	1				2
<u>Bromius obscurus</u> L.	1			8	
<u>Monoxia angularis</u> LeC.				6	2
<u>Pachybrachis melanostictus</u>				2	1
<u>Pyrrhalta carbo</u> LeC.	209i, 2a				
<u>Scelolyperus varipes</u> LeC.	1				1
<u>Syneta hamata</u> Horn			1		
<u>S. simplex</u> LeC.	21	1		7	5
<u>Timarcha intricata</u> Hald.				11	
Unknown					
Cicindelidae					
<u>Omus dejeani</u> Reiche	19	18	4	28	3
Clambidae					
<u>Empelus brunnipennis</u> Mann.			1	1	
Coccinellidae					
<u>Coccinella trifasciata</u> L.				1	1
Cryptophagidae					
<u>Atomaria vespertina</u> Makl.	1				
Curculionidae					
<u>Cnemogonus lecontei</u> Dietz				8	2
<u>Dyslobus granicollis</u> LeC.	41			1	5
<u>D. segnis</u> LeC.	10	32	3	5	1
<u>Dyslobus</u> sp.	2		1		
<u>Geodercodes latipennis</u> Csy.	14		1	1	3
<u>Lobosoma horridum</u> Mann.			1		
<u>Nemocestes incomptus</u> Horn	4				
<u>N. puncticollis</u> Csy.	1				17
<u>Nemocestes</u> sp.	3				
<u>Plinthodes taeniatus</u> LeC.	6	1			7

APPENDIX 2. (Continued)

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Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Coleoptera					
Curculionidae					
<u>Rynchaenus rufipes</u> LeC.				1	1
<u>Scythropus ferrugineus</u> Csy.	1			2	5
<u>Sitona californicus</u> Fahr.			35	1	6
<u>Steremnius carinatus</u> Boh.	16	60		1	
Unknown					
Dascillidae					
<u>Macropogon</u> sp. ?				2	
Elateridae					
<u>Athous varius</u> Lane				1	
<u>Ctenicera suckleyi</u> LeC.		1			
<u>Hemicrepidius morio</u> LeC.	2	1			
<u>Megapenthes caprellus</u> LeC.				2	
<u>Negastrius</u> sp.	1				3
Unknown	1	1i, la			1i, la
Endomychidae					
<u>Stethorhanis borealis</u> Blais.	1			1	
<u>Xenomycetes laversi</u> Hatch					1
Helodidae					
<u>Cyphon concinnus</u> LeC.	1				
<u>Elodes</u> sp.	1				
Lampyridae					
<u>Ellychnia hatchi</u> Fend.					
Unknown	4i	16i	10i	29i	10i
Leiodidae					
<u>Agathidium jasperinum</u> Fall	2			1	
<u>Agathidium</u> sp.					
<u>Catopocerus capizzii</u> Hatch			6		1
<u>Catops basilaris</u> Say					4
<u>Colon</u> sp.					

APPENDIX 2. (Continued)

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Coleoptera					
Leioididae					
<u>Hydnobius</u> sp.				2	1
<u>Leiodes curvata</u> Mann.				2	
<u>Leiodes</u> sp.	2			4	4
<u>Neocyrtusa sternita</u> Hatch				1	
<u>Platycholeus opacellus</u> Fall	5				1i, 1a
Unknown					
Lucanidae					
<u>Platyceroides laticollis</u> Csy.	6			3	1
Meloidae					
<u>Lytta stygica</u> LeC.					1
Melyridae					
<u>Dasytes cruralis</u> LeC.					2
Mycetophagidae					
<u>Mycetophagus pluriguttatus</u> LeC.	1				
Oedemeridae					
<u>Ditylus gracilus</u> LeC.	10	1	5	15	2
<u>D. quadricollis</u> LeC.					
Ostomidae					
<u>Eronyxa pallidus</u> Mots.				1	
Phenogodidae					
<u>Zarhipis integripennis</u> LeC.		1i			
Unknown		5i			
Pselaphidae					
<u>Abdiunguis fenderi</u> Park and Wag.			1		
<u>Actium microphthalum</u> Park and Wag.			1		
<u>Batrisesodes albionicus</u> (Aube)	2	1		2	13
<u>Batrisesodes</u> sp.	2		1		
<u>Cupila clavicornis</u> (Makl.)		1	1		
<u>Lucifotychus impellus</u> Park and Wag.			1		1

APPENDIX 2. (Continued)

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Coleoptera					
Pselaphidae					
<u>Lucifotychus</u> sp.	3		1	2	4
<u>Megarafonus</u> sp.		1		1	1
<u>Oropodes</u> sp.			1		
<u>Pselaptrichus proprius</u> Sch. and Marsh.	3				
<u>P. vanus</u> Schuster and Marsh			2	1	1
<u>Pselaptrichus</u> sp.	1		1	8	2
Unknown					
Ptilidae					
<u>Acratrichis</u> sp.	1				
Rhysodidae					
<u>Clinidium calcaratum</u> LeC.				1	1
Scarabaeidae					
<u>Aphodius opacus</u> LeC.	2			3	1
<u>Aphodius</u> sp.				4	2
<u>Bolboceras obesus</u> LeC.					
<u>Dichelonyx backii</u> Rby.		1			
<u>D. valida</u> LeC.			1		2
<u>Serica</u> sp.					
Scolytidae					
<u>Hylastes nigrinus</u> Mann.	1			1	
Scaptiidae					
<u>Anaspis</u> sp.				1	
Scydmaenidae					
<u>Eutheia scitula</u> Makl.	1	1	1		1
<u>Lophoderus similis</u> Marsh.	2			4	2
<u>Lophoderus</u> sp.	1				
Unknown				1	
Silphidae					
<u>Nicrophorus vesilloides</u> Hbst.			2		

APPENDIX 2. (Continued)

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Coleoptera					
Sphaeritidae					
<u>Sphaerites politus</u> Mann.				1	
Staphylinidae					
<u>Anthobium subcostatum</u> Makl.			1		
<u>Astenus longiusculus</u> Mann.				1	
<u>Atrecus quadripennis</u> Csy.	2				1
<u>Ischnopoda</u> sp.	1				
<u>Lithocharis obsoleta</u> Nordm.	3			2	21
<u>Megarthus pictus</u> Mots.	1			1	
<u>Mycetoporus</u> sp.	3				
<u>Ocypus rutilicauda</u> Horn				5	
<u>Orus pugetanus</u> Csy.				2	1
<u>Orus punctatus</u> Csy.					1
<u>Orus</u> sp.					1
<u>Philonthus picicornis</u> Horn					1
<u>Proteinus basalis</u> Makl.			1		
<u>Stictolinus franciscanus</u> Csy.				3	1
<u>Sunius fenderi</u> Hatch					2
<u>Xestolinus frontalis</u> Hatch					2
Unknown	3		1	1i,2a	5i,16a
Tenebrionidae					
<u>Coelocnemis californicus</u> Mann.		3		3	2
<u>Helops edwardsii</u> Horn				2	
<u>Iphthimus serratus</u> Mann.				2	1
<u>Phellopsis porcata</u> LeC.		3	1	7	2
<u>Usechus nucleatus</u> Csy.	1				
Trixagidae					
<u>Aulonothroscus validus</u> LeC.	4			1	
<u>Pactopus hornii</u> LeC.				2	
<u>Trixagus sericeus</u> LeC.				1	

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Coleoptera					
Trixagidae					
Unknown				1	
Unknown family	3i,2a		1	3i,3a	7i,1a
Collembola^c					
Entomobryidae	p	p	p	p	p
Isotomidae	p	p	p	p	p
Poduridae	p	p	p	p	p
Sminthuridae	p	p	p	p	p
Diptera					
Bibionidae					
Bibio sp.	1	1			
Bombyliidae				1	
Cecidomyiidae	6	7	1	6	3
Ceratopogonidae				1	
Chironomidae	9	1i,1a	2	10	2
Empididae	2			10	1
Lonchaeidae				1	
Milichiidae				5	
Muscidae	1				
Mycetophilidae	2		1		
Phoridae	6		9		1
Sciaridae	9	4	4	7	2
Sphaeroceridae					
Leptocera sp.	3		1		
Sphaerocera sp.	2				
Unknown			12		
Syrphidae		1i			1
Tachinidae		1		1	

APPENDIX 2. (Continued)

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Diptera					
Tipulidae					
Unknown	2 4i	2i 2i,2a	1 5i,1a	3i,1a	1i,2a
Hemiptera					
Aradidae	3			9	3
Berytidae					
Acanthophysa sp.					
Unknown				2	7 12
Cydnidae					
Amnestus sp.					
Unknown					2 1
Lygaeidae					
Geocoris sp.				2	7i,3a
Scolopostethus sp.	14 17	2		11 1i,24a	4 8
Unknown					
Miridae					
Phytocoris sp.				2i 5i	1i
Unknown					
Nabidae					
Pagasa sp.					1
Unknown					3i
Pentatomidae					
Podisus sp.		1			
Unknown				1i	1i
Reduviidae					
Zelus sp.					1
Tingidae					
Acalypta sp.				17	5
Corythuca sp.				1	
Unknown family	9i			10i,2a	23i,2a

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Homoptera					
Achilidae			1		1i
Adelgidae ^c					
Adelges cooleyi (Gillette)			2		2
Aphididae ^c	10		9	10	22
Cicadellidae					
Aceratagallia sp.				1	2
Cuerna hasbrouki Nielson				2i, 15a	10i, 11a
Scaphytropius sp.				1	
Unknown	3i, 1a	1i, 1a	1i	13i, 4a	90i, 10a
Cicadidae					
Okanagana sp.	1			3	3
Psyllidae	1i			12i	2
Unknown family					2i, 1a
Hymenoptera					
Apidae					
Apis mellifera L.	1				1
Braconidae	1			5	2
Diapriidae				9	12
Eucharitidae	1				
Formicidae	610	51	112	1,600	558
Halictidae					2
Ichneumonidae	3	1	2	2	3
Misc. Chalcidoidea and Proctotrupoidea	6	4	1	29	25
Mymaridae					1
Pompilidae				2	3
Scelionidae	1				
Sphecidae					1
Tenthredinidae					2i, 1a
Unknown family	2i				

APPENDIX 2. (Continued)

Species	Plot ^b				
	1	2	3	4	5
	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Isoptera					
Hodotermitidae					
Zootermopsis sp.	1			2	
Lepidoptera					
Geometridae	2i				1i
Noctuidae				2i	5i
Microlepidoptera				1i	1i
Unknown		1i, 1a	1i	2i	12i
Mallophaga ^c					
Trichodectidae		1			
Orthoptera					
Acrididae				3i, 1a	3i
Gryllacrididae	2i, 4a	1i	1	2	1
Plecoptera					
Nemouridae	1				
Nemoura sp.					
Psocoptera ^c	1		1		
Siphonaptera					
Ceratophyllidae				1	
Pulicidae				1	
Thysanoptera ^c		2	2	6	5

APPENDIX 2. (Continued)

Species	Plot ^b				
	1	2	3	4	5
Thysanura ^c Machilidae	Young growth	Midgrowth	Old growth	Clearcut	Clearcut
Unknown Order	1	1	17	1	
	7i		1i	1i	

^a Numbers for Plots 1 and 4 represent 1000 trap nights of sampling each (25 traps per plot open for 48 consecutive hours each week for 20 weeks) and numbers for Plots 2, 3, and 5 represent 600 trap nights (15 traps per plot open for 48 consecutive hours each week for 20 weeks).

^b "i" indicates immature life stage collected. All other numbers refer to the numbers of adults collected (the most common life stage found). "p" indicates present in great numbers, but not counted because collection methods were inconsistent.

^c Groups that were all recorded as adults; no attempt was made to differentiate between immature and adult life stages.

between immature and

APPENDIX 3. Total numbers of arthropods of each species or taxon collected by rotary nets (1.83 m aboveground) from 4 different Douglas-fir stands in the H. J. Andrews Experimental Forest, Oregon from March 27 to August 7, 1973.^a

Species	Plot ^b			
	1	2	3	4
	Young growth	Midgrowth	Old growth	Clearcut
Arachnida ^c				
Acarina	73	6	2	12
Araneida				
Araneidae	3	1	1	3
Gnaphosidae			1	
Lycosidae	1			1
Micryphantidae				1
Salticidae	1			3
Thomisidae	1		1	1
Unknown family	61		1	22
Chelonethida		1		1
Insecta				
Coleoptera				
Alleculidae			1	
<u>Mycetochara procera</u> Csy.				
Anobiidae				
<u>Ernobius gentilis</u> Fall.		1		
<u>Ptilinus basalis</u> LeC.	1			
Bruchidae				
<u>Acanthoscelides pauperculus</u> LeC.				2
Buprestidae				
<u>Agrilus politus</u> Say				1
<u>Anthaxia expansa</u> LeC.				2
<u>Chrysobothris grandis</u> Chamb.				1
<u>Melanophila drummondi</u> Kby.				1
Cantharidae				
<u>Malthodes dorotheae</u> Fend.	2			
<u>M. flexuosus</u> Fend.		7		
<u>M. oregonus</u> Fend.			1	1
<u>Malthodes</u> sp.				1

Species	Plot ^b			
	1	2	3	4
Young growth				
Midgrowth				
Old growth				
Clearcut				
Coleoptera				
Cantharidae				
<i>Podabrus cavicollis</i> LeC.				2
<i>P. conspurcatus</i> Fall	41		1	
<i>P. hackerae</i> Fend.	1			
<i>P. macer</i> LeC.	1			
<i>P. piniphilus</i> Dej.	1			
<i>P. pruinosis</i> LeC.	16	1	2	3
<i>Podabrus</i> sp.				1
<i>Silis insperata</i> Green		1		
<i>Troglomethes oregonensis</i> Witt.	25	1	2	9
Unknown	1	1	2	
Carabidae				
<i>Bembidion iridescent</i> LeC.	1	4		1
<i>B. osculans</i> Csy.	4			3
<i>Bradycellus nigrinus</i> Dej.				1
<i>B. politus</i> Fall				1
<i>Lebia viridis</i> Say				1
Cephaloidae				
<i>Cephaloon tenuicornis</i> LeC.	12			
Cerambycidae				
<i>Anoplopera amabilis</i> LeC.	1			
<i>A. aspera</i> LeC.	1			
<i>A. canadensis</i> Ol.				1
<i>A. crassipes</i> LeC.		2	7	5
<i>A. dehiscent</i> LeC.	10			3
<i>A. dolorosa</i> LeC.			1	11
<i>A. laetifica</i> LeC.	5			2
<i>Dicentrus bluthneri</i> LeC.		2		
<i>Evodina vancouveri</i> Csy.		2		1
<i>Leptura oblitterata</i> Hald.				2

APPENDIX 3. (Continued)

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Coleoptera				
Cerambycidae				
<u>Opsimus quadrilineatus</u> Mann.		1		
<u>Pidonia quadrata</u> Hop.				2
<u>P. scripta</u> Lec.			1	
Chrysomelidae				
<u>Altica tombacina</u> Mann.	1			13
<u>Bromius obscurus</u> L.	1			1
<u>Orsodacne atra</u> Ahr.				1
<u>Pachybrachis circumcinctus</u> Cr.				2
<u>Pyrhalta carbo</u> Lec.	10i, 6a			
<u>Scelolyperus varipes</u> Lec.	1	3	1	9
Unknown	1i			
Cicindelidae				
<u>Cicindela oregona</u> Lec.				1
Clambidae				
<u>Empelus brunnipennis</u> Mann.		1		
Coccinellidae				
<u>Hippodamia convergens</u> Guer.				12
<u>Mulsantina picta</u> Rand.	1			
<u>Scymnillus aterrimus</u> Horn			1	
<u>Scymnus caurinus</u> Horn				5
<u>S. lacustris</u> Lec.				1
<u>S. maculatus</u> Hatch				1
<u>Scymnus</u> sp.				3
<u>Stethorus punctillum</u> Ws.				2
Colydiidae				
<u>Lasconotus schwarzi</u> Kraus		3	1	
Unknown			1i	
Cryptophagidae				
<u>Anchicera gonodera</u> Csy.	2			

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Coleoptera				
Cucujidae				
<u>Pediacus depressus</u> Hbst.		31	43	2
Curculionidae				
<u>Cylindrocopturus furnissi</u> Buch.		1		1
<u>Deporaus glastinus</u> LeC.	1	1		
<u>Gymnaetron pascuorum</u> Gyll.	1			1
<u>Lechriops californica</u> LeC.				1
<u>Miccotrogus picirostris</u> F.				1
<u>Pissodes fasciatus</u> LeC.				
<u>Rhyncolus brunneus</u> Mann.	1			
<u>R. cylindricollis</u> Woll.		2		
<u>Rynchaenus parvicollis</u> LeC.	4			1
<u>R. rufipes</u> LeC.	1			
<u>Scythropus ferrugineus</u> Csy.	2			
<u>Sitona californicus</u> Fahr.	1			
Dascillidae				
<u>Araeopidius monachus</u> LeC.	1			
<u>Macropogon piceus</u> LeC.				39
Derodontidae				
<u>Peltastica tuberculata</u> Mann.	1		2	
Dermestidae				
<u>Anthrenus lepidus</u> LeC.				1
<u>Megatoma perversa</u> Fall	1		1	4
<u>Orphilus niger</u> Rossi				10
<u>Trogoderma</u> sp.				1
Elateridae				
<u>Ampedus apicatus</u> Say				2
<u>A. cordifer</u> LeC.				1
<u>A. varipilis</u> Van D.				6
<u>Athous rufiventris</u> Esch.	1			

APPENDIX 3. (Continued)

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Coleoptera				
Elateridae				
<u>Athous varius</u> Lane	2			
<u>Cardiophorus</u> sp.	2			1
<u>Ctenicera columbiana</u> Brown		1		
<u>C. nebraskensis</u> Bland	1			
<u>C. suckleyi</u> LeC.	2	2		
<u>C. umbripennis</u> LeC.	52	3	15	24
<u>Megapenthes caprellus</u> LeC.				1
<u>Negastrius</u> sp.	4	10	7	2
Unknown				
Erotylidae				
<u>Triplax californicus</u> LeC.		1		
Eucnemidae				
<u>Epiphanis cornutus</u> Esch.	1			
Helodidae				
<u>Elodes angusta</u> Hatch	1			
<u>Elodes</u> sp.			1	
Histeridae				
<u>Isolomalus mancus</u> Csy.	2			
Hydrophilidae				
<u>Crenitis snoqualmie</u> Mil.		1		
<u>Megasternum posticatum</u> Mann.	7	7	12	3
Lampyridae				
<u>Ellychnia hatchi</u> Fend.	1			
Lathridiidae				
<u>Melanophthalma distinguenda</u> C.				
<u>M. pumila</u> LeC.			1	1
<u>Stephostethus liratus</u> LeC.	1			
Leiodidae				
<u>Agathidium maculosum</u> Brown			1	

APPENDIX 3. (Continued)

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Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Coleoptera				
Leiodidae				
<u>Agathidium pulchrum</u> Lec.			1	
<u>Agathidium</u> sp.			2	
<u>Catops basilaris</u> Csy.			2	
<u>Hydnobius longulus</u> Lec.			1	
<u>Leiodes curvata</u> Mann.				1
<u>Platycholeus opacellus</u> Fall	2			
Limnichidae				
<u>Limnichus tenuicornis</u> Csy.	1			
Lucanidae				
<u>Platyceroides laticollis</u> Csy.				1
Melandryidae				
<u>Prothalia holmbergii</u> Mann.	1			
<u>Xylita livida</u> Sahlb.		2		
Meloidea				
<u>Lytta stygica</u> Lec.				5
Melyridae				
<u>Amecocerus provincialis</u> Blais				3
<u>Amecocerus</u> sp.				2
<u>Anthocomus mixtus</u> Horn				1
<u>Dasyrhadus impressicollis</u> Fall			1	
<u>Dasytes cruralis</u> Lec.				2
<u>Eurelymis atra</u> Csy.	1	3		24
<u>Hoppingiana hudsonica</u> Lec.		1		6
<u>Hypebaeus bicolor</u> Lec.				1
Unknown			1	
Mordellidae				
<u>Mordella atrata</u> Melsh.				3

APPENDIX 3. (Continued)

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Coleoptera				
Nitidulidae				
<u>Epuraea aestiva</u> L.	2		8	
<u>E. ambigua</u> Mann.		1		
<u>E. avara</u> Rand.		36	6	
<u>Meligethes nigrescens</u> Steph.				2
<u>Omosita discoidea</u> F.	1		4	
<u>Pocadius fulvipennis</u> Er.		1		1
Oedemeridae				
<u>Asclera nigra</u> LeC.		1		
Ostomidae				
<u>Eronyxa pallidus</u> Mots.		2		20
<u>Nemozoma punctatum</u> Van D.		1		
<u>Temnochila virescens</u> F.		2		
Ptilidae				
<u>Acratrichis</u> sp.			13	1
Scarabaeidae				
<u>Aphodius haemorrhoidalis</u> L.				1
<u>A. opacus</u> LeC.	1			
<u>A. pectoralis</u> LeC.			1	2
<u>Aphodius</u> sp.				2
<u>Dichelonyx valida</u> LeC.	1			
Scolytidae				
<u>Carphoborus vandykei</u> Bruck.		1		
<u>Cryphalus</u> sp.		1		
<u>Dendroctonus pseudotsugae</u> Hopk.	2	2	1	
<u>Dolurgus pumilus</u> Mann.		1		
<u>Gnathotrichus retusus</u> LeC.			1	
<u>Hylastes longicollis</u> Sw.		1		
<u>H. nigrinus</u> Mann.		3	2	
<u>Ips latidens</u> LeC.		7		

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Coleoptera				
Scolytidae				
<u>Leperisinus californicus</u> Sw.		1	1	
<u>Phloesinus</u> sp.			1	
<u>Pityophthorus</u> sp.	1	2		
<u>Pseudohylesinus granulatus</u>	58	23	46	
<u>P. nebulosus</u> LeC.		2		
<u>P. nobilis</u> Sw.		9		1
<u>Scolytus tsugae</u> Sw.	3	19		
<u>S. unispinosus</u> LeC.	4	7	3	
<u>Trypodendron lineatum</u> Ol.				
Scaptiidae				
<u>Anaspis</u> sp.	32	54	19	16
Sphaeritidae				
<u>Sphaerites politus</u> Mann.	1		1	
Staphylinidae				
<u>Amphicroum maculatum</u> Horn		4	2	1
<u>Anthobium fimetarium</u> Mann.			2	1
<u>A. subcostatum</u> Makl.	5		6	
<u>Coprophilus sexualis</u> Leech			1	
<u>Eusphalerum ferrariae</u> Hatch			1	
<u>E. minskae</u> Hatch	2	3		1
<u>Hapalaraea floralis</u> Payk.	5	2	1	2
<u>Ischnopoda</u> sp.			1	1
<u>Lordithon pygmaeus</u> F.				1
<u>Lordithon</u> sp.				1
<u>Medon</u> sp.				
<u>Megarthus pictus</u> Mots.			2	2
<u>Pelecomalium opaculum</u> Csy.	4		5	1
<u>P. puberulum</u> Fauv.		1		5
<u>P. testaceum</u> Mann.	8	7	3	

APPENDIX 3. (Continued)

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Coleoptera				
Staphylinidae				
<i>Philonthus concinnus</i> Grav.		2		2
<i>P. cruentatus</i> Gmel.		1	6	
<i>P. picipornis</i> Horn			1	
<i>Philonthus</i> sp.			1	
<i>Phaeopterus lagrandeuri</i> Hatch	1			
<i>Platystethus americanus</i> Er.				2
<i>Proteinus</i> sp.	1			
<i>Quedius aenescens</i> Makl.	1	2	10	2
<i>Q. capucinus</i> Grav.			1	
<i>Q. laevigatus</i> Gyll.		1		
<i>Q. marginalis</i> Makl.	1			
<i>Q. oculus</i> Csy.	1			
<i>Quedius</i> sp.				
<i>Stenus maritimus</i> Mots.		3	2	
<i>Tachinus contortus</i> Hatch	1		3	
<i>T. semirufus</i> Horn	1			
<i>Tachyporus chrysomelinus</i> L.				1
<i>Trigonurus dilaticollis</i> Van D.		1		1
<i>Xestolinus frontalis</i> Hatch		1	1	1
Unknown				
Tenebrionidae				
<i>Phthora americanum</i> Horn	1			
Trixagidae				
<i>Aulonothroscus validus</i> LeC.	1			
Unknown family	2i, la	1	4	1
Collembola ^c				10
Diptera				
Acarthophthalmidae	2		1	

APPENDIX 3. (Continued)

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Species	Plot ^b			
	1	2	3	4
	Young growth	Midgrowth	Old growth	Clearcut
Diptera				
Acroceridae				
Eulonchus sp.	6	2		2
Agromyzidae	54	11	3	77
Anisopodidae				
Mycetobia sp.	1			
Unknown		1		1
Anthomyiidae	129	303	10	67
Asilidae	16	1		1
Aulacigastridae				
Aulacigaster leucopeza Meigen	2			
Bibionidae				
Bibio sp.				2
Unknown		1		1
Bombyliidae				
Anthrax sp.				3
Villa sp.				2
Unknown				41
Calliphoridae	29	9	3	28
Cecidomyiidae	886	485	655	483
Ceratopogonidae	40	39	67	195
Chamaemyiidae				
Leucopsis sp.	2			
Unknown	4	2	1	171
Chironomidae	638	137	291	1,780
Chloropidae				
Thaumatomyia sp.	18	1	1	10
Unknown		4		143
Conopidae				1
Culicidae	19	1	4	

APPENDIX 3. (Continued)

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Diptera				
Dolichopodidae	17	1	2	3
Drosophilidae				
Amiota sp.		1		2
<u>Scaptomyza</u> sp.	8	1		2
Unknown	1,535	216	102	958
Empididae				
Ephydriidae				
<u>Ditricophora argyrostoma</u> C.				1
<u>Ditricophora</u> sp.	5			22
<u>Hydrellia griseola</u> (Fallen)	1			
<u>Hydrellia</u> sp.	10			
<u>Parydra</u> sp.	4	1		
<u>Psilopa compta</u> (Meigen)				1
Unknown	7		1	14
Heleomyzidae				
<u>Amoebaleria infuscata</u> Gill	1			
<u>Borboropsis steyskali</u> Mathis	12	2	1	7
<u>Suillia</u> sp.	1	2		
Unknown	3			
Lauxaniidae				
<u>Minettia lupulina</u> (Fab.)	1			
<u>Minettia</u> sp.		2		
Unknown	5			
Lonchopteridae				6
Lonchaeidae	111	3	12	32
Milichiidae	27	22	3	913
Muscidae				
<u>Schoenomyza</u> sp.				2
Unknown	178	24	15	102

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Diptera				
Mycetophilidae	1,685	766	580	119
Odiniidae	3			
Otitidae	10	7	2	1
Pallopteridae				
Palloptera sp.	5			
Unknown	9			
Periscelididae				
Periscelis sp.	2			
Phoridae	735	184	39	745
Piophilidae	4	5		1
Pipunculidae	15		1	10
Platypezidae	4			1
Psilidae	2			
Psychodidae	99	18	127	17
Rhagionidae				
Symphoromyia sp.	222	3	10	25
Unknown	138	7	3	6
Sarcophagidae	14	1	7	28
Scatopsidae		1	1	13
Scenopinidae				4
Sciaridae	466	136	201	368
Sciomyzidae				
Limnia sp.	1	1		
Unknown				
Sepsidae				
Sepsis sp.	1	3		11
Unknown	1		1	7
Simuliidae	104	60	182	250
Sphaeroceridae				
Leptocera sp.		6		

APPENDIX 3. (Continued)

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Diptera				
Sphaeroceridae				
Scatophora sp.		2		
Sphaerocera sp.		6		
Unknown	55	12	5	44
Stratiomyidae				
Syrphidae	2			
Tabanidae	108	69	47	2i, 174a
Chrysops sp.	37		1	11
Hybomitra sp.	18	10	7	7
Tabanus sp.	12	1	1	3
Unknown	14		4	4
Tachinidae	231	18	15	138
Tephritidae	7			2
Therevidae		1		4
Tipulidae	18	6	13	14
Trichoceridae	5		4	
Trixoscelididae	2	1		5
Unknown family	50	29	33	135
Ephemeroptera	1		1	
Hemiptera				
Anthocoridae				6
Aradidae	1	2	2	2
Berytidae	1			3
Cydnidae				
Annestrus sp.				1
Lygaeidae				
Geocoris sp.				11
Scolopostethus sp.	7			5
Unknown	5	4		12

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Hemiptera				
Miridae				
<u>Lygus</u> sp.	1			1
Unknown	11, 4a	3	1	8
Pentatomidae				
<u>Cosmopepla</u> sp.				1
<u>Peribalus</u> sp.				1
Unknown	1			
Tingidae				
<u>Corythuca</u> sp.	2			12
<u>Derephysia</u> sp.				2
Unknown family		1i	1	1
Homoptera				
Achilidae	1		3	
Adelgidae ^c				
<u>Adelges cooleyi</u> (Gillette)	10	12	34	387
Aleyrodidae		1		3
Aphididae ^c	173	303	160	1,046
Cicadellidae				
<u>Cuerna hasbrouki</u> Nielson	28	17	2	6
Unknown				42
Delphacidae			1	8
Psyllidae	28		8	108
Unknown family	11, 2a	1i		1
Hymenoptera				
Andrenidae	11	2		81
Anthophoridae				
<u>Nomada</u> sp.	1			2
Unknown	1			2
Apidae				
<u>Apis mellifera</u> L.	3	1		84

APPENDIX 3. (Continued)

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Hymenoptera				
Apidae				
<u>Bombus bifarius</u> Cresson		1		1
<u>B. occidentalis</u> Greene				1
<u>B. vosnesenskii</u> Rad.				1
<u>Bombus</u> sp.	35	11	8	16
Unknown		1		
Argidae				3
Aulacidae				4
Braconidae	2	26	20	286
Chalcididae	109			3
Chrysididae	4		1	8
Cimbicidae				
<u>Zaraea americana</u> Cresson			1	
Colletidae	3			13
Cynipidae	3	2		4
Diapriidae	7	6	7	5
Eucharitidae	1	1		17
Formicidae	3	3	2	70
Gasteruptiidae			9	
Halictidae	67	21	9	95
Ichneumonidae	414	52	59	78
Megachilidae				
<u>Anthidium</u> sp.				2
<u>Heriades</u> sp.				1
Unknown				
Misc. Chalcidoidea and Proctotrupoidea	2	1	1	27
Orussidae	20	27	12	588
Platygasteridae		1		1
<u>Inostemma</u> sp.				
Pompilidae	2	1	1	2

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Hymenoptera				
Sphecidae	27	3	6	15
Tenthredinidae	110	3	10	7
Vespidae	50	26	14	8
Lepidoptera				
Aegeriidae				18
Hesperiidae				1
Lycaenidae				3
Noctuidae				4
Misc. microlepidoptera	27	8	12	37
Nymphalidae				
Boloria sp.				1
Nymphalis californica (Bois.)		4		15
Unknown			1	18
Unknown family	2	1i, 1a		15
Neuroptera				
Chrysopidae		1	1i	
Coniopterygidae	2	3	1	
Hemerobiidae	2			
Inocelliidae				
Inocellia sp.		4		2
Raphidiidae				
Agulla sp.				3
Sialidae	1			
Pleocoptera				
Capniidae				1
Capnia sp.				
Eucapnopsis sp.	1			
Chloroperlidae				
Alloperla sp.	1		1	

APPENDIX 3. (Continued)

Species	Plot ^b			
	1 Young growth	2 Midgrowth	3 Old growth	4 Clearcut
Pleocoptera				
Leuctridae				
Leuctra sp.	2			9
Nemouridae				
Nemoura sp.	2			6
Psocoptera ^c	22		3	2
Strepsiptera				
Stylopidae		19	1	1
Thysanoptera ^c				613
Trichoptera			1	
Rhyacophililidae		2		1
Unknown	2			
Unknown Order	11		6i	

^a A net was run for 2 consecutive hours once a week each in Plots 2 and 3 and run for 2 consecutive hours each on two days weekly in Plots 1 and 4.

^b "i" indicates that the immature stage was collected; all other unmarked numbers represent the adult stage captured. Sometimes "a" is used to indicate adult stage to avoid confusion.

^c No attempt was made with these groups to distinguish immature from adult stages, and all were recorded as adult.